

In this case histology of the biopsy revealed fungal hyphae with very few septa, (resembling those shown in Figure 82.2), and the culture grew out *Rhizopus* spp., confirming the diagnosis of mucormycosis. This child was treated with amphotericin B, and the lesion healed. In this case the infection was probably caused by contaminated elastoplast used in the dressing on the elbow.

Mucormycosis is an infection affecting primarily immunocompromised individuals, such as those with neutropenia, those posttransplant, and those with diabetic keto-acidosis. The Mucorales cause cutaneous infections, pneumonia, and infections of the paranasal sinuses. Infection of the paranasal sinuses can spread rapidly to involve the orbit and brain. A clue to the presence of this organism in the sinus is black discoloration around the nose. Mucormycosis is rapidly progressive and destructive. Therefore making a diagnosis in a suspected case and initiating appropriate therapy is an emergency. Making a diagnosis requires performing a biopsy of tissue, such as sinus mucosa, for histology and for culture for fungi and bacteria. Surface cultures are not appropriate. Histology is more sensitive than culture.

Therapy consists of the following three elements: (i) correcting the underlying defect to the extent possible, (ii) surgical debridement,

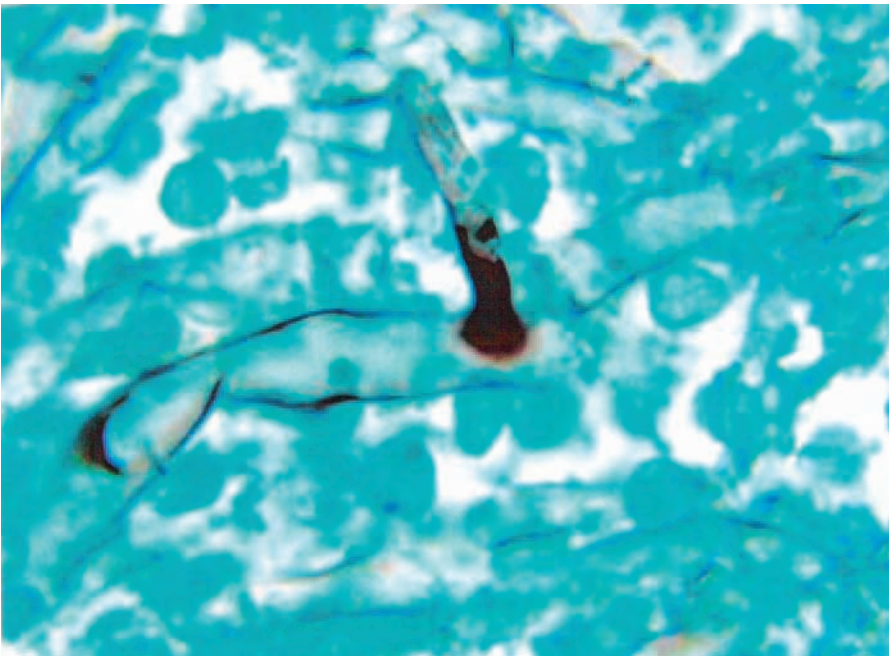


FIG. 82.2. *The histological appearance of a zygomycete in tissue. (Courtesy of the Centers for Disease Control and Prevention/Dr Libero Ajello)*

(iii) antifungal therapy. Antifungal therapy should consist of amphotericin B or one of its lipid-based derivatives. Of the antifungal azoles, only posaconazole is active against the zygomycetes.

Reading:

Greenberg RN, Scott LJ, Vaughn HH et al: Zygomycosis (mucormycosis): emerging clinical importance and new treatments. *Curr Opin Infect Dis* 2004; 17: 517–525.

Gonzalez CE, Rinaldi MG, Sugar AM: Zygomycosis. *Infect Dis Clin N Am* 2002; 16: 895–914.

CASE 83. A 15-month-old girl presents with a 1-day history of fever and vomiting. There is no diarrhea or cough, and no other members of the family are ill. There is no history of foreign travel, day-care attendance, or exposure to animals. On examination she is moderately ill-appearing, but alert and perfusing well. Her temperature is 39.1°C, heart rate 140/minute, and respiratory rate 30/minute. The heart, lungs, ears, throat, and limbs are normal, and there is no lymphadenopathy or neck stiffness. There is mild abdominal tenderness, particularly in the left upper quadrant and flank. However, no masses or enlargement of the liver or spleen can be detected.

- *What is your differential diagnosis?*
- *What would you do?*

This child has a febrile illness with localizing signs in the abdomen. Disease of several different organs can give rise to abdominal pain or tenderness, but those that cause pain in the left upper quadrant are the left lung, colon, spleen, and kidney. The high fever suggests an infection. The absence of diarrhea after 24 hours of symptoms makes enteric infection unlikely; the absence of respiratory symptoms and findings makes pneumonia unlikely; infection of the spleen itself is very rare, and infection in the subphrenic space would imply previous bowel perforation. The most likely cause of this child's symptoms and signs is pyelonephritis. Therefore the tests that are most likely to confirm the diagnosis are chemical analysis, microscopy, and culture of the urine.

The results of these tests showed the following: 2+ protein, 3+ leukocyte esterase, 3+ hemoglobin, leukocytes too numerous to count, 50 red cells per high power field, and many bacteria.

This is sufficient information on which to base a provisional diagnosis of a urinary tract infection, although a culture is the definitive diagnostic test.

On the following day the urine culture grew out *Escherichia coli*, >100,000 organisms per milliliter.

This confirms the diagnosis of a urinary tract infection. Although the urine findings do not determine the site of infection within the urinary tract, the symptoms of high fever, and the signs referable to the left kidney, indicate a diagnosis of pyelonephritis.

Urinary tract infections are usually caused by *Escherichia coli*. In children who have not previously been treated for urinary tract infection, other causative organisms are other enteric rods, in particular *Klebsiella* spp., and *Enterococcus faecalis*.

Making the diagnosis of a urinary tract infection in an infant or young child is often difficult for the following reasons:

- (i) there may be no focal symptoms or signs. When present, focal clinical features include urinary frequency, crying on urination, and suprapubic or abdominal pain or tenderness.
- (ii) obtaining a specimen of urine that is not contaminated with feces, smegma, or vulval flora requires the performance of a suprapubic bladder puncture (in infants only) or bladder catheterization. These require that there is urine in the bladder at the time of the procedure.

Diagnosing a urinary tract infection and treating the patient is important for the following reasons:

- (i) it can be complicated by bacteremia;
- (ii) it indicates the possibility of an underlying urinary tract malformation that has predisposed the patient to the infection, in particular vesicoureteral reflux;
- (iii) it can lead to renal scarring and, over a long period, to chronic renal damage.

Therefore in infants and young children with fever and no apparent site of infection, urinary tract infection should be considered. Because of the above implications of diagnosing this condition, an appropriate specimen of urine must be obtained. If such an infection is diagnosed, further investigation, looking for evidence of a urinary tract abnormality should be performed. This usually consists of a renal ultrasound and a voiding cystourethrogram.

Management consists of antimicrobial therapy directed at the most likely pathogen, namely *Escherichia coli*. In the past ampicillin was active against most strains of this organism. However, this is no longer the case. Therefore therapy should consist of trimethoprim/sulfamethoxazole (to which there is also a rising rate of resistance) or a cephalosporin. Although parenterally

administered third-generation cephalosporins, such as ceftriaxone or cefotaxime are appropriate for ill children, they have not been shown to be superior to oral therapy with cefixime. If imaging reveals a urinary tract malformation that might predispose to urinary tract infection, such as vesicoureteral reflux, the child should receive long-term antimicrobial prophylaxis and be referred to a urologist. Prophylaxis should consist of a drug that does not have significant effects on the bowel flora, for example, trimethoprim/sulfamethoxazole, at about one-fourth the usual therapeutic dosage. Whether such prophylaxis is clearly of benefit is currently being questioned.

Reading:

American Academy of Pediatrics, Committee on quality improvement: practice parameter: the diagnosis, treatment, and evaluation of the initial urinary tract infection in febrile infants and young children. *Pediatrics* 1999; 103: 843–852.

Hoberman A, Wald ER, Hickey RW et al: Oral versus intravenous therapy for urinary tract infections in young febrile children. *Pediatrics* 1999; 104: 79–86.

Wald ER: Vesicoureteral reflux: The role of antibiotic prophylaxis. *Pediatrics* 2006; 117: 919–922.

CASE 84. A 2-year-old girl presents with a history of cough and fever for 2 days.

On examination she has a temperature of 39°C, a respiratory rate of 50/minute, dullness to percussion, and bronchial breathing (tubular breath sounds) over the right upper lobe anteriorly. The rest of her examination is normal.

- *What is the likely diagnosis?*
- *How would you manage her?*

In a child with fever, cough, and tachypnea the most likely diagnosis is pneumonia. The additional findings of dullness to percussion and bronchial breath sounds indicate pulmonary consolidation. Dullness with decreased breath sounds could indicate consolidation and/or the presence of pleural fluid. However, pleural fluid, being dependent, is usually detected over the lower lobes. Pneumonia associated with pulmonary consolidation indicates lobar pneumonia. In such a patient the most likely causative agent is

Streptococcus pneumoniae, although *Haemophilus influenzae* (type b or non-typeable), *Staphylococcus aureus*, *Mycoplasma pneumoniae*, or respiratory viruses are also possible causes. An attempt to make a microbiological diagnosis should be made by blood culture. Sputum cannot be obtained from young children, but in those older than about 6 years, an attempt should be made to obtain sputum for Gram stain and culture. Detecting respiratory viral antigens or culturing these viruses from nasopharyngeal secretions can be performed. However, they are useful only for isolation procedures for hospitalized patients and for withholding antibiotics in those patients with positive results.

A microbiological diagnosis is important in immunocompromised children in whom the diagnostic possibilities are broad. In such cases invasive procedures such as lung puncture, bronchoalveolar lavage, or lung biopsy may be indicated.

Although a chest X ray may not be essential for initial management, it is of use for the following reasons:

- (a) as a baseline, in case the patient does not improve as expected;
- (b) to evaluate for complications;
- (c) to determine whether there is an underlying lung problem predisposing the child to pneumonia. Radiological evidence of consolidation should have cleared within 6–8 weeks. Failure of this to occur should alert one to the possibility of an underlying abnormality, such as bronchial obstruction.

The chest X ray of this child is shown in Figure 84.1.

Management consists of supportive care, ensuring that the child is adequately oxygenated and hydrated, and antimicrobial therapy. Initial antimicrobial therapy should consist of a penicillin, for example, amoxicillin or a third-generation cephalosporin.

Pneumonia means inflammation of the alveoli. This is usually caused by an infectious agent, but it may also be caused by other noxious agents, such as toxins, for example kerosene. Pathologically pneumonia may be patchy (bronchopneumonia) or be localized to a segment or lobe of the lung (lobar pneumonia). These may have different clinical features (see below). The differential diagnosis of pneumonia and associated physical findings are listed in the Table 84.1. The main microbial causes of pneumonia in children of different ages and in those with different risk factors are shown in Table 84.2.

Pathogenesis of pneumonia: Microorganisms can reach the alveoli by descending from the upper airway (aerogenous), the usual route, or hematogenously. In aerogenous pneumonia, there may be descent of organisms

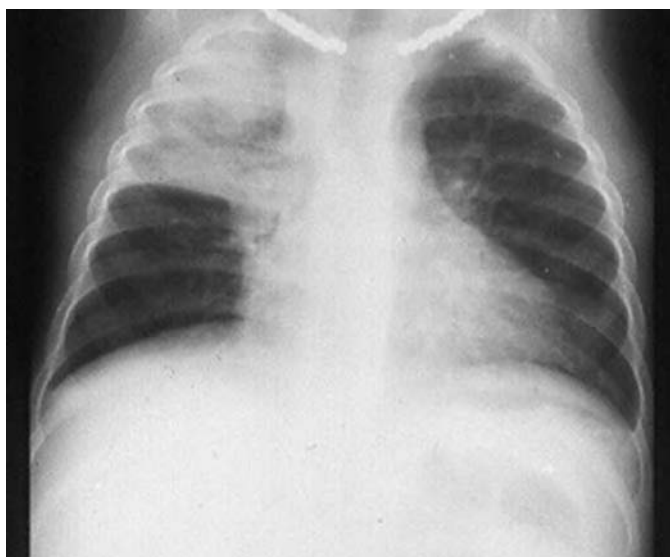


FIG. 84.1. *Chest X ray showing right upper lobe consolidation.*

into many parts of the lung or spread within a lobe through the pores of Kohn.

Clinical features: Fever and tachypnea are the hallmark features of pneumonia in infants and young children. Tachypnea is the most important physical finding in pulmonary disease because it is more sensitive than other findings, such as auscultatory findings. In addition, the respiratory rate is an objective physical finding. Crackles are often present in both patchy and lobar pneumonia, but signs of consolidation are present in only lobar pneumonia. There may be additional signs of respiratory distress or failure, or of the sepsis syndrome. These include intercostal and subcostal retractions, use of accessory muscles of respiration, cyanosis, depressed level of consciousness, agitation, poor perfusion, and hypotension. Lobar pneumonia may be associated with the symptom of localized chest pain and/or tenderness. In the case of pneumonia abutting on the diaphragm, there may be very severe abdominal pain, suggesting the possibility of an acute abdominal emergency. In children with upper lobe pneumonia neck stiffness may be present, suggesting the presence of meningitis. The signs of lobar consolidation include the following: dullness to percussion, decreased intensity but a higher pitch of the breath sounds (“bronchial” or “tubular” breath sounds), “E- to-A changes,” crackles, and increased vocal resonance over the involved area.

Complications of pneumonia may be present at the time the patient presents or may develop during the course of treatment. These can be considered in terms of local spread and distant spread of the infection and of complications specific to the causative organism.

TAB 84.1: The differential diagnosis of pneumonia and associated findings.

Bronchiolitis	fever, tachypnea, crackles, wheeze, hyperinflation
Pulmonary infarction	fever, focal chest pain, tachypnea, consolidation ^a
Acute chest syndrome (sickle cell disease)	fever, focal chest pain, tachypnea, consolidation ^a
Pulmonary edema	tachypnea, crackles
Atelectasis	tachypnea, decreased breath sounds, consolidation ^a
Bronchial obstruction (total)	see atelectasis, mediastinal shift toward the side of obstruction
Bronchial obstruction (partial)	tachypnea, decreased breath sounds, hyperinflation of affected side, mediastinal shift away from the side of obstruction
Pleural fluid	dullness, decreased breath sounds, shift away from the side of the fluid
Pneumothorax	resonance, decreased breath sounds, shift away from the side of the pneumothorax
Tumor	decreased breath sounds, dullness

^aSigns of consolidation: dullness to percussion, bronchial breathing (“tubular” breath sounds), and decreased breath sounds.

1. Local spread:

- (a) *Pleural effusion*: This may occur with bacterial or nonbacterial pneumonia, adjacent to the site of pneumonia. It is termed a “parapneumonic effusion.” However, in the case of bacterial pneumonia, infection may spread to the pleural space, leading to a pleural empyema. This is associated with persistent fever in patients whom one would have expected to improve with therapy. Infection-related pleural effusions can develop in the absence of pneumonia as a result of infradiaphragmatic infection such as a subphrenic abscess, liver abscess, or perforated esophagus. A pericardial effusion can also complicate pneumonia.
- (b) *Lung abscess*: This results from necrosis of lung tissue caused by bacterial pneumonia, especially in cases caused by *Staphylococcus aureus* or anaerobic bacteria (Figure 84.2).

2. Hematogenous spread of infection can occur with any bacterial infection. This is a particular problem in cases of bacteremia caused by

TAB 84.2: Causes of pneumonia in children, according to age and risk factors.

Infectious causes

Normal host, community-acquired

Newborns

Streptococcus agalactiae (group B)*Escherichia coli* and other enteric Gram-negative bacilli*Listeria monocytogenes*

Herpes simplex virus

*Chlamydia trachomatis*The role of *Mycoplasma hominis* and *Ureaplasma urealyticum* in neonatal pneumonia is controversial

Infants and toddlers

Respiratory syncytial virus

Other respiratory viruses

Adenovirus

Parainfluenza virus

Influenza virus

Metapneumovirus

*Streptococcus pneumoniae**Haemophilus influenzae**Staphylococcus aureus*

School aged children and teenagers

*Mycoplasma pneumoniae**Chlamydia pneumoniae**Streptococcus pneumoniae*

Organisms requiring specific exposures that can often be determined by history

Measles

Coxiella burnetii (Q fever)*Chlamydia psittaci*

Hospital-acquired

Respiratory syncytial virus

Other respiratory viruses

Enteric Gram-negative bacilli

*Pseudomonas aeruginosa**Staphylococcus aureus*

Immunocompromised patients:(neutropenia, cancer chemotherapy, transplantation, HIV infection)

All of the above PLUS:

Pneumocystis jiroveci (formerly *P. carinii*)

Cytomegalovirus

Fungi, especially *Aspergillus**Nocardia* species*Rhodococcus equi*

Cystic fibrosis

*Staphylococcus aureus**Haemophilus influenzae**Pseudomonas aeruginosa**Burkholderia cepacia**Stenotrophomonas maltophilia*

Nontuberculous mycobacteria

(Continued)

(Continued)

Subacute and chronic

Mycobacterium tuberculosis

Endemic mycoses

Histoplasma capsulatum

Blastomyces dermatitidis

Coccidioides immitis

Pulmonary infiltrates + eosinophilia

Visceral larva migrans – *Ascaris lumbricoides*, *Toxocara canis*, *T. cati*

Noninfectious causes of pulmonary infiltrates

Food/milk aspiration

Hypersensitivity pneumonia e.g. bird fancier disease

Toxic e.g. kerosene ingestion, paraquat ingestion, chlorine inhalation, cancer chemotherapy

Vasculitis e.g. Goodpasture's syndrome, Wegener's granulomatosis, systemic lupus erythematosus

Pulmonary infarction (thrombus/embolus)

Pulmonary contusion (trauma)

Acute chest syndrome (sickle cell disease)

Heart failure (pulmonary edema)

Malignancy

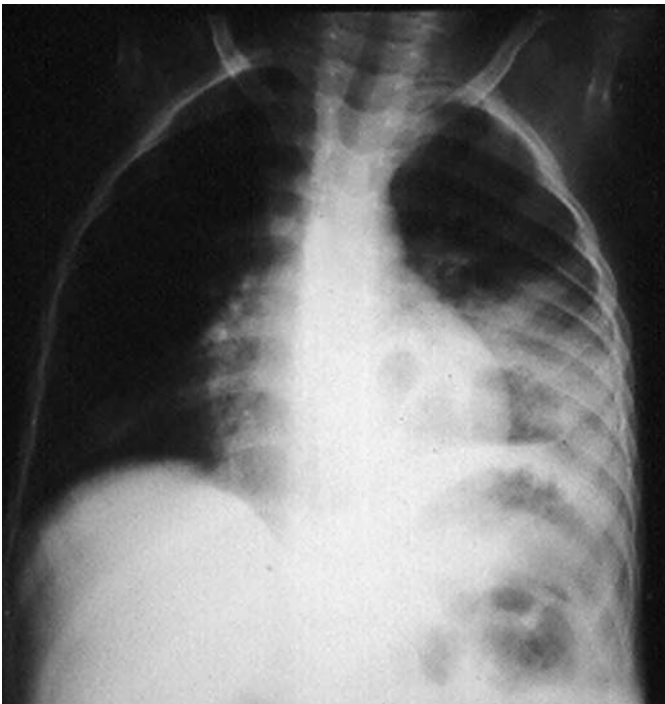


FIG. 84.2. Chest X ray of a child showing left lower lobe consolidation with cavitation due to an abscess.

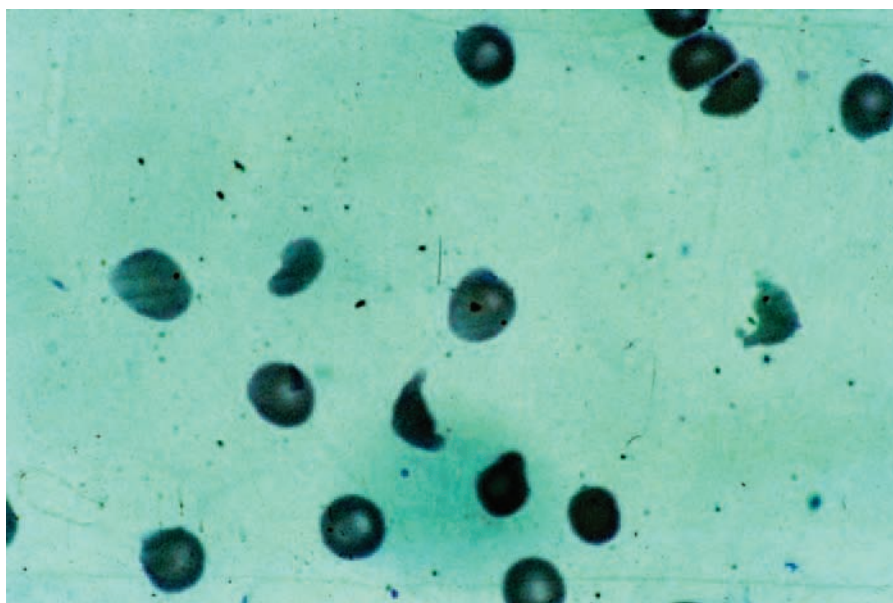


FIG. 84.3. A blood smear from a child with hemolytic uremic syndrome complicating pneumococcal pneumonia. Note the wide spaces between the cells, indicating severe anemia (the hemoglobin concentration was 1.8 g/dl), and fragmented red cells.

Staphylococcus aureus and *Fusobacterium necrophorum* (Lemierre syndrome).

3. Complications related to specific organisms:

- (a) *Streptococcus pneumoniae*: This organism may cause the hemolytic uremic syndrome. The mechanism, which is different from that in cases caused by Shiga-like toxin of enterohemorrhagic *E. coli*, is related to bacterial neuraminidase injuring cell membranes of host cells (Figure 84.3).
- (b) *Mycoplasma pneumoniae* can cause a cold antibody autoimmune hemolytic anemia, as well as erythema multiforme and encephalitis.

Management of complications of pneumonia

Empyema: When this is suspected, a chest radiograph should be taken, preferably in the erect and lateral decubitus positions, in order to determine whether there is fluid in the pleural space and whether it flows or is loculated. If fluid is present, it should be aspirated for the purpose of confirming the diagnosis and with the hope of obtaining a culture of the causative organism. (This is seldom accomplished if the patient has already received antimicrobial therapy.) Ultrasound can also be helpful in localizing the pleural fluid. If empyema is present, the treatment is surgical, either by closed

chest tube drainage or by thoracoscopy-directed debridement. This latter procedure is associated with a shorter hospital course.

Lung abscess: Treatment in this case entails antimicrobial therapy for several weeks. Surgical management is seldom necessary.

Reading:

Bradley JS: Old and new antibiotics for pediatric pneumonia. *Semin Respir Infect* 2002; 17: 57–64.

CASE 85. An 18-month-old girl presents with a history of fever for 4 days and bilious vomiting for 1 day. She has not defecated for 3 days. She was seen in an emergency room 1 day ago and was treated with ceftriaxone. On examination she is moderately ill-appearing, but alert and perfusing well. The examination is normal except for the abdominal examination which reveals mild distension and marked generalized tenderness, particularly marked on the right side, especially inferiorly.

- ***What is the differential diagnosis?***
- ***How would you manage the child?***

This child has three main clinical features: fever, vomiting, and marked abdominal tenderness. This suggests an infectious process within the abdominal cavity. Although acute lobar pneumonia can cause abdominal pain, tenderness, and fever, tenderness in that case is mainly upper abdominal. The absence of respiratory symptoms and signs are also against the diagnosis of pneumonia. Organs within the abdominal cavity that are potential foci of disease include the kidneys, the liver, the gallbladder, the intestine, and the pancreas. The absence of diarrhea is against the diagnosis of acute gastroenteritis, but the infections related to the intestine that can present in this fashion are those associated with inflammation of the subepithelial layers, such as typhoid fever or acute appendicitis, especially if perforation has occurred. The most likely diagnosis is a perforated appendix with abscess formation. The bile-stained vomitus suggests the possibility of a high bowel obstruction as might occur with a midgut volvulus, associated with intestinal malrotation. Although this is the most serious consideration, a patient with this condition is likely to be very ill-appearing due to bowel ischemia and systemic acidosis.

Management should entail the following:

- (a) ensuring adequate perfusion and hydration;
- (b) nasogastric drainage;

- (c) laboratory tests: blood culture; electrolyte concentrations; serum amylase and lipase; urinalysis and urine culture;
- (d) antimicrobial therapy directed against enteric rods and anaerobes should be initiated, such as a combination of a third-generation cephalosporin and metronidazole.
- (e) imaging studies, which should be done in the following sequence:
 - (i) plain abdominal X ray with the X ray beam horizontal, to look for free gas. If this is seen the child should undergo laparotomy immediately.
 - (ii) contrast study of the upper gastrointestinal tract to look for malrotation, which is the predisposing factor to midgut volvulus. If this is abnormal, the child should undergo laparotomy immediately.
 - (iii) computer tomography (CT) scan of the abdomen and pelvis with intravenous and intrabowel contrast to look for foci of intra-abdominal infection. This might need to be delayed if the contrast used in the earlier upper gastrointestinal study interferes with the imaging.

In this case the plain abdominal X ray and upper gastrointestinal contrast study were unrevealing. After 2 days, during which the child received antimicrobial therapy as described above, and during which her condition

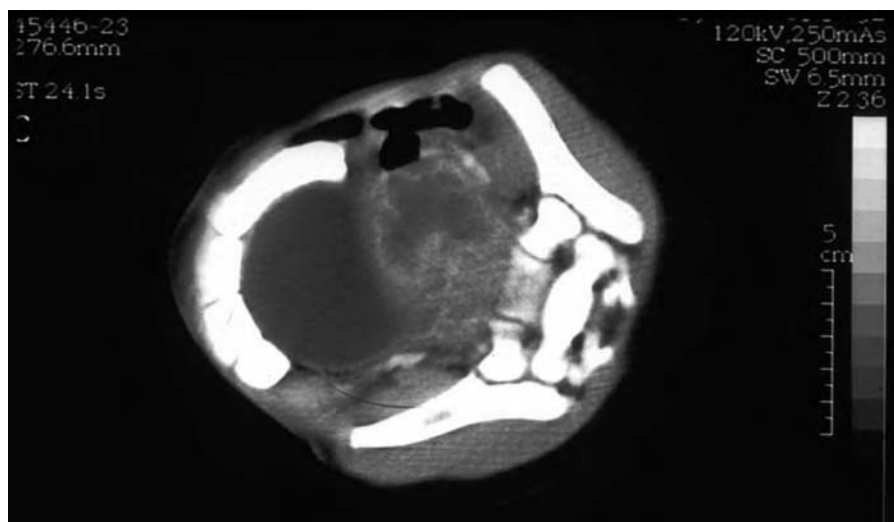


FIG. 85.1. CT scan showing a large pelvic abscess resulting from a perforated appendix.

improved, a CT scan was performed, revealing a pelvic abscess probably arising from a perforated appendix (Figure 85.1).

She was treated with antimicrobial therapy for 10 days and made an uneventful recovery. The diseased appendix was removed laparoscopically 4 weeks later.

Reading:

Solemkin JS, Mazuski JF, Baron EJ et al: Guidelines for the selection of anti-infective agents for complicated intra-abdominal infections. Clin Infect Dis 2003; 37: 999–1005.

CASE 86 (COMP). An 11-year-old boy presents with fever and severe headache, followed by confusion. He had a “cold” about 2 weeks ago. On examination he is febrile, drowsy, and has mild swelling and erythema of the right upper eyelid. The eyes have normal movements, but there is papilledema on the right. There is some neck stiffness and slight weakness of the left upper and lower limbs. During the examination he suffers a seizure, which is controlled with intravenous clonazepam.

- *What is the differential diagnosis?*
- *What would you do?*

The clinical features of fever, headache, and changes in mental state strongly suggest an infection affecting the brain. The swelling and erythema around the right eye suggest bacterial sinusitis, and the papilledema indicates raised intracranial pressure. This whole clinical picture indicates that bacterial sinusitis has been complicated by intracranial suppuration such as a subdural or epidural empyema, a brain abscess, and/or meningitis. The intracranial empyemas in this circumstance can be present over the convexity of the brain or within the interhemispheric fissure, while abscesses affect the frontal lobes. Had external ophthalmoplegia been present, this would have suggested the additional possibility of orbital cellulitis, while the presence of internal and external ophthalmoplegia and chemosis would have suggested the complication of cavernous sinus thrombosis.

Management should consist of the following:

- (a) *Supportive care:* Controlling the seizure, ensuring an adequate airway and breathing, and reducing the intracranial pressure;
- (b) Initiating antimicrobial therapy directed at the most likely pathogens, which include *Streptococcus pneumoniae*, other streptococci, in particular those belonging to the *Streptococcus milleri* group,



FIG. 86.1. CT of the patient's head showing a right epidural empyema and right hemispheric brain swelling with midline shift, complicating sinusitis.

Haemophilus influenzae, *Staphylococcus aureus*, anaerobes, and Gram-negative bacilli, including enteric bacilli. A combination of vancomycin, a third-generation cephalosporin and metronidazole, or a combination of vancomycin and meropenem is suitable.

- (c) *Confirming and clarifying the diagnosis:* This involves imaging the head by a computer tomography (CT) scan, with contrast, or by magnetic resonance imaging (MRI) (Figure 86.1); this shows a right hemisphere epidural empyema, with right hemisphere swelling and shift of the midline.
- (d) *Surgery:* The need for drainage of an abscess will depend on the actual findings on imaging. Specimens should be sent to the laboratory for Gram stain, and aerobic and anaerobic bacterial culture.

Similar complications can occur with middle ear infection. In this case the brain abscess affects the temporal lobe or cerebellum, and thrombosis affects the lateral sinuses.

This patient illustrates several very important points:

- (i) Eyelid swelling is an important feature of paranasal sinusitis; this is not an ocular problem.
- (ii) Severe headache or vomiting in individuals with sinusitis or otitis media suggests the possibility of intracranial complications of the infection. These include osteomyelitis of the skull, cerebral venous sinus thrombosis, epidural empyema, subdural, empyema, meningitis, and brain abscess.

Reading:

Piccirillo JF: Acute bacterial sinusitis. *N Engl J Med* 2004; 351: 902–910.

Bair-Merritt MH, Shah SS, Zaoutis TE et al: Suppurative intracranial complications of sinusitis in previously healthy children. *Pediatr Infect Dis J* 2005, 24: 384–386.

Subcommittee on Management of Sinusitis and Committee on Quality Improvement, American Academy of Pediatrics: Clinical practice guideline: management of sinusitis. *Pediatrics*. 2001; 108;798–808.

CASE 87 (COMP). A 6-year-old previously healthy boy presents to a hospital in South Africa with a history of fever and abdominal pain for 1 week, and extreme fatigue for 1 day. There is no diarrhea or hematochezia. On examination he is moderately ill-appearing, with a temperature of 39°C, heart rate of 120/minute, blood pressure of 100/50 mm Hg, good peripheral perfusion, mild jaundice, and marked mucosal pallor. The heart and lungs are normal. The abdominal examination reveals mild generalized tenderness, and the spleen is palpable 3 cm below the left costal margin. The rectal examination is normal, and the stool appears normal and tests negative for occult blood. There is no significant lymphadenopathy, and the neuromuscular examination is also normal. The urine is pink, and tests positive for hemoglobin.

- *What are the diagnostic possibilities?*
- *What should be done to differentiate between them?*

There are several abnormalities. Can they be linked in a single unifying diagnosis?

- (a) *Fever*: This is most likely caused by an infectious process, whether viral, bacterial, fungal, or parasitic.
- (b) *Abdominal tenderness*: This is not severe, tending to exclude generalized peritonitis, and diffuse, tending to exclude a focal process in

a site such as the liver or appendix. In young children, however, the lack of focality of abdominal tenderness may be misleading, both because they cannot vocalize their symptoms, and, in young infants, because the omentum is not large enough to seal perforations of the bowel. The organ that is present in every area of the abdominal cavity is the bowel. These symptoms may be compatible with bowel inflammation or distension.

- (c) *Jaundice*: This suggests either cholestasis, as might occur with hepatitis or biliary duct disease (unusual in previously healthy children), failure of conjugation, as occurs with hepatitis, or an excessive bilirubin load, as occurs with hemolysis.
- (d) *Pallor*: In the absence of shock this indicates anemia. The history suggests an acute anemia. This can be due to acute blood loss (for which there is no evidence), acute hemolysis, or acute bone marrow failure. Because there are abdominal symptoms, bleeding into the intestine should be considered. Acute anemia due to bone marrow failure caused by parvovirus B 19 (“aplastic crisis”) can occur in an individual with a shortened red cell lifespan due to a chronic hemolytic state such as sickle cell disease or hereditary spherocytosis, of which there is no history in this child.
- (e) *Pink urine*: This can be caused by certain foods, such as beetroots, but in the context of an ill patient should be considered likely due to blood, free hemoglobin, or myoglobin. All of these give a positive test for hemoglobin. If the urine is pink and turbid, this suggests hematuria, as occurs with disease of the urinary tract; if it is pink and clear (like rosé wine), this indicates hemoglobinuria or myoglobinuria. Hemoglobinuria is due to intravascular hemolysis, while myoglobinuria is due to rhabdomyolysis, of which there is no clinical evidence.

The acute anemia, jaundice, and pink urine suggest the likelihood of intravascular hemolysis. The few disorders that cause this include some autoimmune hemolytic anemias, which may complicate several different infections; macroangiopathic hemolytic anemia due to heart valve abnormalities (for which there is no evidence); microangiopathic hemolytic anemia, as occurs with hemolytic uremic syndrome, a possibility; and glucose-6-phosphate dehydrogenase (G6PD) deficiency, also a possibility. Hemolytic crises in individuals with G6PD deficiency are usually precipitated by infections but may be precipitated by oxidant drugs.

What infection might the child have? Of systemic viral infections infectious mononucleosis due to Epstein–Barr virus (EBV) might explain the

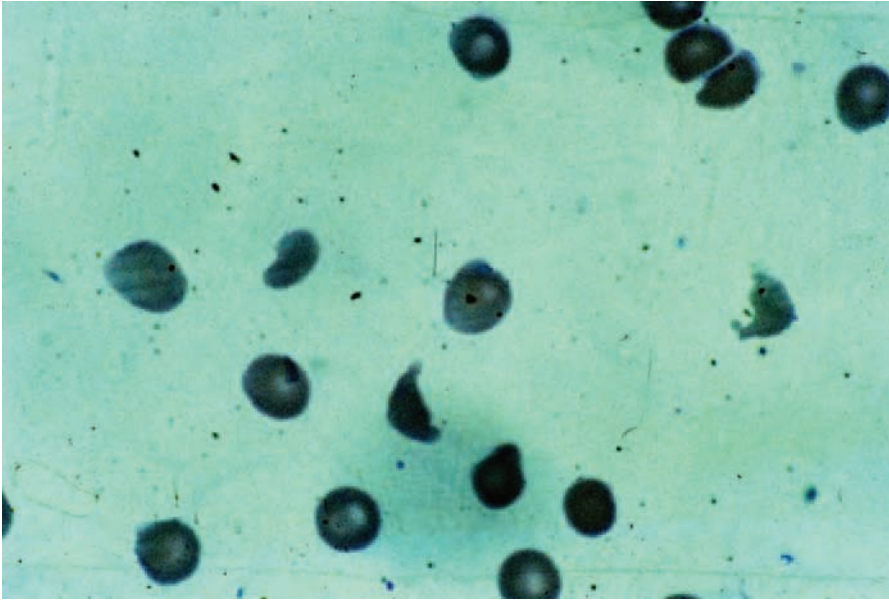


FIG. 87.1. *Schistocytes in the blood of a child with microangiopathic hemolytic anemia associated with hemolytic uremic syndrome. Note the large spaces between the red cells, indicating severe anemia. (The hemoglobin concentration was 1.8 g/dl.)*

anemia, jaundice, and splenomegaly best but not the abdominal tenderness, unless there is splenic rupture. If anemia were due to splenic rupture, there would likely be features of shock. The systemic bacterial infections that would best explain the abdominal pain, tenderness, and splenomegaly is typhoid fever (*Salmonella typhi* infection). Although malaria causes fever, hemolytic anemia, and splenomegaly, the hemolysis in this infection is generally not intravascular. A few tests might provide all the necessary information for a complete diagnosis of this child's problems:

hemoglobin concentration – to determine the severity of the anemia;

blood smear for red cell morphology and parasites – for schistocytes of angioathic hemolysis;

hemolysis (Figure 87.1), “bite” cells of G6PD deficiency (Figure 87.2), malaria parasites (see Case 63), and Downy cells of infectious mononucleosis (see Case 6);

Coomb's test – to diagnose autoimmune hemolysis;

urine dipstix and microscopy – to differentiate between causes of pink urine;

blood culture – to diagnose bacteremia, especially that caused by *Salmonella typhi*;

heterophile antibody test – to diagnose infectious mononucleosis.

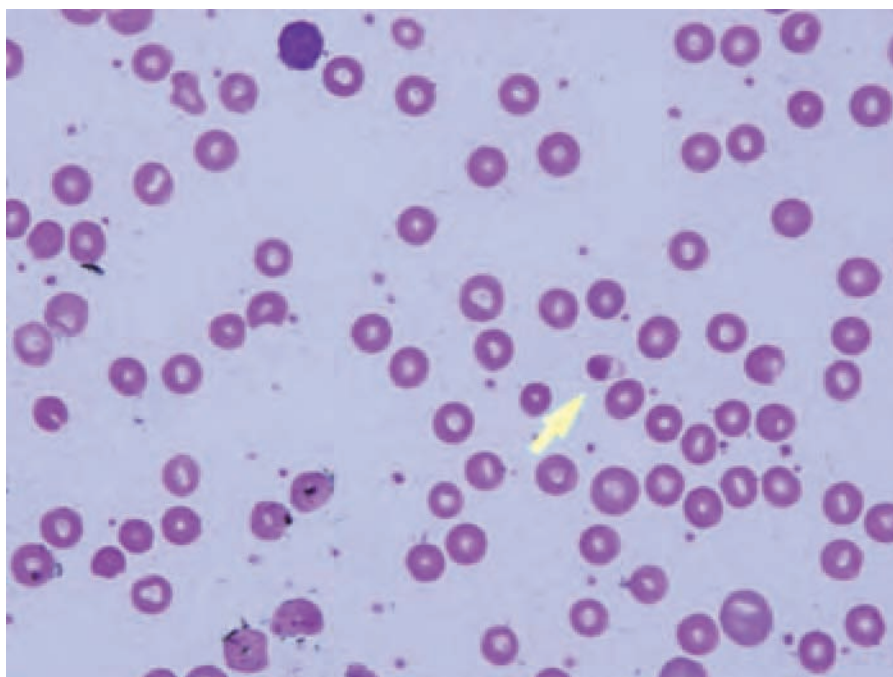


FIG. 87.2. *Blood smear of a child with hemolysis caused by glucose-6-phosphate dehydrogenase deficiency demonstrating a red cell with uneven distribution of hemoglobin within the cell ("bite cell"). (Courtesy of Dr Carlos Alvarado and Dr Carlos Abramowsky, Emory University)*

Although management will depend on the likely diagnosis or diagnoses, supportive care should be instituted immediately. The most important element of this is to ensure adequate hemoglobinization with a blood transfusion.

The diagnosis in this child was typhoid fever with underlying glucose-6-phosphate dehydrogenase deficiency.

Reading:

Berkowitz FE: Hemolysis and infection: categories and mechanisms of their interrelationship. *Rev Infect Dis* 1991; 13: 1151–1162.

CASE 88. An 18-month-old girl presents with fever and swelling on the right side of her neck of 3 days' duration. There is no history of living in or visiting another country, trauma, or upper respiratory tract symptoms. On examination she is mildly ill-appearing, with a temperature of 38.5°C. The right side of her neck has a swelling of approximately 3 cm diameter just below the angle of the mandible. It is firm and tender, and the overlying skin is erythematous. Movement of the neck is limited. The rest of the examination is normal.

- *What is the differential diagnosis?*
- *What would you do?*

This child has an acute infection in the submandibular area. The most likely focus of infection is an upper cervical lymph node, but other possible foci include the submandibular salivary gland, a branchial cleft cyst, or extension of infection from the lateral pharyngeal space. The likely causative organisms are *Staphylococcus aureus*, *Streptococcus pyogenes*, and anaerobes. The possibility of a mycobacterial infection and cat-scratch disease affecting the cervical lymph node should also be considered.

There are several different strategies for managing such a patient. These are:

- (i) Administer antimicrobial therapy active against the above pathogens, especially against *Staphylococcus aureus*, and re-examine the child after 1 or 2 days. Such therapy would depend on the local prevalence of methicillin-resistant *Staphylococcus aureus*. This might be cephalexin, dicloxacillin, or clindamycin. Although trimethoprim/sulfamethoxazole is active against *Staphylococcus aureus*, it is not very active against *Streptococcus pyogenes*.
- (ii) Aspirate the swelling and perform a Gram stain and culture of any aspirated material. If pus is aspirated this indicates the presence of an abscess, and adequate drainage should be performed. If no pus is aspirated, begin antimicrobial therapy as described above.

At the first visit a PPD should be placed.

If, at the follow-up visit, the abnormality has progressed, the following strategies, which are not mutually exclusive, should be considered:

- (a) Aspiration of the swelling.
- (b) Image the neck (by computer tomography) to determine more precisely the location of the disease. This should certainly be performed if conditions other than cervical lymphadenitis are considered likely. If an abscess is demonstrated this should be drained surgically.
- (c) Admission of the patient to hospital for intravenous antimicrobial therapy and for observation of the course of illness. If there is no improvement or the disease progresses, surgical drainage or biopsy should be performed.

If the PPD is 5 mm or greater in diameter at 48 hours, a mycobacterial infection is likely. This should be addressed as follows, assuming that there is no specific tuberculosis exposure history: a specimen from the node for

histology, acid-fast stain, and mycobacterial culture should be obtained, either by needle aspiration or biopsy, preferably excisional biopsy.

Also a chest X ray should be performed.

If the chest X ray suggests the possibility of pulmonary tuberculosis, three early morning gastric aspirate specimens for mycobacterial stain and culture should be obtained and antituberculosis therapy should be initiated. This strategy should also be used if there is a history of a tuberculosis contact.

If the chest X ray is normal:

- (a) if the PPD is 5–9 mm in diameter, await the results of the culture. If a nontuberculous mycobacterium grows and the node has been removed, no further management is necessary; if the node could not be removed, antimicrobial therapy according to the organism's identity should be attempted.
- (b) if the PPD is 10 mm or greater in diameter, initiate antituberculosis management while awaiting culture results. If the culture remains negative, a course of therapy for tuberculosis should be completed.

CASE 89. A 17-year-old boy presents with a 1-day history of back pain followed by loss of urinary and bowel control, ability to walk, and sensation in his legs. There is no history of preceding trauma, febrile illness, diarrhea, respiratory symptoms, foreign travel, or contact with other individuals with similar symptoms. He is, however, sexually active. He was fully immunized as a child. Examination reveals a generally healthy-appearing boy who is afebrile. The only abnormalities are referable to the nervous system. He cannot move his lower limbs or sit up unsupported, and the deep tendon reflexes in the lower limbs are absent. Sensory examination reveals a level in the mid-thoracic area below which he has decreased or absent sensation. The cremasteric and superficial abdominal reflexes are absent. The anal “wink” is absent, and he has lost bladder control. The upper limbs and cranial nerves are normal, as are his higher functions. There is no back tenderness.

- *What might this be due to?*
- *What should be done?*

This patient has acute paraplegia, indicating spinal cord disease. The differential diagnosis, in the absence of trauma, includes diseases within the cord itself (intramedullary) or outside the cord but within the spinal canal and compressing the cord (extramedullary). They include the following:

Abscess in the spinal epidural or subdural space. Such abscesses are hematogenous in origin and are usually caused by *Staphylococcus aureus*.

Hemorrhage (intra- or extramedullary) due to a tumor, cyst, or arteriovenous malformation.

Transverse myelitis: This may be due to an infection or due to an immune reaction to an infection. Several different infectious agents have been implicated, including Epstein–Barr virus, cytomegalovirus, human immunodeficiency virus (HIV), and *Schistosoma mansoni*.

Infarction of the cord, due to vasculitis, as can occur with spinal tuberculosis, or due to emboli.

Acute spinal cord disease constitutes a medical emergency. The most important elements of management are (a) to ensure adequate ventilation and perfusion, which was the case here, and (b) to diagnose or exclude immediately remediable causes of cord compression (compressive myelopathy), namely a tumor, an abscess, or a hemorrhage. This necessitates gadolinium-enhanced magnetic resonance imaging (MRI) of the spine and cord. If this cannot be performed, a computer tomography scan or a myelogram should be performed. Dexamethasone should be administered to reduce swelling of the cord. Compressive lesions should be treated surgically, with submission of any removed material for histology and for culture for bacteria, mycobacteria, and fungi. If a compressive myelopathy has been excluded, spinal fluid should be obtained to determine whether the disease is inflammatory or not. Further imaging may help to determine the extent of disease, and thus the particular nosologic entity, such as multiple sclerosis, neuromyelitis optica, acute disseminated encephalomyelopathy (ADEM), or only transverse myelitis.

In this patient the MRI showed an abnormal signal in the thoracic cord, with mild cord swelling, but no well-circumscribed lesion. This was most compatible with acute transverse myelitis. He was treated with dexamethasone, but showed very little improvement. He subsequently developed demyelination in his brain, leading to a diagnosis of ADEM. He has remained paraplegic and severely disabled.

Reference:

Kaplin AI, Krishnan C, Deshpande DM et al: Diagnosis and management of acute myelopathies. *The Neurologist* 2005; 11: 2–18.

CASE 90. An 18-month-old girl presents with a 1-day history of fever, unwillingness to eat, and inability to move her neck. Also her voice has developed a muffled quality. On examination she is mildly distressed from pain. Her temperature is 38.6°C, and her respiratory rate is 30/minute. She keeps her neck still, not flexing or moving it from side-to-side. There is mild

cervical lymphadenopathy. Her throat is difficult to visualize due to the pooling of secretions there, and the cry is slightly muffled. However, she is not in any respiratory distress, and her lung examination is normal. Her neurological examination, including her mental status, is normal.

- ***What is the differential diagnosis?***
- ***What would you do?***

This child has an inflammatory disease, probably infectious, in the pharyngeal area. The differential diagnosis includes peritonsillar abscess, which is unusual in children as young as this child, retropharyngeal abscess, and lateral pharyngeal space abscess. These latter two conditions arise from infection spreading to the lymphoid tissue around the pharynx. A retropharyngeal abscess can also arise from infection in the cervical vertebra (e.g. due to tuberculosis) rupturing anteriorly. The presence of stridor and breathing difficulty, together with pooling of secretions would suggest the possibility of epiglottitis. Although meningitis can present with neck stiffness and fever, it is often associated with some change in mental status such as extreme irritability or depressed level of consciousness. In addition it is not associated with voice changes or with pooling of secretions in the pharynx (unless the patient is in coma).

Management of a patient such as this should entail ensuring an adequate airway (which is the case here) and establishing an anatomic diagnosis. The importance of making a diagnosis is for the consideration of surgical drainage of an abscess. A lateral X ray of the neck can demonstrate the airway and abnormal tissue impinging on it (Figure 90.1). Normally in this view, taken when the neck is in extension, the width of the retropharyngeal space is approximately equal to that of the anteroposterior width of the cervical vertebral body.

This figure shows the width of the retropharyngeal space far exceeding that of the vertebra and the airway being pushed forward. These are characteristic of a retropharyngeal abscess or retropharyngeal cellulitis (phlegmon). Better definition of the anatomy of the disease, which might be necessary for deciding about the need for surgical drainage, can be obtained with a computer tomography scan (CT) scan. This is shown in Figure 90.2.

This shows a large hypodense area behind the pharynx, suggestive of an abscess.

Therapy consists of two elements: antimicrobial and surgical drainage. In many cases the retropharyngeal infection consists of a phlegmon, without clear abscess formation. In such cases, antimicrobial therapy alone usually results in clinical improvement. This should be directed at *Staphylococcus aureus*, streptococci, anaerobes, and small Gram-negative rods, such as *Haemophilus influenzae* and *Eikenella corrodens*. A combination of clindamycin and a third-generation cephalosporin would be suitable. Surgical drainage



FIG. 90.1. *Lateral neck X ray, showing abnormal swelling anterior to the cervical spine, pushing the airway anteriorly.*

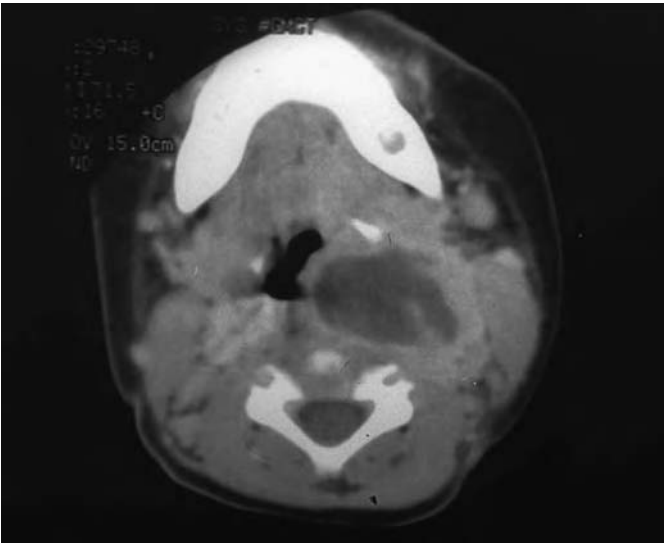


FIG. 90.2. *A CT scan of the patient's neck, showing a large retropharyngeal abscess.*

might become necessary in the case of a large abscess. This was performed in this child. The main complications of pharyngeal abscesses are impingement on the airway with airway obstruction, compression or erosion of the large vessels in the neck (jugular vein or carotid artery), and extension along the prevertebral fascia into the mediastinum.

Reading:

Craig FW, Schunk JE: Retropharyngeal abscess in children: clinical presentation, utility of imaging, and current management. *Pediatrics* 2003; 111: 1394–1398.

CASE 91. A 2-year-old boy presents with a history of limping and fever for 2 days. There is no history of trauma. On examination he is mildly ill-appearing with a temperature of 38.7°C. He has multiple mosquito bites on his legs. His gait suggests that he has pain when moving and bearing weight on his right lower limb. When he is examined lying supine, he does not allow the examiner to move his right thigh and hip. If the thigh is held still, movement of the ankle is normal. Movement of the knee cannot readily be tested without movement around the hip. Examination of the bones does not clearly reveal bony tenderness.

- *What is the differential diagnosis?*
- *What should be done?*

In making a diagnosis of the cause of a limp one should ask three questions: (a) Is the limp due to pain or weakness? (b) Where is the abnormality? (c) What is the pathology?

- (a) In this case the limp seems to be due to pain, not to weakness.
- (b) The areas that can give rise to pain on walking are, broadly, the back, pelvis, hip joint, thigh, knee joint, leg, ankle joint, foot, and shoe. In this case it seems to be the hip joint, the thigh, the knee joint, or proximal tibia.
- (c) The presence of fever suggests that the cause is probably an infection, although a noninfectious, but inflammatory disease, for example, juvenile rheumatoid disease, or a malignancy, can also cause fever

The disease with the potential to cause the greatest damage is septic arthritis of the hip, which, if not appropriately treated, can result in avascular necrosis of the femoral head. The other important diagnostic considerations are septic arthritis of the knee and osteomyelitis of the femur or the tibia.

Therefore the child should be evaluated for these possibilities.

Septic arthritis of the hip joint: An ultrasound examination of the hip joint can be used to determine whether or not there is fluid in this joint. If this shows that fluid is present, the child should undergo aspiration of the hip joint, with cell count, Gram stain, and culture of the fluid obtained. If the fluid is purulent, the joint should be washed out. Antimicrobial therapy should be directed at *Staphylococcus aureus*, unless the Gram stain of the fluid suggests the presence of a different organism. In populations in which immunization against *Haemophilus influenzae* is not in widespread use, this organism is also a likely cause of septic arthritis in children. Other causes of

septic arthritis in children are *Kingella kingae*, *Streptococcus pyogenes*, *Streptococcus pneumoniae*, and *Salmonella* spp.

If the hip ultrasound is normal further imaging studies should be performed. The optimal study is a magnetic resonance imaging (MRI) scan of the lower limb, which can demonstrate areas of osteomyelitis or arthritis. If this cannot be done a technetium bone scan should be performed. This can demonstrate increased blood flow around an inflamed joint or increased uptake in an area of osteomyelitis.

Although plain X rays can usually be performed rapidly, they are rarely helpful in the early stages of acute osteomyelitis because bony changes take 10–14 days to become manifest radiologically. Their value lies primarily in demonstrating fractures.

Management of skeletal infection consists of antimicrobial therapy, and, in the case of septic arthritis, washing out of the joint to reduce the damage caused by the inflammatory reaction to synovial cartilage. When to initiate antimicrobial therapy can be influenced greatly by the availability of the above imaging studies and the apparent severity of the child's clinical condition, which determines the urgency with which therapy should be initiated. Ideally, if a precise focus of bone or joint disease can be determined clinically, this should be aspirated, with specimens sent for Gram stain and culture, before antimicrobial therapy is started. The microbiological yield might be improved by injecting some of the specimen into a blood culture bottle. In patients such as the one presented, the site for aspiration should be guided by the results of imaging. Blood cultures should also be performed. However, they are positive in only about 50% of cases of osteomyelitis in children. If antimicrobial therapy is initiated before appropriate specimens are obtained, a microbiological diagnosis might never be made and the full course of therapy for skeletal infection will need to be empiric.

Antimicrobial therapy should be directed primarily at *Staphylococcus aureus*. In areas with low prevalence rates of methicillin-resistant *Staphylococcus aureus* (MRSA), an antistaphylococcal penicillin or first-generation cephalosporin should be used. In areas with high prevalence rates of MRSA, initial therapy should consist of vancomycin or clindamycin. Vancomycin should be chosen if the child is severely ill or if the prevalence rate of clindamycin-resistance is high. The duration of therapy should be at least 3 weeks for septic arthritis and 4 weeks for osteomyelitis. In cases in which an organism is identified and antibiotic susceptibilities can be determined, oral therapy can be used once the patient's condition has improved, provided that there is a suitable agent, for example, cephalexin or clindamycin.

Reading:

Hambleton S, Berendt AR: Bone and joint infections in children. In: Pollard AJ, McCracken GH, Jr Finn A (editors): Hot Topics in Infection and Immunity in Children. Kluwer Academic/Plenum Publishers, New York, 2004, pp. 47–62.

CASE 92. (HYP). A newborn boy is noted to have purple skin lesions at birth. He is born by normal vaginal delivery at term. The maternal history is normal, except that the mother had a mild febrile illness at about 24 weeks of pregnancy, lasting a few days. She also has a 3-year-old daughter who is well. On examination the baby is slightly lethargic. The weight is 2.6 kg, length 45 cm, and head circumference 32 cm. There is moderate pallor. The heart and lungs are normal, and the abdominal examination reveals marked enlargement of the liver and spleen. The skin shows diffuse petechiae. There is no lymphadenopathy.

- *What is the differential diagnosis?*
- *What would you do?*

There are three main immediate potential threats to this baby's life: bacteremia, complications from severe anemia, and hemorrhage. Therefore initial attention should be directed at detecting or quantifying these threats, by performing blood culture and urine cultures, blood count, blood smear, platelet count, Coombs test, prothrombin time (PT), and partial thromboplastin time (PTT). In view of the petechiae, a lumbar puncture should not be performed.

Bacteremia: Antibiotic therapy active against *Streptococcus agalactiae*, *Listeria monocytogenes*, and enteric bacilli should be administered intravenously.

Severe anemia: Considering the clinical findings, in particular hepatosplenomegaly, this might be caused by hemolytic disease of the newborn due to maternal–fetal blood group incompatibility. Arrange for Group O Rhesus negative blood, which might be necessary to transfuse into the infant before the cause of the anemia has been determined.

Hemorrhage: Management strategies to prevent or treat the infant for life-threatening hemorrhage will depend on the results of the above investigations.

Attention can now be paid to other diagnoses which are less threatening in the short term. These are:

- (a) intrauterine infection of which the following are the most important:
 - (i) cytomegalovirus (CMV) infection. This is the most likely intrauterine infection because it is the most frequent;

- (ii) congenital rubella;
- (iii) congenital toxoplasmosis;
- (iv) congenital syphilis – this is often associated with maculopapular skin lesions, mucosal lesions, and lymphadenopathy. The manifestations might not be present at birth;
- (v) congenital human immunodeficiency virus (HIV) infection, which is also usually associated with lymphadenopathy.

Unusual congenital infections that can present in this fashion include malaria, tuberculosis, American trypanosomiasis, relapsing fever, and lymphocytic choriomeningitis virus infection.

(b) congenital leukemia.

A good ophthalmological examination can be very helpful in distinguishing between several intrauterine infections. Congenital rubella causes diffuse speckled pigmentation of the retina (“salt-and-pepper” pigmentation), congenital CMV infection causes chorioretinitis in any area, while congenital toxoplasmosis causes chorioretinitis particularly at the posterior pole of the eye (see Case 65).

The use of serologic tests, utilizing IgM titers to differentiate maternal infection from fetal infection are of value in this clinical circumstance. These are used for diagnosing rubella, and toxoplasmosis, and may be helpful in diagnosing congenital syphilis. CMV infection should be diagnosed by culture of the infant’s urine, and HIV infection is diagnosed by DNA PCR testing of the infant’s blood.

Treatment is available for patients with the bacterial or protozoal infections, and HIV infection. Gancyclovir, which is active against CMV, has not been shown to be of significant benefit in cases of congenital CMV infection.

Reading:

Andrews JI: Diagnosis of fetal infections. *Curr Opin Obstet Gynecol* 2004; 16: 163–166.

CASE 93. An 18-month-old, previously healthy boy presents with fever and difficulty breathing. On examination he is ill-appearing, with grunting, a temperature of 39°C, heart rate of 180/minute, blood pressure of 80/40, and respiratory rate of 80/minute. The chest examination reveals intercostal, subcostal, and suprasternal retractions. The lungs are resonant to percussion and clear to auscultation. A diagnosis of pneumonia with possible

bacteremia is made, and the child is hospitalized, given oxygen by nasal cannula, and treated with cefotaxime intravenously. A chest X ray is read as showing patchy right lower lobe pneumonia. Two days after admission he becomes obtunded. The peripheral pulse cannot be palpated, and the heart sounds are soft. He is intubated and ventilated, and given intravenous fluids and pressors. However, his pulse remains impalpable. Review of the initial chest X ray suggests cardiomegaly, and the admission blood culture is reported to be growing small Gram-negative rods. An electrocardiogram shows low amplitude complexes.

- ***What might have happened?***
- ***What would you do?***

The child is in shock, has cardiomegaly, soft heart sounds, low amplitude complexes on electrocardiogram, and has Gram-negative rod bacteremia.

Although Gram-negative sepsis might have explained shock on admission, it seems less likely to have developed after the child has received broad-spectrum antimicrobial therapy for 2 days. Refractory shock, soft heart sounds, and the radiologic appearance of cardiomegaly suggest myocarditis or pericarditis with cardiac tamponade.

Furthermore, the organism present in the blood could have localized within the pericardium causing a purulent (suppurative) pericarditis. Therefore an echocardiogram should be performed immediately to differentiate between myocarditis and pericarditis, and with a view to directing the immediate performance of pericardiocentesis.

In this case, due to the unavailability of echocardiography, a pericardiocentesis was performed at the bedside. After withdrawal of a few milliliters of purulent fluid, the blood pressure rose. A pericardiotomy was subsequently performed, and the child recovered. The blood culture isolate was *Haemophilus influenzae* type b.

Suppurative (purulent) pericarditis is a potentially lethal disease because it can cause cardiac tamponade. It is difficult to diagnose in children because its signs and symptoms are often ascribed to pneumonia, as initially occurred in this patient. Furthermore, it is relatively uncommon, and thus seldom considered as a diagnosis. The diagnosis is particularly difficult to make in young children. Older children can complain of chest pain that is worse on lying than sitting and that may radiate to the upper back or they may complain of abdominal pain. A pericardial rub may be heard. This is easier to detect when the patient is sitting up and leaning forward. Once a significant effusion has developed, the heart sounds are soft, and the signs of cardiac tamponade may be present, namely hypotension, elevated jugular venous

pressure, and an increased pulsus paradoxus. Bacteria can reach the pericardium via several different routes, namely hematogenously, from infection in adjacent lung tissue, from the mediastinum, or from an infradiaphragmatic infection. The main causes are *Staphylococcus aureus*, *Haemophilus influenzae* type b, *Streptococcus pneumoniae*, and *Neisseria meningitidis*. In adults anaerobes are also important causes of purulent pericarditis.

Management consists of pericardiocentesis, both to establish the diagnosis and to relieve the tamponade, pericardial drainage, and antimicrobial therapy. This should be directed initially at the likely causative organisms and adjusted once an organism has been cultured. The optimal form of surgical drainage is controversial. Such procedures range from catheter drainage, the least invasive, through pericardiostomy tube drainage, to pericardiectomy, the most invasive. Constrictive pericarditis is a late sequela of this infection.

Reading:

Dupuis C, Gronnier P, Kachaner J et al: Bacterial pericarditis in infancy and childhood. *Am J Cardiol* 1994; 74: 807–809.

Park S, Bayer AS: Purulent pericarditis. *Curr Top Infect Dis* 1992; 12: 56–82.

CASE 94. A 5-year-old boy presents with a history of left-sided facial swelling and pain for 1 day. There is no history of rhinorrhea or of nasal congestion. On examination he has a temperature of 38°C but does not appear ill. He has fairly marked swelling and erythema of the left side of his face extending from the mouth to the lower eyelid, with some swelling of the lower eyelid. There is tenderness over the swollen area. There is no conjunctival injection, and the ocular movements are full. There is mild submandibular lymphadenopathy.

- *What is the differential diagnosis?*
- *What should be done at this time?*

The main differential diagnosis includes maxillary sinusitis and odontogenic infection. The above scenario does not include an oral examination, which is essential.

This shows severe and extensive dental caries, with swelling and marked tenderness of the gums in the left upper jaw. The diagnosis is an odontogenic abscess, and the treatment is antimicrobial therapy and surgical drainage of the abscess. A dental X ray can help in localizing the specific tooth affected (Figure 94.1), and a panoramic radiograph (orthopantomogram) is helpful in determining the extent of the dental and periodontal disease (Figure 94.2).

Such abscesses can spread through the bones of the jaw into several spaces around the mouth and the pharynx, including the lateral pharyngeal space, the sublingual space, the pterigomandibular space, the buccal space, the submandibular space, the submasseteric space, and the temporal space. A computer tomography scan can help to delineate such spread. In severe cases, as described here, the affected tooth or teeth must be removed. If there is evidence of abscess formation in one of the above-mentioned spaces, surgical drainage should be performed. Antimicrobial therapy should be directed mainly at streptococci and anaerobes. Although penicillin has been very effective in patients with such infections, there is a rising rate of resistance to this drug among anaerobes. Alternatives include amoxicillin/clavulanate (or ampicillin/sulbactam for parenteral therapy) or clindamycin.

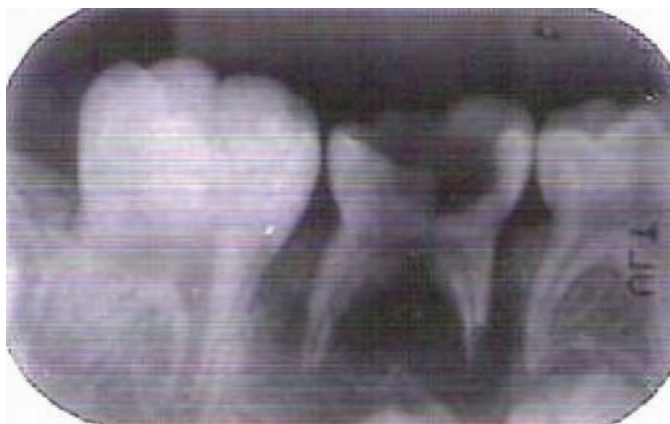


FIG 94.1. A dental abscess resulting from a carious tooth in a 6-year-old girl.



FIG 94.2. Orthopantomogram showing a lytic lesion of mandible due to an odontogenic abscess (arrow head).

Prevention depends on good dental hygiene and nutrition, including the drinking of fluoride-fortified water to prevent the development of dental caries.

Reading:

Flynn TR: The swollen face. Severe odontogenic infections. *Emerg Med Clin N Am* 2000; 18: 481–519.

Wayne DB, Trajtenberg CP, Hyman DH: Tooth and periodontal disease: a review for the primary-care physician. *Southern Med J* 2001; 94: 925–932.

CASE 95. A 2-year-old girl presents with a history of refusing to walk for 1 day and the suggestion of abdominal and back pain for a few days. There is no history of trauma or of preceding respiratory tract infection. On examination the child is afebrile and well appearing but in some discomfort. She is reluctant to bear weight on her legs and refuses to walk. The abdomen is soft and nontender; however, there is tenderness over the lumbar spine. Movement of the hips, knees, and ankles is normal, and a rectal examination is normal.

- *What is the differential diagnosis?*
- *What should be done?*

The clinical features suggest a problem in the lumbar spine. The possible disease entities include infections, injuries, and tumors. The most serious considerations are those diseases that can result in spinal cord compression, such as spinal epidural abscess or hematoma, which are rare in children, or a tumor such as neuroblastoma. The most likely infections of the spine are discitis and vertebral osteomyelitis. Several imaging studies may need to be employed to make a diagnosis:

- (i) A plain X ray of the spine: this may show narrowing of the disc space in discitis, and bony erosion in osteomyelitis, but these changes are absent early on in these disease processes;
- (ii) A technetium bone scan will often show increased uptake in the case of osteomyelitis, but may be normal in discitis;
- (iii) A magnetic resonance imaging (MRI) scan of the spine has the advantage of assisting in the diagnosis of all the likely diagnoses, including osteomyelitis, discitis, as well as diseases of the soft tissue, epidural space, and spinal cord. If available, this is the study of choice.

In the case of discitis or osteomyelitis, it is optimal to obtain an aspirate of presumed infected material in order to make a microbiological diagnosis, and

thus provide optimal antimicrobial therapy. This requires imaging guidance and is frequently not possible. A blood culture and a tuberculin skin test should be performed. Therapy should be directed at the most likely pathogen, namely *Staphylococcus aureus*. However, consideration should also be given to other pathogens such as *Kingella kingae* and *Salmonella enteritidis*.

Imaging studies in this child failed to show any abnormality. A provisional diagnosis of discitis was made. She was treated with analgesia and nafcillin and made a rapid recovery.

Discitis represents inflammation of the intervertebral disc. It affects mainly children younger than about 4 years of age and teenagers. There is controversy as to whether all cases are due to infection. In cases of infection, microorganisms are believed to reach the disc from the metaphyseal arteries supplying the vertebra. When a microorganism is isolated, the most common is *Staphylococcus aureus*. The main clinical findings are irritability, refusal to walk, abdominal pain, and back pain or tenderness. There may or may not be fever. These findings may have an insidious onset, over days or weeks.

Reading:

Early SD, Kay RM, Tolo VT: Childhood diskitis. J Am Acad Orthop Surg 2003; 11: 413–420.

CASE 96 (HYP). An 8-month-old girl presents to a South African hospital in 1976 with a history of fever and irritability. On examination she has a temperature of 39°C and is slightly obtunded. Her cerebrospinal fluid (CSF) reveals 2000 leukocytes per mm³, of which 95% are neutrophils, and protein and glucose concentrations of 150 and 27 mg/dl, respectively. The Gram stain shows numerous Gram-positive diplococci. A provisional diagnosis of pneumococcal meningitis is made. She is treated with intravenous penicillin G, at a dosage of 300,000 units per kilogram per 24 hours divided into six doses. Her condition progressively deteriorates. Twenty-four hours after her admission the CSF cultures reveals *Streptococcus pneumoniae*. Her therapy is continued. However, her condition deteriorates progressively and she dies 36 hours after admission.

• **Why might her condition have deteriorated?**

Clinical deterioration in a patient thought to be treated appropriately may be due to:

- (a) the natural progression of or a complication of the disease, irrespective of treatment;

- (b) wrong diagnosis – in the case of an infection, this would include wrong microbial/etiological diagnosis (“wrong bug”);
- (c) in the case of an infection, the use of an antimicrobial agent to which the causative agent is resistant (“wrong drug”).

In this case, the diagnosis was clearly pneumococcal meningitis. Deterioration might have been due to cerebral swelling or brain infarction, well-recognized complications of this disease. However, resistance of the organism to penicillin would be another possibility. In this case the minimal inhibitory concentration (MIC) of the isolate of *Streptococcus pneumoniae* to penicillin was >2 $\mu\text{g/ml}$. This indicates resistance to penicillin.

Historically and geographically, this child would have represented one of the early cases of the problem of penicillin-resistant *Streptococcus pneumoniae*, which became a major clinical, including nosocomial, problem in South Africa at that time, and which has since become a major problem worldwide.

Reference:

Jacobs MR, Koornhof HJ, Robins-Brown RM et al: Emergence of multiply resistant pneumococci. N Engl J Med 1978; 299: 735–740.

CASE 97 (HYP). A 10-year-old boy presents with fever and pain in his left leg, a few days after scraping the leg in a bicycle fall. On examination he has a temperature of 39°C , and swelling and erythema of the left leg, extending from the ankle to the mid-calf. He is treated with nafcillin 1 g every 6 hours intravenously and with elevation of the leg. After 48 hours he remains febrile and he complains of pain in his right knee. On examination there is marked tenderness just above the medial femoral condyle.

- **Why might he not be improving and why might he have developed new symptoms?**

The initial presentation suggests a diagnosis of cellulitis, the most common causes of which are *Staphylococcus aureus* and *Streptococcus pyogenes*. Nafcillin would be expected to be active against both these organisms. Failure of the patient’s condition to improve suggests:

- (a) a complication, such as abscess formation, for which surgical drainage is necessary;
- (b) a different causative organism, which is resistant to nafcillin, for example, a Gram-negative rod (very unlikely unless the injury had occurred in water or the wound had been contaminated with soil); or

- (c) a resistant strain of staphylococcus or streptococcus. The new clinical manifestation, namely tenderness in the opposite femur, strongly suggests the development of osteomyelitis. Its manifestation during therapy provides further evidence that the initial causative organism is resistant to nafcillin. In 2005 methicillin-resistant *Staphylococcus aureus* strains (which are resistant to all penicillins and cephalosporins) are widespread in the United States. Penicillin-resistant strains of *Streptococcus pyogenes* have not yet been reported.

Discussion of cases 96 and 97

Antimicrobial resistance: There are two different definitions of this phenomenon:

(a) clinical resistance, in which the organism is resistant to the antimicrobial agent at a concentration attainable in infected tissue with the use of nontoxic dosages of the agent; and (b) microbiological resistance, in which the organism is more resistant than it was previously, even if it does not meet criteria for clinical resistance. Resistance affects antiviral, antibacterial, antifungal, and antiparasitic agents, but this discussion will be confined to antibacterial agents.

Bacteria develop resistance to antimicrobial agents by changing their genetic make-up. They accomplish this by acquiring genes from other sources, including from other bacteria, or by mutations. Mutations occur approximately every 10^8 to 10^9 divisions. For a single organism dividing every 30 minutes, it takes less than 1 day for a mutation to occur. Increase in the prevalence of resistant bacteria results from selective pressures placed on bacterial populations by their exposure to antimicrobial agents. The susceptible organisms are eliminated and the resistant ones survive, becoming the predominant population. This phenomenon (natural selection) was described by Charles Darwin in 1859, before bacteria had been identified.

How do resistant bacteria spread? The genetic material confirming resistance can spread among organisms, including across genera. More important, however, is spread of the resistant organisms themselves, in feces, secretions, and, of extreme significance within hospitals, on the hands of health-care workers.

Mechanisms of antimicrobial resistance in bacteria: There are four main mechanisms:

1. Barrier to entry of the organism into the bacterial cell. This operates mainly in Gram-negative bacteria, in which it is due to changes in porin proteins, which constitute the channels through which macromolecules enter the organism.

2. Efflux mechanisms: These are energy-dependent mechanisms whereby the organism pumps the antibiotic out of itself before the drug can exert its effect. This is important in resistance to tetracycline as well as to other agents.
3. Enzymatic alteration or destruction of the antibiotic: This mechanism is the most important mechanism of resistance to beta-lactams (the enzymes are beta-lactamases) and to aminoglycosides (the enzymes are acetylases, adenylylases, and phosphorylases). The following are examples: penicillin resistance in staphylococci (counteracted by antistaphylococcal penicillins such as oxacillin, cloxacillin, nafcillin, and methicillin); ampicillin resistance in *Haemophilus influenzae* and penicillin resistance in *Neisseria gonorrhoeae* (counteracted by third-generation cephalosporins); resistance to third-generation cephalosporins in enteric bacilli (conferred by “extended-spectrum beta-lactamases” and other types of beta-lactamases); resistance to aminoglycosides in enteric bacilli and *Pseudomonas aeruginosa*.
4. Alteration of the molecular targets of the antimicrobial agent, for example alteration of penicillin-binding proteins, resulting in penicillin resistance in pneumococci (Case 96), methicillin resistance in staphylococci (Case 97), and ampicillin resistance in enterococci.

More than one of the above mechanisms may operate in one organism synergistically.

Reading:

Berkowitz FE: Antibiotic resistance in bacteria. *Southern Med J* 1995; 88: 797–804.

Chambers HF: Community-associated MRSA – resistance and virulence converge. *N Engl J Med* 2005; 352: 1485–1487.

Kaplan SL: Treatment of community-associated methicillin-resistant *Staphylococcus aureus* infections. *Pediatr Infect Dis J* 2005; 24: 457–458.

CASE 98. A 17-year-old girl with short gut syndrome, a complication of systemic lupus erythematosus, is hospitalized with a tunnel infection of her Broviac catheter, used for hyperalimentation. Her weight is 56 kg. She is treated intravenously with vancomycin 1 g every 12 hours and amikacin 420 mg every 12 hours. Her catheter is removed. Her renal function deteriorates

as shown by a serum creatinine rising from 0.8 to 3.2 mg/dl over a period of a few days.

- ***What are the possible reasons for the deterioration in renal function?***

There are several possible reasons for her deterioration in renal function:

- (a) underlying renal disease complicating systemic lupus. This is unlikely to have caused such a sudden change, however.
- (b) infective endocarditis complicating the bacteremia and complicated by glomerulonephritis (infection of venous catheters has become an important cause of infective endocarditis).
- (c) toxicity from the antibiotics. Both vancomycin and amikacin can cause nephrotoxicity. Because they have narrow therapeutic ranges, their blood concentrations should be checked, the peaks to ensure therapeutic concentrations and the troughs to ensure adequate clearance. This was the cause of the renal dysfunction in this patient.

CASE 99A (HYP). A 3-year-old boy had bacterial meningitis as an infant. This was complicated by hydrocephalus, for which he has a ventriculoperitoneal shunt, and epilepsy, which has been well controlled with phenytoin, 40 mg twice per day. He presents with fever, headache, and vomiting. Cerebrospinal fluid (CSF) from the shunt reveals 200 leukocytes per mm³ with a Gram stain showing a few Gram-positive cocci in clusters. A shunt infection is diagnosed and the lower end of the shunt is externalized. Antimicrobial therapy with vancomycin intravenously and rifampin orally is initiated. The CSF culture grows out a coagulase-negative staphylococcus, and the antimicrobial therapy is continued. By the third day the child is afebrile and feeling much better, but on the fifth day he has a generalized seizure.

- ***Why might he have had the seizure?***

Possible causes for seizures in a child whose seizures have previously been well controlled:

- (a) brain injury resulting from the ventriculitis and meningitis. This is unlikely considering his improving condition.
- (b) decreased serum concentration of phenytoin resulting from the drug interaction between the metabolisms of phenytoin and rifampin. Rifampin induces the enzyme responsible for phenytoin metabolism,

resulting in an increased rate of phenytoin metabolism, and hence a decreased blood concentration of phenytoin, predisposing the child to a seizure.

CASE 99B (HYP). A 10-year-old boy with moderate persistent asthma, managed with albuterol inhalations and oral theophylline, presents with a cough and fever. He is diagnosed with pneumonia based on the finding of pulmonary infiltrates on chest X ray, and he is treated with erythromycin by mouth. After 4 days he develops vomiting and experiences a grand mal seizure.

- *What is the likely cause of these new clinical problems?*

Vomiting and seizures are manifestations of theophylline toxicity. The likely cause of theophylline toxicity in this child is interference with theophylline metabolism by erythromycin, resulting in an increased blood concentration of theophylline.

CASE 99C (HYP). A 17-year-old girl weighing 65 kg is diagnosed with pulmonary tuberculosis and treated with isoniazid 300 mg, rifampin 600 mg, and pyrazinamide 1250 mg daily by directly observed therapy. She is sexually active and taking oral contraceptive pills. After 2 months on treatment she complains to her health-care worker of nausea and vomiting of 1 week's duration. On examination by a physician there are no abnormal findings, specifically no jaundice and no right upper quadrant tenderness nor hepatomegaly.

- *What are the possible causes of her symptoms?*

The most important immediate consideration in an individual who is taking antituberculosis therapy and who develops nausea and vomiting is the possibility of drug-induced hepatotoxicity. If the therapy is not discontinued immediately, this can result in progression to hepatic failure that can prove fatal. Therefore when antituberculosis therapy is initiated, patients should be warned that if they suffer this symptom they should discontinue therapy and be seen by their physician immediately. Hepatotoxicity, which is rare in children, can be diagnosed by the demonstration of significantly elevated hepatic transaminase levels.

Another important cause of this symptom is pregnancy. According to the history she is sexually active and taking an oral contraceptive. This can be

rendered less effective by rifampin due to the induction of enzymes by rifampin, leading to an increased rate of metabolism of the oral contraceptive.

Other considerations, unrelated to therapy, include gastritis and cholecystitis.

Therefore she should have tests of liver function and damage, and a pregnancy test.

Discussion of cases 98, 99A–C:

Drug interactions are important considerations in any individual receiving more than one drug, because they may result in significant toxicities or reduction in therapeutic effects. Most interactions are the result of pharmacokinetic effects, in which the presence of one drug affects the blood or tissue concentration of another drug, but a few can result from pharmacodynamic effects. There are a few broad mechanisms by which interactions affect drug pharmacokinetics:

- (a) Drug A interferes with the intestinal absorption of drug B, thus reducing its activity, for example, aluminum hydroxide interferes with the absorption of several cephalosporins, isoniazid, ethambutol, itraconazole, ketoconazole, rifampin, and tetracyclines.
- (b) Drug A stimulates enzymes (particularly cytochrome oxidase P450 enzymes) that metabolize drug B, resulting in the increased rate of metabolism of drug B, and hence a decreased therapeutic effect of drug B, for example, rifampin causes a decreased therapeutic effect of atovaquone, benzodiazepines, oral contraceptives (Case 99C), corticosteroids, cyclosporine, several antiretrovirals, phenytoin (Case 99A), valproic acid, and warfarin.
- (c) Drug A inhibits enzymes that metabolize drug B, resulting in toxicity of drug B, for example, erythromycin increases the blood concentrations of amiodarone, benzodiazepines, carbamazepine, digoxin, phenytoin, theophylline (Case 99B), valproic acid, and warfarin. This phenomenon can be used therapeutically.
- (d) Drug A inhibits the excretion of drug B, for example, probenecid inhibits the excretion of penicillin. This is also used therapeutically in order to increase the serum concentration of penicillin.
- (e) Drug A and drug B have similar toxicities, resulting in additive toxicities, for example, both vancomycin and aminoglycosides are nephrotoxic, and their use in combination may increase the risk of nephrotoxicity (Case 98).
- (f) Drug A causes a toxicity that can affect the excretion of drug B, for example, amphotericin B causes renal dysfunction, which can result in

decreased excretion of several other drugs, such as 5-flucytosine, vancomycin, and aminoglycosides.

Antimicrobial agents may have theoretical adverse pharmacodynamic interactions, the evidence for which is largely controversial. For example, a drug inhibiting protein synthesis, for example, tetracycline, could reduce the rate of production of bacterial cell wall, the site of action of beta-lactams, thus reducing the effect of the beta-lactam.

Therefore it is important to consider drug interactions whenever prescribing. Specific antimicrobial agents that are at particularly high risk for causing interactions include rifampin, triazoles, and protease inhibitors.

Reading:

Rang HP, Dale MM, Ritter JM, Moore PK: Pharmacology. 5th edition. Churchill Livingstone, Edinburgh, 2003, pp. 718–723.

CASE 100. An 11-year-old girl who has recently immigrated to the United States from Honduras presents with a history of pain and swelling of her left knee for about 3 weeks, which was preceded by injury to the knee. She has lost about 2 kg in weight in the past few weeks. She developed fever the night before presentation. She has no known contact with an individual with tuberculosis. On examination she is well appearing and afebrile. The proximal part of her left tibia is mildly swollen, erythematous, and tender. The knee joint has full range of movement and no fluid can be detected within the joint. Of note is the absence of pallor, jaundice, lymphadenopathy, hepatomegaly, and splenomegaly. X rays of the knee are shown in Figures 100.1 and 100.2.

- ***What is your differential diagnosis?***
- ***What would you do?***

This child has several lucencies within the proximal tibial metaphysis. The differential diagnosis includes bone cysts, tumors, and infections. The indolent nature of the illness suggests that, if this is an infection, it may be caused by an organism that is slow-growing such as *Mycobacterium tuberculosis*. Nevertheless, pyogenic infections, such as those caused by *Staphylococcus aureus*, can become chronic in the absence of therapy.

It is important to make a specific diagnosis, by performing a biopsy, with performance of histology, routine bacterial cultures, and cultures for mycobacteria and fungi. The pace of this illness is slow, so there is no urgency in instituting any form of management.



FIG 100.1. X ray showing lucencies in the proximal tibial metaphysis (anterior-posterior view).

The child underwent a bone biopsy. The specimens revealed Gram-positive cocci, which were *Staphylococcus aureus*. This is a case of subacute or chronic staphylococcal osteomyelitis resulting in the formation of Brodie's abscesses. Chronic osteomyelitis is characterized by bone necrosis, due to ischemia. This area of bone, the sequestrum, becomes surrounded by new bone formation, the involucrum.

There are several different types of subacute osteomyelitis, whose differential diagnosis varies according to the radiological appearance. These are shown in Table 100.1.

This child's X rays indicate Brodie's abscesses of type IA.



FIG 100.2. X ray showing lucencies in the proximal tibial metaphysis (lateral view).

As opposed to the treatment of patients with acute hematogenous osteomyelitis, which is medical (antimicrobial), the treatment of patients with chronic osteomyelitis entails surgical debridement of necrotic bone as well as antimicrobial therapy active against the causative organism. This should be prolonged and can usually be administered by the oral route (for about 3 months). However, there is a substantial chance of relapse.

TAB. 100.1: Different types of subacute osteomyelitis.

Type	Site	Radiologic appearance	Differential diagnosis
IA	metaphysis	lucency; no reactive bone	eosinophilic granuloma
IB	metaphysis	lucency; sclerotic margins	typical Brodie's abscess
II	metaphysis	cortical erosion	osteogenic sarcoma
III	diaphysis		osteoid osteoma
IV	diaphysis	"onion peel"	Ewing sarcoma

Reading:

Ramos OM: Chronic osteomyelitis in children. *Pediatr Infect Dis J* 2002; 21: 431–432.

Lopes TD, Reinus WR, Wilson AJ: Quantitative analysis of the plain radiographic appearance of Brodie's abscess. *Invest Radiol* 1997; 32: 51–58.

CASE 101 (COMP). Three days after undergoing an exchange transfusion for hyperbilirubinemia, a full-term, 3.1 kg boy develops temperature instability, vomiting, abdominal distension, and bloody stools. Examination reveals a very ill, gray-appearing, hyporesponsive infant, with poor perfusion, heart rate of 180/minute, blood pressure of 30/10 mm Hg, and respiratory rate of 80/minute. No abnormalities of the heart or lungs are detected. The abdomen is markedly distended, and bowel sounds are absent. The anterior abdominal wall is a deep red to purple color.

- *What do the history and physical examination indicate?*
- *What has caused this problem?*
- *How would you manage the child?*

The infant is in shock. He has peritonitis due to bowel perforation, which is the likely cause of the shock. The main causes of bowel perforation in a newborn infant are necrotizing enterocolitis or gastric perforation. In very low birthweight infants, isolated small bowel perforation may occur in the absence of necrotizing enterocolitis. The history in this patient is highly suggestive of the diagnosis of necrotizing enterocolitis. Although very rare, this is a recognized complication of exchange transfusion.

The immediate management should entail fluid resuscitation of the infant and artificial ventilation. During this time, antimicrobial therapy active against bowel organisms should be initiated, and abdominal X rays should be performed to attempt to clarify the cause of the problem. As soon as the infant is stable, an exploratory laparotomy should be performed.



FIG 101.1. *Abdominal X ray showing pneumatosis intestinalis.*

Necrotizing enterocolitis is a disease of the bowel characterized by mucosal necrosis, with entry of bacteria and gas from the bowel lumen into the bowel wall. Although its pathogenesis is incompletely understood, it appears to be due to a combination of ischemia, infection, and the presence of protein within the bowel lumen. The vast majority of cases affect premature infants who have received some feeding, but conditions that affect intestinal blood flow, such as exchange transfusion and congenital heart disease, can also result in this disease.

The clinical features of necrotizing enterocolitis are vomiting (or increased gastric residuals in a tube-fed infant), abdominal distension, and hematochezia. There are often associated features suggesting a systemic

infection such as hypothermia, apnea, and poor perfusion. The early radiological feature is dilated bowel loops, and the pathognomonic feature is pneumatosis intestinalis (gas within the bowel wall) (Figure 101.1).

If the disease progresses, intestinal perforation may occur, resulting in discoloration of the abdominal wall and scrotum, and free intraperitoneal gas (Figure 101.2).

Gas within the portal venous system and the liver may also be observed in severe cases (Figure 101.3).

Management consists of ensuring adequate perfusion, nasogastric tube suction, broad-spectrum antimicrobial therapy, directed at streptococci, enteric Gram-negative rods, and anaerobes, and frequent clinical and radiological monitoring for signs of progression, in which case surgery might be indicated. Such signs include free gas, intraperitoneal fluid, palpation of an abdominal mass, and persistence of dilated bowel loops.

Complications include the sepsis syndrome resulting from either the intestinal disease or vascular catheters required for management, enteric fistulae, short gut syndrome, and intestinal strictures.



FIG 101.2. *Abdominal X ray showing free intraperitoneal gas.*



FIG 101.3. An abdominal X ray showing gas in the portal vein and the liver, resulting from necrotizing enterocolitis in a 9-month-old child who had suffered shock as a result of acute gastroenteritis.

Reading:

Kafetsis DA, Skevaki C, Costalos C: Neonatal enterocolitis: an overview. *Curr Opin Infect Dis* 2003; 16: 349–355.

Caty MG, Azizkhan RG: Necrotizing enterocolitis. In: Ashcraft KW, Murphy JP, Sharp RJ, Sigalet DL, Snyder CL (editors): *Pediatric Surgery*. 3rd edition. WB Saunders, Philadelphia, 2000.

CASE 102. A 6-year-old shepherdess from rural South Africa presents with a 3-month history of abdominal discomfort. There are no other symptoms. On examination she is well appearing and afebrile. The only abnormality is a vague, nontender mass in the epigastrium, possibly within the liver (Figure 102.1).

- *What is your differential diagnosis?*
- *What would you do?*

A tender hepatic mass associated with fever would suggest the presence of a hepatic abscess, either bacterial or amebic in origin, while the findings in this child suggest a noninflammatory mass, such as a cyst or tumor in the

liver or pancreas. The child's occupation exposes her to echinococcosis (infection with the dog tapeworm, *Echinococcus granulosus*).

A simple diagnostic test is an ultrasound of the abdomen (Figure 102.2).

This shows a large cyst within the liver, compatible with a hydatid cyst.



FIG 102.1. The swelling in the child's epigastrium.

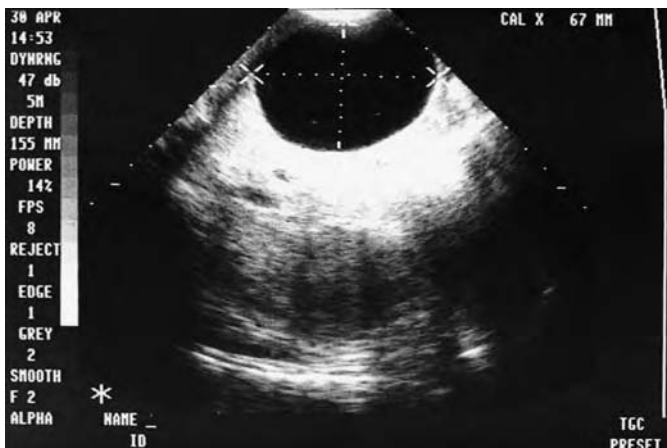


FIG 102.2. A large cyst within the liver, probably a hydatid cyst (*Echinococcus*).

Life Cycle of *Echinococcus granulosus*

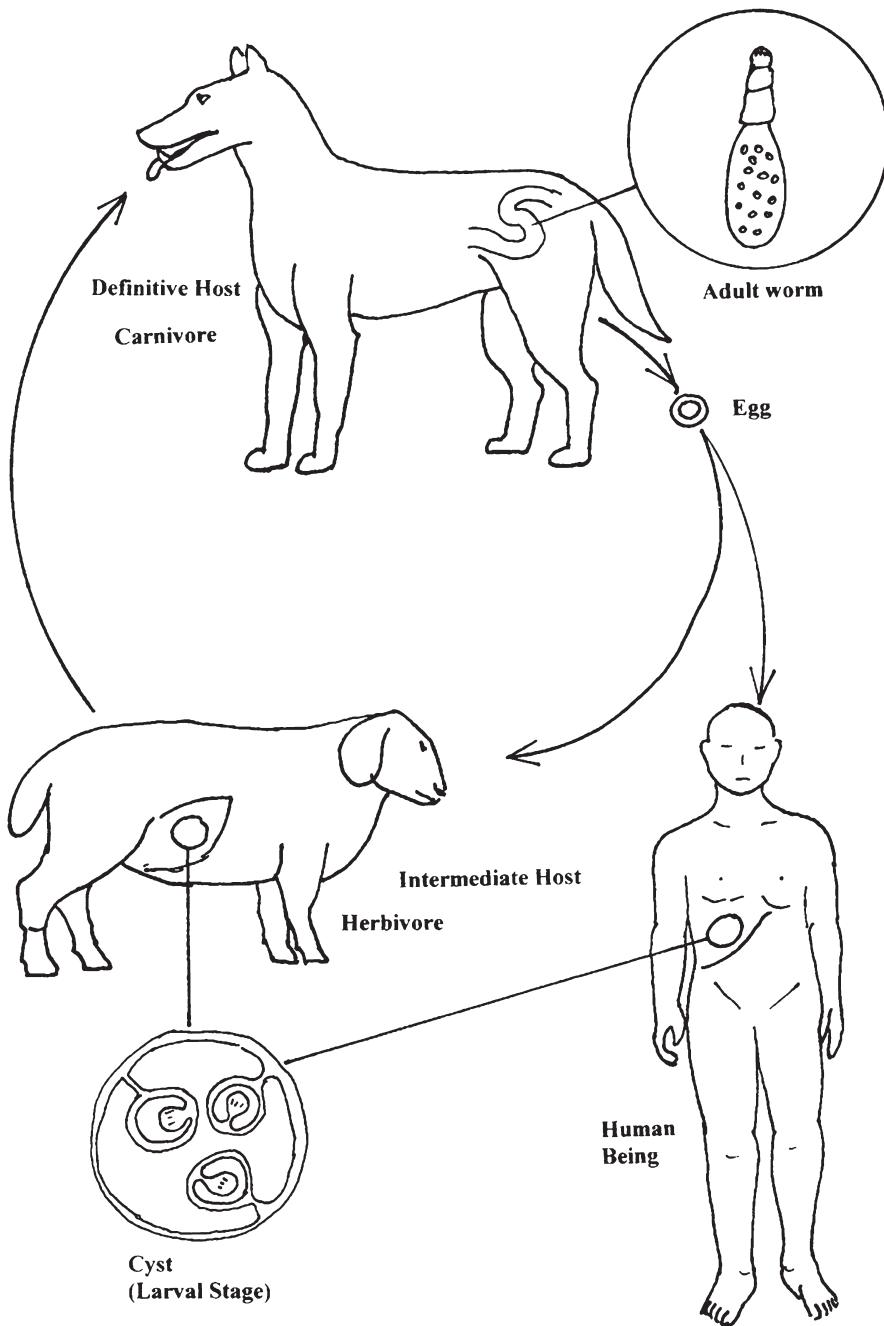


FIG 102.3. The life cycle of *Echinococcus granulosus*.

Hydatid disease is the larval stage of the small tapeworm *Echinococcus granulosus*. This worm has a carnivore (e.g. dog, wolf) as its definitive host and a herbivore (e.g. sheep, reindeer, camel) as the intermediate host. The larval stage forms the cyst. The life cycle is shown in the figure (Figure 102.3).

The adult worm lives in the intestine of the carnivore. Eggs are passed in its stool on to grass on which they are eaten by a herbivore and sometimes by a human being. The eggs hatch and the larvae enter the portal circulation and pass to the liver and sometimes on to the systemic circulation. The larvae form cysts that contain daughter larvae. These are infectious for the carnivore. The cysts may be multiple, and may become very large, causing disease by compression. Although usually located in the liver, they may also be present in lung, bone, or other viscera (Figure 102.4).

If a cyst ruptures, the contents form daughter cysts wherever the contents have spread to, such as throughout the peritoneal cavity. In addition, an anaphylactic reaction can occur.

Management consists of antimicrobial therapy with albendazole, followed, when necessary, by surgical removal. When surgery is performed, great care must be taken to prevent spilling of cyst contents and seeding of the field with larvae. Various techniques have been developed to minimize this risk. Prior antiparasitic treatment also reduces this risk.

This patient underwent surgical removal of the cyst. Microscopy of the cyst contents revealed hydatid “sand,” with larvae showing hooklets (Figure 102.5).

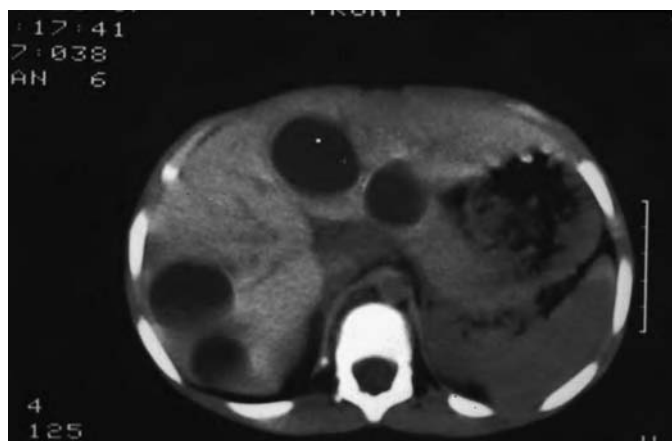


FIG 102.4. Computer tomography scan showing multiple hepatic hydatid cysts.

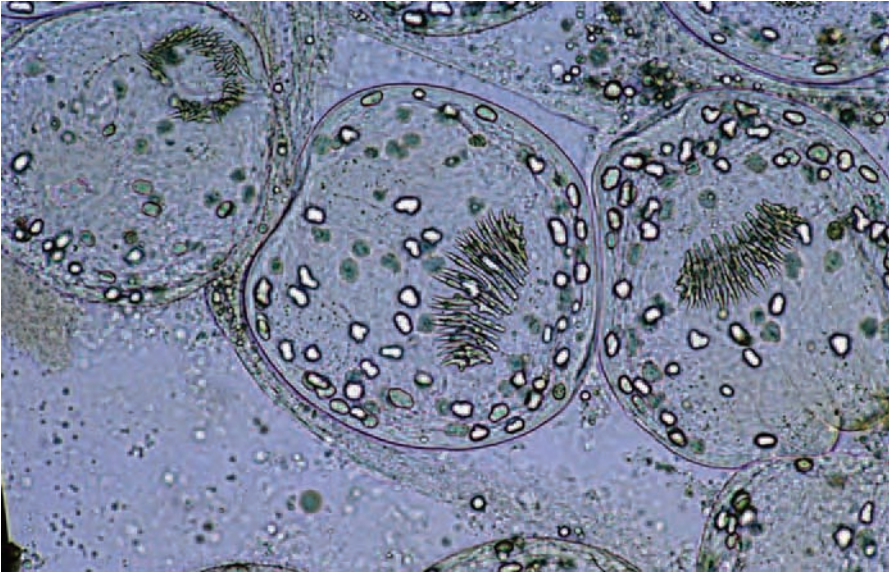


FIG 102.5. Contents of this patient's cyst, showing hydatid "sand." Note the row of hooklets on the larval segment.

Reading:

McManus DP, Zhang W, Li J et al: Echinococcosis. *Lancet* 2003; 362: 1295–1304.

CASE 103. A 9-year-old girl presents with a history of fever and joint pains for about 3 days and a rash for 1 day. On examination she is mildly ill-appearing, with a temperature of 38.7°C. She has pain on movement and swelling of her left ankle, and pain in her left knee. There is a macular rash on her limbs and trunk. The heart, lungs, abdomen, and nervous systems are normal, and there is no lymphadenopathy. The rash is shown in Figure 103.1.

- *What is your differential diagnosis?*
- *What would you like to know to help make a diagnosis?*

This patient has a clinical syndrome characterized by fever, arthritis, and a fairly diffuse macular rash. The differential diagnosis should include primarily infectious diseases but also rheumatologic diseases. In patients with diseases of unclear origin, knowledge about risk factors can be very helpful.



FIG 103.1. The child's rash.

About 1 week before onset of her symptoms this patient had acquired a pet rat. She often kept it inside her shirt, where she had several scratch marks. This made the diagnosis of rat bite fever very likely.

There are two forms of rat bite fever. (a) That caused by *Streptobacillus moniliformis*, a Gram-negative bacillus that can be transmitted by ingestion as well as by skin inoculation. The incubation period is usually 3–10 days. (b) That caused by *Spirillum minus* (Soduku) occurs mainly in Asia. Both these infections are characterized by fever, a macular or petechial rash present mainly on the extremities, and polyarthralgia or arthritis. The main complications are severe visceral infections, especially infective endocarditis. A series of three fatal cases has recently been reported in the United States. Treatment consists of penicillin.

This patient responded rapidly to intravenous penicillin.

■ **TAB. 103.1: Diseases associated with fever, rash, and arthritis and their risk factors.**

Infections characterized by diffuse rash and arthritis	Risk factor
Virus infections:	
arboviruses (including dengue, chikungunya, Onyong-nyong)	mosquitoes, travel
rubella	no immunization
parvovirus B19	contact with infected human
Bacterial infections:	
Septic arthritis associated with scarlet fever	<i>Streptococcus pyogenes</i> , <i>Staphylococcus aureus</i>
Meningococcal bacteremia	travel to area of high prevalence
Disseminated gonococcal infection	sexual activity
Infective endocarditis	heart disease
Leptospirosis	exposure to body of unclean water
Syphilis	sexual activity
Rat bite fever (<i>Streptobacillus moniliformis</i> , <i>Spirillum minus</i>)	rat exposure
Rocky Mountain spotted fever and other rickettsial diseases are associated with a rash similar to that observed in this patient, but are not associated with arthritis.	tick exposure
Rheumatological diseases to be considered are:	
systemic lupus erythematosus and rheumatoid arthritis	family history
juvenile rheumatoid disease	
acute rheumatic fever	prior pharyngitis

Reference:

CDC. Fatal rat-bite fever—Florida and Washington, 2003 MMWR 2005; 53: 1198–1202.

CASE 104. A 21-year-old girl presents in shock and with renal failure to a community hospital. She is resuscitated, placed on a ventilator, and transferred to a tertiary care facility. Prior to transfer she had blood cultures drawn and antimicrobial therapy initiated with ceftazidime, vancomycin, and metronidazole.

Her history is as follows: at the age of 4 years she underwent repair of an atrial septal defect. As a result of a complete heart block, she had an intraventricular pacemaker placed. About 4 months before the current admission she had the power source of her pacemaker replaced. This was complicated, 1 month later, by a pocket space infection, which necessitated removal of the pacemaker and its replacement with an epicardial pacemaker. On examination she is very ill, sedated, and on a ventilator, requiring

vasopressor therapy and continuous veno-venous hemofiltration. She has the following physical findings: subconjunctival hemorrhages, hemorrhagic skin lesions, hemorrhagic lesions on the tips of the fingers (Figure 104.1), splinter hemorrhages, purple discoloration of the distal third of the feet (Figure 104.2), and auscultation of her heart reveals a 3/6 holosystolic murmur at the apex.

- *What is the likely diagnosis?*
- *How might it have developed?*
- *How would you manage her?*

This patient has heart disease suggestive of mitral insufficiency and multiple conjunctival and cutaneous abnormalities indicative of emboli. These strongly suggest the diagnosis of infective endocarditis. The shock is probably due to a combination of cardiac failure and bacteremia (septic shock). The recent risk factor that might have caused the endocarditis is the pocket infection that could have led to infection of the intraventricular pacemaker. This is an entity called “pacemaker endocarditis.”

Management should entail resuscitation and ventilation, as is described above, followed by attempts to make a microbiological diagnosis. This entails performance of blood cultures. Three cultures done a few minutes apart should be performed, and antimicrobial therapy, directed against the



FIG 104.1. The skin lesions on the tips of her fingers.



FIG 104.2. The appearance of her foot.

most likely pathogens, should be initiated. The most likely pathogens causing infective endocarditis are *Staphylococcus aureus* and viridans group streptococci. Empiric therapy should include vancomycin (for its activity against methicillin-resistant staphylococci), nafcillin (for its rapid activity against methicillin-susceptible staphylococci), gentamicin (for its synergistic activity against streptococci and enterococci), and a third-generation cephalosporin (for its activity against many Gram-negative rods) all dosed according to her renal function. Additional information can be provided by obtaining culture results from her prior hospitalization for the drainage of the pocket infection. In this case the pocket infection had been caused by *Pseudomonas aeruginosa*, which was subsequently cultured from the patient's blood on this admission. It is very important that any pacemaker leads from the original pacemaker be removed.

She was treated with ceftazidime and tobramycin, and after a difficult course, during which she suffered a cerebral embolus, she recovered from the infection.

A more complete discussion of endocarditis follows.

Infective endocarditis (IE) is an infection of the valvular or mural endocardium. It occurs mainly in individuals who have underlying structural heart disease but can affect previously normal hearts.

Pathogenesis: The first stage in the development of IE is the formation of a thrombus on the endocardium. Predisposing factors to this are turbulence,

abnormal jets of blood, or trauma to the endocardium, for example, due to a catheter. If bacteremia is present at this time, the thrombus can become infected. This leads to accumulation of platelets and leukocytes at the site, within which are embedded the bacteria. This mass is called a vegetation (Figure 104.3).

The infection can result in local tissue injury, for example, valve destruction, and can spread to adjacent structures. Abnormal jets caused by the infection can result in infection in noncontiguous structures in the heart (e.g. spread from the aortic to the mitral valve). Pieces of vegetation can break off, resulting in emboli. In right-sided endocarditis, these usually impact in the lung, whereas in left-sided disease, they may impact anywhere in the systemic circulation.

Valvular endocarditis can result in valvular insufficiency or, if there is a large vegetation, in valvular obstruction.

Microbiology: The most common microorganisms causing IE are those that stick readily to the endocardium, namely staphylococci and viridans group streptococci. In native heart endocarditis (i.e. cases in which there is no underlying heart abnormality or prosthetic material) *Staphylococcus aureus* accounts for almost all cases of staphylococcal endocarditis. However,

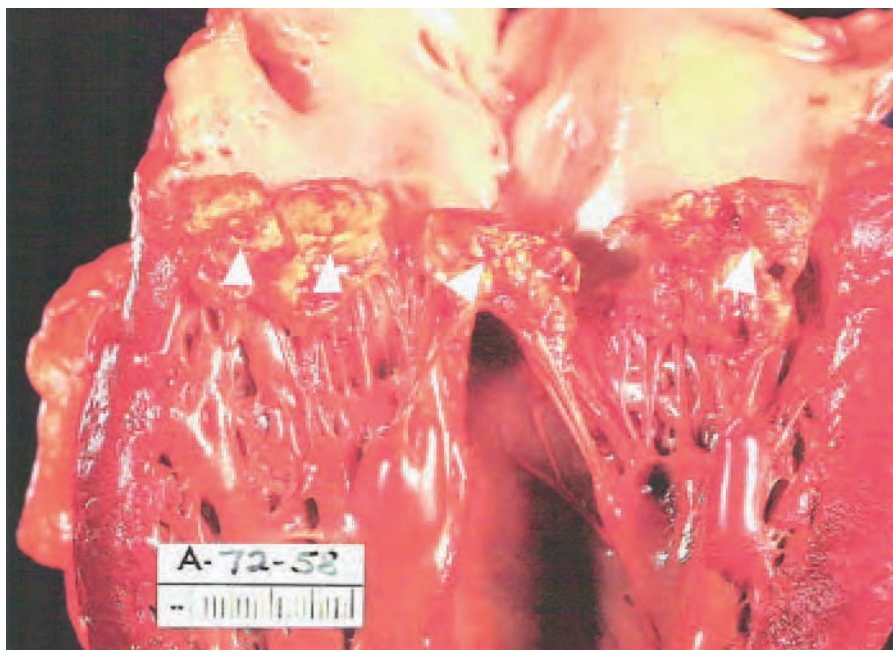


FIG 104.3. Vegetations (arrows) on the mitral valve of a different patient with infective endocarditis. (Courtesy of the Centers for Disease Control and Prevention/Dr Edwin P Ewing, Jr)

after heart surgery, coagulase-negative staphylococci are also important. Many cases of postsurgical endocarditis are nosocomial. In this circumstance *Candida* spp. and *Corynebacterium* spp., as well as staphylococci, are important. Gram-negative rods are important causes of nosocomial bacteremia in general, but less common causes of endocarditis. The “HACEK” group of organisms (*Haemophilus aphrophilus*, *Actinobacter actinomycetemcomitans*, *Cardiobacterium hominis*, *Eikenella corrodens*, and *Kingella kingae*) are rare causes of endocarditis, as are *Coxiella burnetii* and *Bartonella* spp.

Predisposing conditions: In infants and children the main underlying diseases predisposing to endocarditis are various congenital heart diseases. Because most predisposing conditions are either left-to-right shunts (ventricular septal defect, patent ductus arteriosus) or right-sided lesions (pulmonary stenosis, Fallot’s tetralogy), most IE in children is right-sided. In countries where rheumatic heart disease is prevalent, this is also an important predisposing condition. Nosocomial bacteremia, particularly with *Staphylococcus aureus* and *Candida* spp., can lead to endocarditis, even in patients with normal hearts. An increasing number of cases of IE are associated with the presence of central venous catheters, which can remain in the superior vena cava or right atrium for prolonged periods.

Clinical manifestations: (a) General (systemic) symptoms and signs: fever is the most common and important feature of IE. Fatigue, malaise, digital clubbing, anemia, weight loss, and arthralgia may also occur. (b) Embolic and immune phenomena: splinter hemorrhages, Janeway lesions and Osler’s nodes on the fingers, Roth spots in the retina (hemorrhages with a white center), petechiae, missing pulses, hematuria (due to immune-mediated glomerulonephritis), splenomegaly, and mental status changes. (c) Cardiac manifestations: the predisposing condition, changing or new heart murmur, chest pain due to coronary emboli, and heart failure.

Nowadays fewer cases are diagnosed after prolonged illness, when many of the classic signs of “subacute bacterial endocarditis (SBE)” would have developed, and more are diagnosed early. There are two main clinical syndromes: (1) acute infective endocarditis, usually caused by *Staphylococcus aureus*, in which the predominant manifestations are those of the sepsis syndrome, with hemodynamic instability, and (2) a less virulent form, in which prolonged fever occurs in an at-risk individual.

Differential diagnosis: As the manifestation of IE are protean, the differential diagnosis is broad. However, it generally includes conditions that are associated with fever and heart murmurs. They are shown in Table 104.1.

The diagnosis of IE is very important because, without treatment, the condition is almost universally fatal. On the other hand, treatment entails intravenous therapy for several weeks, which is undesirable in an individual

TAB. 104.1: Differential diagnosis of infective endocarditis.

Fever and heart murmur
Congenital heart disease + viral infection
Flow murmur + infection
Heart murmur, arthralgia/arthritis
Systemic lupus erythematosus
Juvenile rheumatoid disease
Acute rheumatic fever
Postcardiac surgery bacteremia
Vascular catheter-associated
Pneumonia-associated
Wound-associated
Urinary catheter associated
Other Postcardiac surgery fever
Postpericardiotomy syndrome
Post-bypass infectious mononucleosis (postperfusion syndrome)
Heart murmur and emboli
Atrial myxoma

without the condition. The main diagnostic test is blood culture. Three separate blood cultures should be taken, with a minimum of 5 ml of blood in each bottle. This is to demonstrate continuous bacteremia and to confirm or reject the hypothesis that a single positive culture is due to a contaminant (such as coagulase-negative staphylococcus). Echocardiography is useful to assist in the diagnosis, especially to demonstrate the location of the lesion and the extent of tissue damage that it has caused. A set of diagnostic criteria have been established in adults, which have been shown to be useful in children. These are known as the “Duke” criteria.

Management: The mainstay of management is antimicrobial. The role of surgery and prophylaxis is discussed later.

The main decision to make is to treat or not to treat, depending on how certain the diagnosis is. The disadvantage of treating a patient in whom the diagnosis is unclear is that once therapy has been instituted, if the blood cultures are subsequently negative, further blood culture information will not become available, and, in the absence of an alternative diagnosis, the course of therapy should be completed. Therefore, from a management viewpoint, I consider patients in three different categories. (a) Patients with suspected acute endocarditis. These patients should have three sets of blood cultures performed over a period of about 15 minutes, and therapy should then be started. (b) Patients with many features of endocarditis, whom one would treat irrespective of the blood culture results. These patients should have three sets of blood cultures performed over several hours, and then

therapy should be started. (c) Patients who might have endocarditis (possible). These patients should have three sets of blood cultures over 24 hours but should NOT receive antimicrobial therapy. Their clinical course and blood culture results should be followed until a diagnosis of endocarditis or an alternative diagnosis can be made.

Choice of antimicrobial therapy: This should be bactericidal. Empiric therapy should be active against the likely pathogens (taking into account their likely resistance patterns). Once the pathogen and its resistance pattern becomes known, therapy can be adjusted. My initial empiric therapy is directed at *Staphylococcus aureus* that might be resistant to methicillin, streptococci (including enterococci), which might be relatively resistant to penicillin, and Gram-negative rods. Therefore I use a combination of vancomycin, nafcillin (optimal for methicillin-susceptible *Staphylococcus aureus* infection), ceftriaxone, and low-dose gentamicin.

Complication of IE: (a) Cardiac complications: The most important complication of IE is heart failure, which can lead to shock. Intractable heart failure is the main indication for surgery in patients with IE. It is usually due to valvular insufficiency but can be due to obstruction of a valve aperture caused by a massive vegetation. IE of the aortic valve can be associated with many different complications, mainly as a result of infection spreading through the valve ring. This can result in the following: spread to the conducting system, resulting in heart block; rupture into the pericardial space, resulting in tamponade; extension into the coronary sinus; extension to the mitral valve. (b) Emboli: These can occur anywhere. The most serious systemic emboli are those to the brain, resulting in a stroke; however, they can shoot to the heart itself, gut, kidney, or limbs, resulting in a threat to the function of those organs. Emboli from right-sided endocarditis usually lodge in the lungs.

Role of surgery in management: When a patient with IE has intractable heart failure, cardiogenic shock, or persistent bacteremia (at least 1 week's duration) despite optimal antimicrobial therapy, a cardiac surgeon should be consulted with a view to surgical removal of the infected tissue and replacement with prosthetic material.

Prevention of IE: Most patients with IE do not have a history of a preceding predisposing event (other than the cases following heart surgery). Nevertheless those at risk of IE should receive antibiotic prophylaxis when undergoing procedures that are associated with transient bacteremia. The benefit of this has not been shown in randomized controlled trials (it would require many thousands of cases to be studied to show a benefit). The recommendations of the American Heart Association regarding the types of underlying heart abnormalities, and types of procedures for which

prophylaxis is recommended have been published. In patients with rheumatic heart disease, the prophylaxis against *Streptococcus pyogenes* should not be confused with prophylaxis against IE.

Reading:

Moreillon P: Infective endocarditis. *Lancet* 2004; 363: 139–149.

Durack DT, Lukes AS, Bright DK, Duke Endocarditis Service: New criteria for diagnosis of infective endocarditis: utilization of specific echocardiographic findings. *Am J Med* 1994; 96: 200–209.

Dajani AS, Taubert KA, Wilson W et al: Prevention of bacterial endocarditis. Recommendations by the American Heart Association. *JAMA* 1997; 277: 1794–1801.

CASE 105 (COMP). A 17-year-old boy develops fever, rash, and a severe headache 1 week after a safari in South Africa. On examination he is moderately ill-appearing with a temperature of 39.5°C; the other vital signs are normal. He has mild photophobia and mild conjunctival hyperemia but no neck stiffness. There is a macular rash on his trunk and limbs, and two eschars on his lower abdomen (Figures 105.1 and 105.2), and there is bilateral inguinal lymphadenopathy. The rest of his examination is normal.

- *What is the differential diagnosis?*
- *How would you manage him?*

The combination of a rash, fever, and headache suggest a systemic infection. The eschars suggest further that these sites might have been the inoculation site of an infectious agent, as a result of an injury or arthropod bite. An injury leading to focal and then systemic infection is likely due to *Staphylococcus aureus* or *Streptococcus pyogenes*. However, in that case the inoculation site would have more evidence of inflammation or suppuration than is present here. Furthermore the rash is not scarlatiniform as can occur with these infections and is too diffuse for embolic disease due to infective endocarditis. Therefore one should consider infection introduced by an arthropod. Many mosquito-borne virus infections (arbovirus infections) such as dengue and West Nile virus infection are associated with headache and rash, but they are not associated with eschars. *Trypanosoma brucei*, the cause of African sleeping sickness, which is transmitted by a Tsetse fly, is associated with an eschar at the bite site, fever, and rash, and generalized lymphadenopathy but is not transmitted in South Africa. This presentation is typical of



FIG 105.1. *The rash and lesions on the patient's trunk.*

certain tick-borne rickettsial infections, namely African tick bite fever, caused by *Rickettsia africae*, and Mediterranean spotted fever, caused by *Rickettsia conorii*.

As in the case of Rocky Mountain spotted fever, the decision to provide antimicrobial therapy (doxycycline) for patients suspected of having these infections should be based on clinical evaluation, not on any laboratory study.

The spotted fever rickettsiae have a worldwide distribution, with each geographic area having its particular species. They constitute an important and under-appreciated cause of febrile illness in travelers. They infect endothelial cells and consequently can affect any organ. Although African tick bite fever (African tick typhus) does not have the high case fatality rate associated with Rocky Mountain spotted fever, it can be fatal, the highest risk groups being infants and the elderly.



FIG 105.2. *The rash on one of his limbs.*

This patient was treated with doxycycline and he made a rapid recovery.

Reading:

Jensenius M, Fournier P-E, Kelly P et al: African tick bite fever. *Lancet Infect Dis* 2003; 3: 557–564.

Raoult D, Fournier PE, Fenollar F et al: *Rickettsia africae*, a tick-borne pathogen in travelers to sub-Saharan Africa. *N Engl J Med* 2001; 344: 1504–1510.

CASE 106. A 10-year-old girl presents with a skin abnormality of several weeks' duration. It is not itchy and is only of esthetic concern to her. On examination it appears as seen in Figure 106.1. There are multiple areas of hypopigmentation with slight very superficial scaliness. The lesions have normal sensation.

- *What is the diagnosis?*
- *How can you confirm it?*
- *How would you treat the patient?*

This is the typical appearance of Pityriasis versicolor, which is a very superficial fungal infection caused by *Malassezia furfur*. This yeast can be part of the normal flora of the skin and causes lesions, as seen in the picture, mainly in adolescents, although it may affect younger children and adults. The infection usually affects the shoulders, neck, and upper trunk. The diagnosis can be confirmed by microscopic examination of a skin scraping stained with methylene blue stain or after clearing with potassium hydroxide. This shows round yeast cells and hyphae, the so called “spaghetti-and-meatballs” (Figure 106.2).

The treatment consists of selenium sulfide shampoo, applied for 10 minutes to the affected skin, daily for 1 week then weekly for 1 month.

This organism can cause invasive infection in individuals receiving total parenteral nutrition that includes a lipid preparation. The organism requires



FIG 106.1. The child's rash.

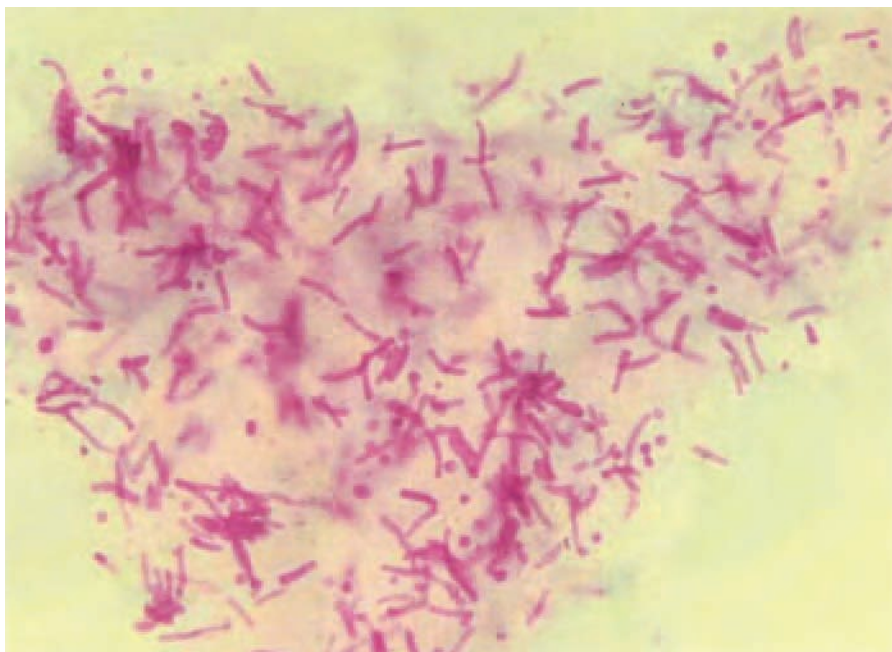


FIG 106.2. A photomicrograph of *Malassezia furfur*. (Courtesy of Dr Lucille K Georg/Centers for Disease Control and Prevention)

lipid for its growth. One of the methods used in the laboratory for identifying it is demonstration of growth in the presence of lipid, for example, olive oil, but no growth in its absence.

CASE 107. A 5-year-old girl presents with a history of having recurrent episodes of headache, abdominal pain, and stuffy nose, followed within a few hours by the onset of fever. This is associated with swelling of the lymph nodes in the neck and often with mouth ulcers. These episodes, which last about 4 days, occur every 4–6 weeks and have occurred her “whole life.” She has been exposed to dogs, cats, hamsters, tropical fish, and ticks. No other family member has had similar symptoms. On examination she is well appearing and afebrile, and the only abnormal findings are enlarged, tender upper cervical lymph nodes, large tonsils, and a slightly enlarged spleen.

- *What is the differential diagnosis?*
- *How should she be managed?*

This child’s current clinical findings suggest an infectious mononucleosis-like illness. This is caused by Epstein–Barr virus, cytomegalovirus, human

immunodeficiency virus, and toxoplasmosis. These conditions do not, however, cause recurrent episodes of febrile illness over years. This child's symptoms suggest a "periodic fever" syndrome. This is a group of disorders characterized by recurrent episodes of fever without evidence of an infectious etiology. Some of them are inherited disorders of proteins involved in inflammation. The most important of these is Familial Mediterranean fever (FMF), which is characterized by recurrent episodes of fever, arthritis, and serositis. Affected individuals, who have ancestry from the eastern Mediterranean region, can develop amyloidosis. The cause is an abnormality of "pyrin," a protein involved in the inflammatory cascade. The treatment is colchicine. Other inherited disorders in this group are Muckle-Wells syndrome, also due to an abnormality of pyrin, Hibernian fever, due to an abnormality of the tumor necrosis factor receptor, and the Hyper IgD syndrome, due to a mutation of the enzyme mevalonate kinase.

This child's illness is highly suggested of a syndrome called the "Periodic Fever, Aphthous stomatitis, Pharyngitis, Cervical Adenitis" (PFAPA) syndrome. It does not appear to have serious sequelae, and there is no specific therapy.

Reading:

Feder, H: Periodic fever, aphthous stomatitis, pharyngitis, adenitis: a clinical review of a new syndrome. *Curr Opin Pediatr* 2000; 12: 252–256.

Scholl, P: Periodic fever syndromes. *Curr Opin Pediatr*. 2000; 12: 563–566.

Edwards MS, Millon JC, Perez MD: Recurrent fever in a healthy-appearing child. *Semin Pediatr Infect Dis* 2004; 15: 220, 229–234.

CASE 108. A 14-year-old previously healthy boy presents with a history of progressively worse pain in the left hip developing over the past 1 week, and fever for 3 days. He jumped off a porch 1 week ago but did not injure himself. On examination he is moderately ill-appearing with a temperature of 39.8°C, heart rate of 132 beats per minute, blood pressure of 111/64 mm Hg, and respiratory rate of 20 per minute. The most notable finding is warmth and tenderness over the left gluteal region and greater femoral trochanter area, and limitation and pain on rotation of the hip. Flexion and extension of the hip are fairly normal. There is a healed abrasion on the posterior aspect of the left thigh. There is mild conjunctival injection, and the heart examination is normal except for a 2/6 ejection systolic murmur. The rest of the examination is normal, in particular there are no enlargement of the liver or spleen, no lymphadenopathy, and no other skin lesions.

- *Where might the illness be located?*
- *What might its cause be?*
- *What would you do?*

In patients with pain around the hip, one should consider disease within the hip joint, the bones around the hip joint (pelvic bones, sacrum, and femora), the bones and soft tissue of the lower back, disease within the contents of the pelvic cavity (intestine, genital tract, urinary bladder, and muscles, including the obturator internus, piriformis, gemelli, coccygeus, pubococcygeus, and levator ani), and disease affecting nerves around the hip.

Disease within the hip joint is very important to diagnose early because raised pressure within the joint from inflammation can lead to compromise of the vascular supply to the head of the femur and subsequent avascular necrosis. The ability of the patient to flex and extend the hip but not rotate it suggests that the disease is not within the hip joint itself, while tenderness over the gluteal region suggests the possibility of disease within the gluteal muscles.

Given the patient's fever, the most likely cause of the disease is an infection. The presence of a healed abrasion on the thigh suggests that the likely organism is *Staphylococcus aureus* or *Streptococcus pyogenes*. In addition to the local infection he could be bacteremic and at risk for the hemodynamic effects of bacteremia and for dissemination of the infection.

A specific clinical examination that should be performed in this situation is a rectal examination. This is the only clinical access available to examine the inside of the pelvis and the pelvic bones in males and prepubertal females. In this patient it revealed a boggy swelling anterior to the sacrum, suggesting that the problem is a pelvic abscess.

The clinical situation poses the following dilemma: should one immediately initiate antimicrobial therapy active against the most likely organisms in this moderately ill patient or should one first try to delineate the focus of disease by imaging, with the hope that this will enable a surgeon to aspirate or drain the infected material, after which antimicrobial therapy would be started?

In this child a blood culture was performed and antimicrobial therapy was initiated, directed against staphylococci, including methicillin-resistant *Staphylococcus aureus*, *Streptococcus pyogenes*, and enteric rods, with vancomycin, nafcillin, and cefotaxime. A magnetic resonance imaging (MRI) scan of the pelvis is shown in Figure 108.1.

This showed a large pelvic abscess compressing the rectum. There was also associated swelling around the posterior branch of the first sacral nerve root, which might have accounted for some of his pain. Although surgical



FIG 108.1. An MRI scan of the pelvis showing a large abscess compressing the rectum.

drainage was considered, the inaccessibility of the abscess led to a delay in performance of such a procedure.

The first blood culture grew out *Staphylococcus aureus*, susceptible to penicillin; the blood culture taken 2 days after initiation of therapy, grew out the same organism but there were a few colonies that were methicillin resistant. He remained febrile, and a blood culture taken 3 days after initiation of therapy grew out a methicillin-resistant *Staphylococcus aureus*. He then underwent surgical drainage of the abscess through the rectum. The pus grew out methicillin-resistant *Staphylococcus aureus*. After the drainage he became afebrile and made a speedy recovery.

How did this abscess develop? Possibly he sustained an injury to one of the pelvic muscles when he jumped off the porch, causing a muscle hematoma. This might have become seeded during a transient staphylococcal bacteremia arising from the thigh abrasion.

This patient demonstrates the following:

- (a) pain around the hip may be due to disease inside the pelvis;
- (b) a rectal examination can help to locate disease around the hip;
- (c) antimicrobial therapy may not be effective if pus is not drained;
- (d) antibiotic resistance can emerge within a patient during antimicrobial therapy, due to natural selection of resistant organisms.

Staphylococcus aureus inhabits the skin and the anterior nares. It is an opportunist in that minor skin trauma may be followed by local infections. These include impetigo (pyoderma) and furuncles, which may progress to cellulitis and abscesses, and surgical wound infections. These foci can be associated with asymptomatic bacteremia. This bacteremia can, however, result in metastatic infections that can be severe, and, in turn, lead to severe symptomatic bacteremia. These foci include bone (osteomyelitis), joint (septic arthritis), muscle (pyomyositis), lung (hematogenous pneumonia), and endocardium (infective endocarditis). Recently a syndrome of severe sepsis, associated with pneumonia and sometimes deep venous thrombosis, has been described, affecting particularly adolescent boys. This is caused by a community-acquired methicillin-resistant strain or strains of *Staphylococcus aureus* that produce a virulence factor called Pantin-Valentine leukocidin (pvl).

Although the above-described patient did not have manifestations of this syndrome, he was treated aggressively on presentation with the concern that this might develop.

Reading:

Crawford SE, Daum RS: Epidemic community-associated methicillin-resistant *Staphylococcus aureus*. Modern times for an ancient pathogen. *Pediatr Infect Dis J* 2005; 24: 459–460.

Gonzalez BE, Martinez-Aguilar G, Hulten KG et al: Severe staphylococcal sepsis in adolescents in the era of community-acquired methicillin-resistant *Staphylococcus aureus*. *Pediatrics* 2005; 115: 642–648.

Zetola N, Nuermberger EI, Bishai WR: Community-acquired methicillin-resistant *Staphylococcus aureus*: an emerging threat. *Lancet Infect Dis* 2005; 5: 275–286.

CASE 109. A 10-year-old boy, previously healthy, was diagnosed with pneumococcal meningitis caused by a fully susceptible strain of *Streptococcus pneumoniae*. He was treated with intravenous ceftriaxone and steadily improved. On the eighth day of therapy he complains of severe right upper quadrant pain and nausea. He lacks a cough or other respiratory tract symptoms. Examination is normal except for right upper quadrant tenderness. Of note is the fact that he is not jaundiced.

- *What might explain his problem?*
- *What additional information would you like?*
- *What would you do?*



FIG 109.1. An ultrasound picture of the gallbladder showing a large amount of sludge within the lumen.

The sites of disease most likely to cause the above symptoms are the liver, gallbladder, and lower lobe of the right lung. The lack of respiratory symptoms makes lung disease unlikely. A specific risk factor for disease of the gallbladder is ceftriaxone, 50% of which is excreted in bile. It can cause the formation of “sludge” in the gallbladder where it is concentrated up to 120 times the serum concentration. The additional information of interest is the dosage of this drug that he received, which was 2 g every 12 hours. His weight was 45 kg. This dosage is the highest dose recommended for adults and was probably too high for him. An appropriate dose might have been 1.5 g every 12 hours. The presence of biliary sludge can be demonstrated by ultrasound. An ultrasound (Figure 109.1) revealed a large amount of sludge in the gallbladder.

Management should entail discontinuing the drug if appropriate or lowering its dosage. If symptoms persist or become worse, cholecystectomy might become necessary.

He improved initially after ceftriaxone therapy was discontinued. However, he presented a few weeks later with severe epigastric pain and was found to have acute pancreatitis. After this had resolved, he underwent cholecystectomy.

Reading:

Ko C, Sekijima JH, Sum P: Biliary sludge. *Ann Intern Med* 1999; 130: 301–311

CASE 110. A 4-year-old boy, previously diagnosed with autoimmune lymphoproliferative syndrome (ALPS) and being treated with systemic corticosteroids, presents with a history of fever and cough for a few days. There is no vomiting, diarrhea, dysuria, or bone pain, and he has not traveled outside his home town in Georgia, USA. On examination he is miserable and difficult to examine but not severely ill. His temperature is 39°C, and his vital signs are normal other than a heart rate of 120/minute. There is no pallor or jaundice. He has enlargement of cervical, axillary, and inguinal lymph nodes and a slightly enlarged spleen. After multiple attempts, his throat is visualized. This reveals inflamed tonsils covered with extensive exudates. The rest of his examination is normal.

- *What is your differential diagnosis?*
- *What would you do?*

This child is immunocompromised by virtue of receiving corticosteroids and possibly by his underlying disease itself. This places him at increased risk for severe viral infections. His only localizing finding is an exudative tonsillitis. This is the likely cause of his symptoms. This, together with the generalized lymphadenopathy and splenic enlargement suggests the diagnosis of infectious mononucleosis. However, the splenic enlargement could be due to his underlying disorder.

Other causes of tonsillitis are *Streptococcus pyogenes*, *Corynebacterium diphtheriae* (to which he has no known exposure risk), and several respiratory viruses, including adenovirus. *Arcanobacterium haemolyticum* infection of the pharynx is associated with a diffuse scarlatiniform rash, which this child lacks.

Therefore a test for streptococcal pharyngitis should be performed. If positive he should be treated with penicillin. If negative, specific tests for viruses, by antigen detection or by culture, should be considered. In most patients these are not of value because they do not influence therapy. However, in a child such as this, a positive test might restrain one from pursuing other diagnostic tests and from using antibiotics.

In this child, his fever persisted for several days. A culture from his throat grew out adenovirus.

There are more than 40 different serotypes of adenoviruses, which are large DNA viruses. Most of these cause mucosal disease, particularly of the respiratory tract and eye, but also of the urinary bladder. Specific serotypes, types 40 and 41, are well-recognized causes of acute gastroenteritis. Adenoviruses occasionally cause visceral disease such as hepatitis and severe pneumonia in normal hosts. However, they constitute a very important cause of

severe and sometimes fatal pneumonia and hepatitis in immunocompromised hosts, in particular transplant patients.

Although antiviral therapy has not been clearly shown to be of benefit to affected patients, there are anecdotal reports of improvement in patients treated with cidofovir, which is a very toxic drug.

This child gradually improved without receiving any antiviral therapy.

Reading:

Leen AM, Looney CM: Adenovirus as an emerging pathogen in immunocompromised patients. *Bri J Haematol* 2004; 128: 135–144.

Ison MG: Adenovirus infection in transplant recipients. *Clin Infect Dis* 2006; 43: 331–339.

CASE 111. A 16-year-old boy presents with a 4-day history of fever, headache, generalized muscle aches, and weakness. He has also had odynophagia and upper abdominal pain. On examination he has a temperature of 38.4°C, heart rate of 93/minute, and blood pressure of 98/53. His conjunctivae are mildly inflamed, but the rest of his mucosae are normal. His abdomen is mildly tender with slight hepatomegaly but no splenomegaly. There are enlarged axillary, cervical, and inguinal lymph nodes. His strength is generally reduced and he has diffuse muscle tenderness. The rest of his examination is normal.

- *What is your differential diagnosis?*
- *What more would you like to know?*
- *What would you like to do?*

This patient clearly has a systemic disease associated with fever. An infectious etiology is most likely. The lymphadenopathy suggests an infectious mononucleosis-like illness. The differential diagnosis includes:

Viral infections

Epstein–Barr virus (EBV)
 cytomegalovirus (CMV)
 human immunodeficiency virus (HIV)
 hepatitis A, B or C

Bacterial infections

rickettsial or ehrlichial infection
 syphilis (secondary)
 leptospirosis

Parasitic infection

toxoplasmosis

Noninfectious etiologies

systemic lupus erythematosus

leukemia

It is very important to enquire from him about possible exposures to these agents, in particular travel and recreational activities, including sexual activity. Laboratory studies should be used both to define the extent of organ system abnormalities and to determine possible etiologies, both for treatment and for public health reasons.

Such studies in his case revealed elevated hepatic transaminases (alanine aminotransferase 661 U and aspartate aminotransferase 935 U), indicating hepatitis, and elevated creatine phosphokinase (660 U), indicating myositis.

Serological studies revealed that he had evidence of a past infection with EBV and that he had a negative antibody test for HIV and *Toxoplasma gondii*.

Negative antibody studies do not exclude the diagnosis of acute (primary) HIV infection, because seroconversion can take 3–6 months from the time of infection. Therefore direct tests of viral presence should be performed. This generally means viral nucleic acid detection, by a polymerase chain reaction test, which was positive in his case. This is, in fact, a typical presentation of acute HIV infection. This occurs in about 70% of cases of horizontally acquired HIV infection. The symptoms and signs include fever and fatigue (almost all patients), night sweats, myalgia, arthralgia, nausea, vomiting, pharyngitis, lymphadenopathy, enlargement of liver or spleen, rash, oral or genital ulcers, oral thrush, headache, meningitis, myelitis, encephalopathy, peripheral neuropathy, and “cotton-wool” retinal spots.

The diagnosis should be considered in individuals who are at risk for HIV infection, and who have features of an infectious mononucleosis-like illness.

The importance of making a diagnosis is for long-term management and public health management, including counseling the patient about his risk of transmitting the infection to others and following-up his sexual contacts. The overall benefit of antiretroviral therapy in such patients is controversial.

Reading:

Schacker T, Collier AC, Hughes J et al: Clinical and epidemiologic features of primary HIV infection. *Ann Intern Med* 1996; 125: 257–264.

Kahn JO, Walker BD: Acute human immunodeficiency virus type 1 infection. *N Engl J Med* 1998; 339: 33–39.

Perlmutter BL, Glaser JB, Oyugi SO: How to recognize and treat acute HIV syndrome. *American Family Physician* 1999; 60: 535–546.

CASE 112. A 4-year-old boy complains of sudden severe pain in his left eye while playing outside. At an initial evaluation, he is diagnosed with a corneal abrasion, but on the following day he has periorbital edema, proptosis, ophthalmoplegia, and markedly decreased visual acuity. There is a vitreous opacity, indicating a diagnosis of endophthalmitis. An ultrasound indicates the presence of a metallic foreign body in the eye. A surgical exploration of the eye is performed during which large necrotic areas of sclera and retina are noted. Therefore the eye is enucleated. Histology of the eye is shown in Figure 112.1, as is a Gram stain of the excised material (Figure 112.2).

- *What is the likely organism?*
- *Why did it cause such rapid destruction of the eye?*

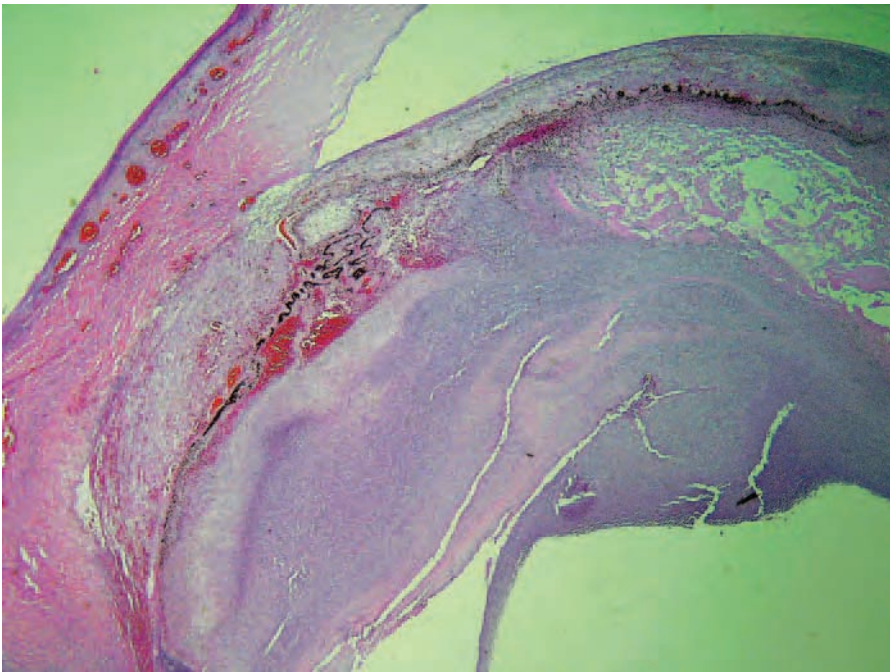


FIG 112.1. An abscess behind the iris ($\times 100$). (Courtesy of Dr Hans Grossniklaus, Department of Ophthalmology, Emory University)

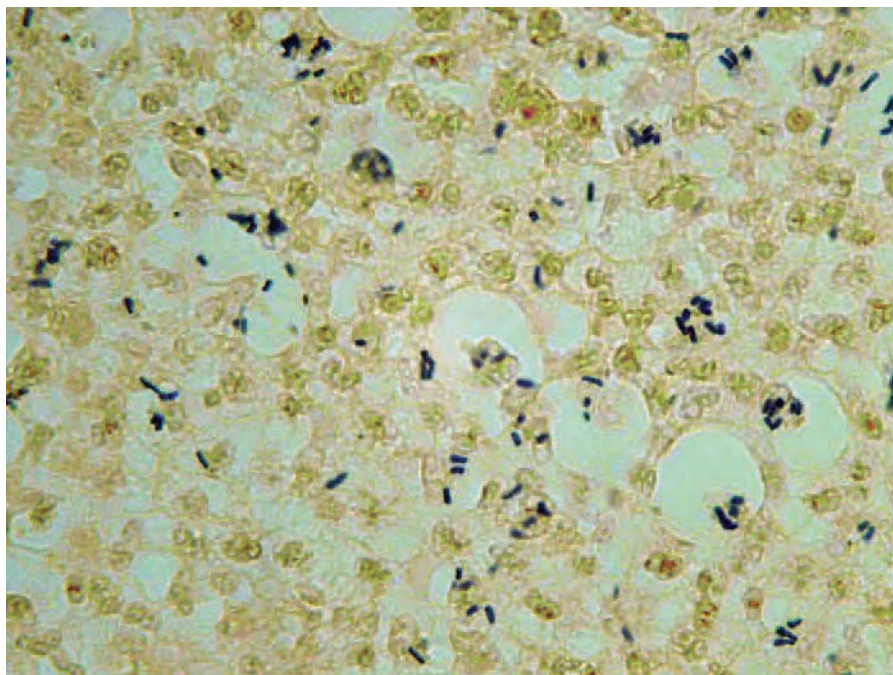


FIG 112.2. A Gram-stained preparation of the contents of the intraocular abscess ($\times 250$). (Courtesy of Dr Hans Grossniklaus, Department of Ophthalmology, Emory University)

The organisms seen are large, fat, Gram-positive bacilli. This suggests a *Bacillus* spp. or a *Clostridium* spp. infection. (The culture subsequently grew out *Bacillus cereus*.) Both these organisms can cause infections complicating injuries, which must have occurred in this child, to explain the foreign body within his eye. This scenario is well described as being caused by *Bacillus cereus*, which is a very important cause of infection complicating penetrating eye injuries. This organism produces several exotoxins, including a lecithinase, which probably plays a role in the ocular damage. Endophthalmitis (inflammation of the intraocular structures) results from penetrating injury of the eye by trauma or surgery (such as cataract surgery) or from bloodstream infection with bacteria or fungi. Systemic antimicrobial therapy is not very effective for treatment of patients with this condition. Optimal therapy entails vitrectomy and instillation of antimicrobial agents, in particular vancomycin and amikacin, directly into the vitreous.

Bacillus cereus also causes other infections. In normal hosts it causes two acute food poisoning syndromes, namely an emetic syndrome and a diarrheal syndrome. The organism causes infections of vascular catheters and other indwelling devices, and, in severely compromised hosts, can cause bacteremia and other severe infections.

Reading:

David DB, Kirkby GR, Noble BA: *Bacillus cereus* endophthalmitis. Br J Ophthalmol 1994; 78: 577–580.

Gaur A, Shenep JL: The expanding spectrum of disease caused by *Bacillus cereus*. Pediatr Infect Dis J 2001; 20: 533–534.

CASE 113. A 12-year-old boy with nephrotic syndrome for several years presents with a 1-day history of fever and severe abdominal pain. He always has some edema and ascites. Treatment for his nephrotic syndrome consists of a low salt diet and prednisone. He requires infusions of albumin followed by furosemide when his ascites causes discomfort. There is no history of diarrhea or cough. On examination he is very ill-appearing and grunting. His heart rate is 120/minute, blood pressure 100/60 mm Hg, respiratory rate 40/minute, temperature 38.7°C, his peripheral perfusion is good, and he has marked anasarca. His lung examination reveals dullness to percussion and decreased breath sounds at both bases. His abdomen is distended and diffusely very tender. The rest of his examination is normal.

- *What is your differential diagnosis?*
- *How would you manage him?*

The child's nephrotic syndrome places him at risk for systemic infections, particularly those caused by *Streptococcus pneumoniae*, and for thromboses. In addition the corticosteroid therapy predisposes him to opportunistic infections, such as those caused by cytomegalovirus and *Pneumocystis jiroveci*. Individuals with ascites, whether due to nephrotic syndrome or liver disease, are at risk for the development of spontaneous bacterial peritonitis due to hematogenous spread of bacteria. In patients with nephrotic syndrome such infections are caused mainly by *Streptococcus pneumoniae*, other streptococci, and by enteric Gram-negative bacilli. Systemic infections are an important cause of morbidity and mortality in patients with nephritic syndrome, probably due to loss of immunoglobulin and complement through the urine.

His clinical picture strongly suggests an intraperitoneal infection and sepsis syndrome. Although peritonitis from a perforated bowel, such as the appendix, is possible, spontaneous bacterial peritonitis is much more likely in this case.

Management should consist of the following:

1. Ensuring adequate perfusion – Although he is edematous, his intravascular volume is likely depleted. An infusion of intravenous albumin would therefore be indicated.

2. Obtaining cultures of blood and the peritoneal fluid – Peritoneal fluid should be filtered and the filters cultured, because the concentration of organisms in the fluid is usually very low.
3. Antimicrobial therapy – This should include drugs that are active against pneumococci as well as enteric bacilli. A suitable drug is a third-generation cephalosporin, such as ceftriaxone or cefotaxime.
4. Prevention – Patients with the nephrotic syndrome should be immunized against pneumococcal infection with the heptavalent conjugate pneumococcal vaccine. Those older than 2 years should also receive the 23-valent polysaccharide vaccine.

This patient was treated as described here, and he recovered. The blood culture grew out *Streptococcus pneumoniae*.

Reading:

Gorensek MJ, Lebel MH, Nelson JD: Peritonitis in children with nephrotic syndrome. *Pediatrics* 1988; 81: 849–856.

Krensky AM, Ingelfinger JR, Grupe WE: Peritonitis in childhood nephrotic syndrome. *Am J Dis Child* 1982; 136: 732–736.

CASE 114. An 18-month-old girl presents at about 2 AM with a 1-day history of a noisy cough and fever. The illness started 2 days ago with a runny nose. There is no history of a choking episode. She is brought at this time because she has started having difficulty breathing and her breaths are noisy.

On examination she is in marked respiratory distress and has obvious inspiratory stridor. Her heart rate is 140/minute, respiratory rate 40/minute, and temperature 38.7°C. Although she is alert, she is very agitated and moving around a lot, while lying in her father's arms. She is mildly cyanosed and has a palpable pulsus paradoxus. Her respiratory tract examination reveals marked suprasternal and subcostal retractions on inspiration. The lungs are resonant to percussion, and, on auscultation, breath sounds are barely audible.

- ***What is your initial assessment?***
- ***What do you want to do?***
- ***What is the differential diagnosis?***

This child is in respiratory failure, due to obstruction of the extrathoracic airway, and respiratory arrest is imminent. Therefore endotracheal intubation

should be performed immediately. Once this has been accomplished and the child's condition has been stabilized, a differential diagnosis can be considered. The only condition which can present with stridor, and in which the management might be different, is a foreign body inhalation. In that case attempts should be made to dislodge it.

The mechanism of stridor and the differential diagnosis follows.

Stridor is an inspiratory noise that reflects obstruction of the larynx and extrathoracic trachea. Resistance to airflow is inversely proportional to the airway radius to the fourth power. Thus reduction of airway radius by one half causes a sixteenfold increase in airflow resistance. The noise and phase of respiration during which the noise occurs is very important in determining the site of respiratory obstruction. This is illustrated in Figure 114.1.

During inspiration, as the diaphragm descends and the thoracic wall moves outward, the lungs and intrathoracic airways expand; the airways outside the thorax (larynx and upper trachea) become narrower, due to the negative pressure within the lumen. Therefore any narrowing of the larynx or upper trachea, for example, that caused by mucosal swelling, is exacerbated during inspiration, resulting in stridor. During expiration the reverse occurs. Therefore narrowing in the intrathoracic airways, for example, asthma or bronchiolitis, is exacerbated during expiration. In such cases the noise produced is wheezing, which is more pronounced during expiration.

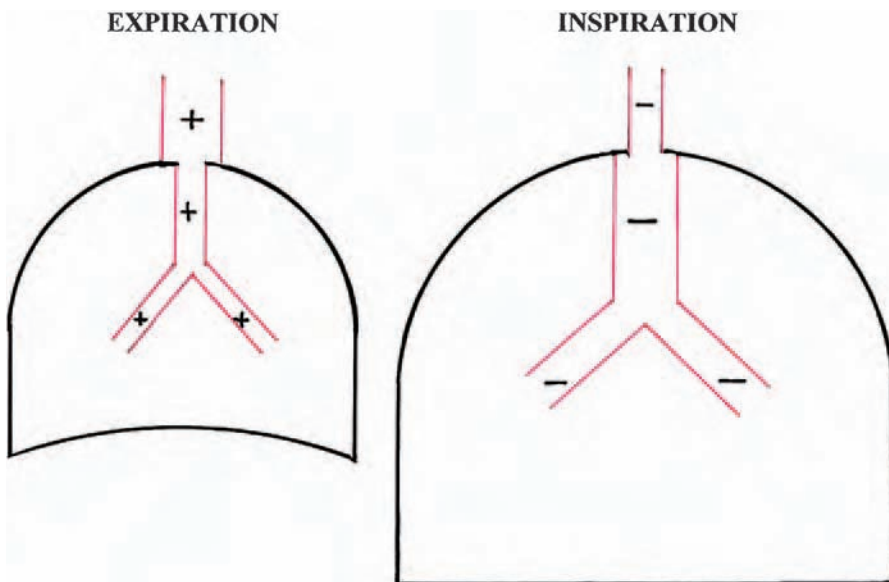


FIG 114.1. Changes in airway pressure during the stages of respiration (+, positive pressure; -, negative pressure).

■ **TAB. 114.1: Differential diagnosis of acute stridor.**

1. Mechanical obstruction
 - foreign body inhalation
2. Infections
 - epiglottitis
 - retropharyngeal abscess
 - diphtheria
 - viral croup (laryngotracheobronchitis)
 - bacterial tracheitis
3. Chronic obstruction aggravated by a superimposed viral infection
 - laryngotracheomalacia
 - laryngeal web
 - laryngeal cyst, tumor, hemangioma
 - laryngeal papilloma
4. Bilateral recurrent laryngeal nerve palsy
5. Hypocalcemia
6. External pressure
 - rapidly enlarging lymph node due to malignancy or tuberculosis

Clinical features indicating severe obstruction are:

mental status changes, including agitation
 marked sternal, suprasternal, or subcostal retractions
 palpable pulsus paradoxus
 marked tachycardia
 poor or absent breath sounds

The commonest cause of stridor is laryngotracheobronchitis, which was the diagnosis in this child.

A discussion of epiglottitis, viral croup, and bacterial tracheitis follows.

Epiglottitis: This is a bacterial infection of the epiglottis *and* the periepiglottic tissue. It was formerly caused mainly by *Haemophilus influenzae* type b, but as a result of widespread use of a vaccine against this organism, it is now caused mainly by *Streptococcus pneumoniae* and *Streptococcus pyogenes*. The disease is characterized by marked swelling of the epiglottis and aryepiglottic folds, resulting in a circle of swelling around the airway, which may totally occlude it. The clinical features are fever, cough, stridor, drooling, and respiratory distress. The child prefers to sit, with the neck forward. Because the airway in this condition can be severely compromised, this must be treated as an extreme emergency. When this condition is suspected, under ideal circumstances, the patient should be taken to an area, for example, operating room, where he/she can be have the airway examined by somebody skilled in endotracheal intubation. The airway obstruction can be so severe that the only clue to the presence of an airway is a bubble. The

management is endotracheal intubation to secure the airway. Once this is accomplished, antimicrobial therapy can be addressed. This should be directed at *Haemophilus influenzae*, *Streptococcus pneumoniae*, and *Streptococcus pyogenes*. Ceftriaxone and cefotaxime are suitable choices.

Viral croup (Laryngotracheobronchitis): This is caused by several respiratory viruses, the most common of which is parainfluenza virus. Infants and young children present with a history of fever, a barking cough, and stridor, often preceded by rhinorrhea. The stridor often develops during the middle of the night. This diagnosis, which is usually made clinically, is the most common of those affecting the middle respiratory tract (larynx and extrathoracic trachea).

Management: Most patients require no specific treatment. Humidification of the atmosphere has been used traditionally and may be helpful, but its value has not been demonstrated in a clinical trial. Dexamethasone, administered orally or parentally has been shown to bring about clinical improvement. Inhalation of epinephrine may also be of value in more severe cases. Inhalation of a 70% helium/30% oxygen mixture may help severe cases by lowering the resistance to airflow. Patients with the physical findings of severe disease, as described in the case scenario above, should undergo endotracheal intubation immediately.

Bacterial tracheitis: This is a severe infection in which the patient is toxic-appearing and has the features of middle respiratory tract obstruction. He/she coughs up (or is found at endotracheal intubation to have) large amounts of purulent secretions arising from the trachea. It usually complicates a previous epithelial injury such as that caused by a viral infection, in particular that caused by measles or influenza. The most common bacterial cause is *Staphylococcus aureus*. The diagnosis is based on clinical features and sometimes on findings at endoscopy. Treatment consists of establishing an adequate airway and of antibiotics directed at the likely agent. Examination of secretions by Gram stain is very important for choosing empiric antibiotic therapy.

Reading:

Brown JC: The management of croup. *British Medical Bull* 2002 ; 61: 189–202.

CASE 115. A 15-year-old previously healthy girl presents with a history of abdominal pain for 2 days. It started in the lower abdomen and has remained there. It is constant, aggravated by movement, but not crampy in nature. Her last menstrual period began 2 weeks ago and lasted 1 week.

There is nausea but no vomiting or diarrhea. There are no respiratory or urinary symptoms. On examination she is moderately ill-appearing. Her temperature is 37.9°C, and the rest of her vital signs are normal. The main finding is marked abdominal tenderness across the whole of the lower abdomen, on the right side more than the left side. She also has tenderness in the right upper quadrant, but neither the liver nor gallbladder can be palpated.

- *What is your differential diagnosis?*
- *What would you do?*

Pain in the lower abdomen is derived primarily from one of three organs, namely the intestine, urinary tract, or genital tract. In addition the organs that can give rise to right upper quadrant tenderness are the liver, gallbladder, pancreas, intestine, and kidney. Therefore the differential diagnosis is broad. Diseases of the liver, gallbladder, and pancreas do not, as a rule, cause lower abdominal pain, which is her main symptom. However, inflammatory disease of the Fallopian tubes can result in spread of inflammation up the right paracolic gutter to the perihepatic region, to cause perihepatitis (Fitz-Hugh-Curtis syndrome).

The differential diagnosis includes the following:

Intestinal disease

acute appendicitis – her symptoms and signs are too diffuse, and she does not appear to have generalized peritonitis

Genital tract – salpingitis (pelvic inflammatory disease)

ruptured ovarian cyst

ovarian torsion

ectopic pregnancy

labor – this is crampy in nature

endometriosis

Urinary tract

infection – the symptoms seem too extensive for this condition

It is very important to enquire about a history of sexual intercourse (although denial by the patient does not necessarily exclude this possibility, and does not necessarily exclude the possibility of a sexually transmitted infection or pregnancy) and to perform a genital examination.

This patient was, indeed, sexually active. Pelvic examination revealed a purulent discharge issuing from the cervical os. In addition there was tenderness of the adnexa, and movement of the cervix elicited pain. These features strongly suggest the diagnosis of pelvic inflammatory disease.

The following tests should be performed:

- (a) Microscopic examination of the discharge (wet preparation) to look for leukocytes, “clue cells,” which are characteristic of bacterial vaginosis, and for *Trichomonas vaginalis*.
- (b) A culture from the cervix for *Neisseria gonorrhoeae* and a test for *Chlamydia trachomatis* (antigen, nucleic acid). Although *Chlamydia trachomatis* can be cultured in tissue culture, this is expensive and should be reserved for use in prepubertal children (Figure 115.1).
- (c) Screening tests for evidence of other sexually transmitted infections, including hepatitis B, syphilis, and human immunodeficiency virus (HIV) infection.
- (d) A pregnancy test. A positive pregnancy test has the following implications:
 - (i) the diagnosis might be an ectopic pregnancy;
 - (ii) if treatment for pelvic inflammatory disease is to be administered, a tetracycline should NOT be given;
 - (iii) radiological (excluding ultrasound) procedures of the abdomen are contraindicated.

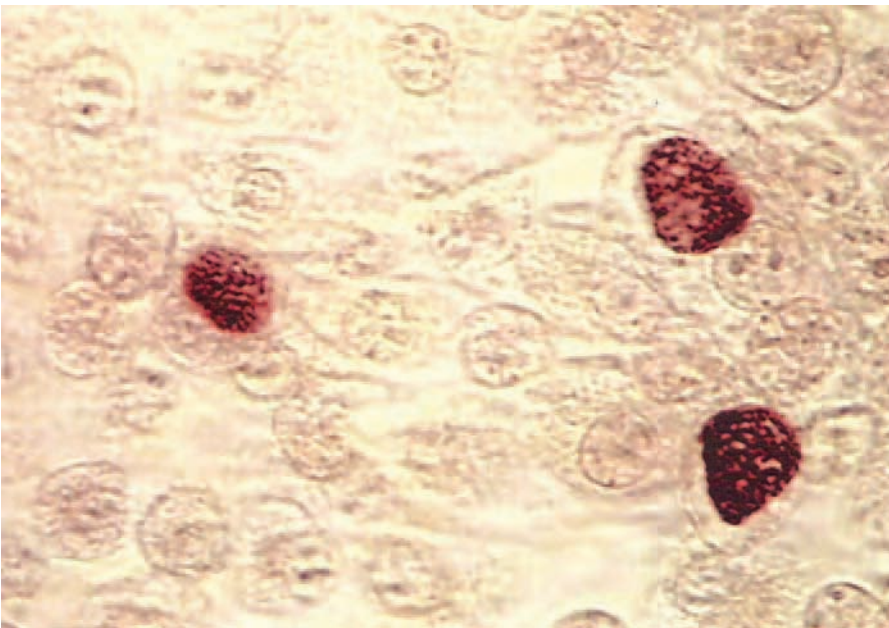


FIG 115.1. Tissue culture (McCoy cells) showing intracellular inclusions representing *Chlamydia trachomatis*. (Courtesy of the Centers for Disease Control and Prevention/Dr E Arum, Dr N Jacobs).

- (e) Urinalysis, microscopy, and culture. However, if there is pus in the vagina, which can readily interfere with obtaining a clean urine specimen, these tests may show numerous leukocytes and the culture may be falsely positive.

If the pregnancy test is negative and there is no vaginal or cervical pus, the other diagnostic possibilities become more likely. Therefore, depending on the progression of her course, other diagnostic tests should be considered, including an abdominal and pelvic ultrasound or computer tomography scan.

This patient's pregnancy test was negative.

She was diagnosed with pelvic inflammatory disease, complicated by the Fitz-Hugh-Curtis syndrome.

She received antimicrobial therapy with cefoxitin and doxycycline, and she had largely recovered within 2 days. Her test for *Chlamydia trachomatis* was positive. She was counseled to use condoms during sexual intercourse and to advise her sexual partner(s) to seek medical treatment. Contraception was also discussed with her.

Pelvic Inflammatory Disease(PID) is the term used to refer to infection of the Fallopian tubes (salpingitis) and endometrium. It is an important cause of morbidity in adolescent girls and an important cause of infertility.

Pathogenesis: Infectious agents, primarily *Chlamydia trachomatis*, and, to a lesser extent, *Neisseria gonorrhoeae*, ascend from the cervix, into the uterus, and thence into the tubes, where they cause tubal epithelial damage. There is secondary invasion by normal vaginal flora, such as enteric rods, and anaerobes. The purulent discharge within the tubes can spread into the peritoneal cavity, resulting in localized peritonitis. A tubo-ovarian abscess may also develop. The discharge may extend to the capsule of the liver, resulting in perihepatitis. The main long-term complications resulting from the tubular damage are infertility and ectopic pregnancy. Adhesions resulting from the peritonitis can result in intestinal obstruction.

Diagnosis: The diagnosis is made clinically. Confirming the diagnosis is seldom accomplished because the "gold standard" for the diagnosis, namely visualizing the tubes at laparoscopy, is seldom done. Although the diagnostic features (adnexal tenderness, cervical motion tenderness, and lower abdominal pain) are suggestive, they are nonspecific. The differential diagnosis is discussed above.

Management: Whether patients should be hospitalized or not should be determined by their severity of illness and the degree of certainty of the diagnosis. I prefer to hospitalize patients with severe abdominal pain, until

their symptoms have significantly improved. Firstly this allows for observation of the patient for improvement, particularly since deterioration would suggest an alternative diagnosis, for which they might require different management, and, secondly, it ensures that the patient complies with therapy initially.

The mainstay of therapy is antimicrobial. Several options are available. The chosen agents should provide antimicrobial activity against *Chlamydia trachomatis*, *Neisseria gonorrhoeae*, enteric rods, and anaerobic bacteria. I follow the recommendations of the US Public Health Service (see Reading). My preferred regimen is:

Doxycycline 100 mg every 12 hours, orally or intravenously,

PLUS

Cefoxitin 2 g intravenously every 6 hours OR cefotetan 2 g every 12 hours intravenously

Once improvement has occurred, therapy can be completed on an out-patient basis with a course of doxycycline to complete 14 days of therapy.

Outcome: If the patient has not improved within 48 hours, one should consider the possibility that the diagnosis is mistaken or that a complication has developed, the most likely of which is a tubo-ovarian abscess. Imaging studies, in particular ultrasonography, may be helpful at this point.

Reading:

McCormack WM: Pelvic inflammatory disease. N Engl J Med 1994; 330: 115–119.

US Public Health Service. Guidelines for the treatment of sexually transmitted diseases. 2002. MMWR 2002; 51; RR-6: 48–52.

CASE 116 (HYP). A 12-year-old previously healthy boy presents with a history of fever and cough for about 5 days. In the past day he has become short of breath and weak. On examination he is ill-appearing, with a respiratory rate of 50 per minute, heart rate of 130 per minute, and blood pressure of 100/40, and he is perfusing well. His mucosae are pale, and he has mild jaundice. The heart is normal except for a 2/6 ejection systolic murmur, and the lungs are remarkable for retractions and diffuse crackles throughout the lower zones. The abdominal and neuromuscular examinations are normal.

- *What might be the problem?*
- *What would you do?*

The physiological problems are respiratory difficulty and pallor, but not shock. Therefore the first item of management is to provide additional oxygen. Pallor in the absence of shock indicates anemia. The findings of fever and pulmonary crackles suggest the presence of pneumonia. Although the crackles could be due to pulmonary edema, there are no signs of heart disease to suggest a cardiac cause for this problem. What might be causing pneumonia in a previously healthy boy? The preceding symptoms suggest a respiratory virus infection, such as influenza. What about the pallor and jaundice? These suggest a hemolytic anemia. The association of pulmonary disease and anemia should also suggest the possibility of pulmonary hemorrhage, as can occur with Goodpasture's syndrome and Wegener's granulomatosis. There is no preceding history of a genetic cause of hemolytic anemia, such as sickle cell disease. However, a sickle cell crisis affecting the lung (acute chest syndrome) could explain the child's condition very well. Infections can cause hemolysis by stimulating the production of autoantibodies, as in the case of *Mycoplasma pneumoniae* infection, or by precipitating a hemolytic crisis in the case of underlying glucose-6-phosphate-dehydrogenase (G6PD) deficiency. Making a hematologic diagnosis in this case is important for the following reasons: if there is an autoimmune hemolytic anemia, corticosteroid therapy might be beneficial; if this is a sickle cell acute chest crisis, there may be a very rapid deterioration, and a regular blood transfusion or an exchange transfusion

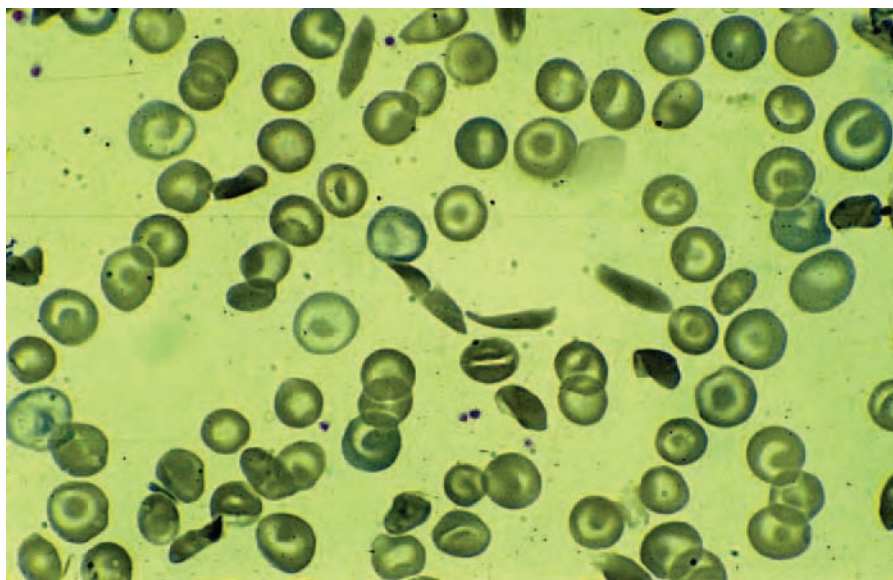


FIG 116.1. Blood smear showing target cells, a feature of hemoglobinopathies, and sickle cells.

might become necessary; if the diagnosis is G6PD deficiency, education about risk situations and drugs that can cause a hemolytic crisis is important.

Therefore tests that might help in making a diagnosis are a blood smear to examine the red cell morphology, a Coomb's test, hemoglobin electrophoresis and G6PD activity. The blood smear may show: diffuse basophilia (indicative of young red cells), agglutination of red cells in cold-agglutinin autoimmune hemolytic anemia, "bite cells," characteristic of G6PD deficiency, or sickle cells (Figure 116.1).

The patient nevertheless requires treatment for the pneumonia, with agents active against *Streptococcus pneumoniae* and *Mycoplasma pneumoniae*, such as a combination of a penicillin or third-generation cephalosporin together with a macrolide.

Reading:

Berkowitz FE: Hemolysis and infection: categories and mechanisms of their interrelationship. Rev Infect Dis 1991; 13: 1151–1162.

CASE 117. A 6-year-old girl presents with acute onset of fever and headache and is diagnosed with pneumococcal meningitis. Her left pupil does not respond to light, and the optic nerve is white and has a paucity of blood vessels. She has a history of a severe head injury as a result of a motor vehicle accident about 1 year ago. When examined a few days after institution of appropriate antimicrobial therapy, she is afebrile, alert, and has no nuchal rigidity.

- **Why might she have pneumococcal meningitis?**
- **How would you further evaluate her?**

The ocular findings are those of optic atrophy. This suggests that she indeed suffered a significant head injury, with damage to the optic nerve. Bacterial meningitis, in particular pneumococcal meningitis, after a significant head injury, strongly suggests a communication between the external environment and the subarachnoid space. When there is no obvious skull fracture, the likely site is a fracture through the base of the skull, either through the middle cranial fossa into the middle ear cavity or through the anterior cranial fossa through the cribriform plate. The latter can sometimes be demonstrated by having the patient lie prone with the head hanging over the edge of the bed. Cerebrospinal fluid (CSF) may begin to drip (CSF rhinorrhea). When performed on this patient, this maneuver elicited CSF rhinorrhea (Figure 117.1).



FIG 117.1. *The patient lying prone demonstrating CSF rhinorrhea.*

A computer tomography scan with narrow cuts can demonstrate a bony defect in the cribriform plate, as was accomplished in this case. This was repaired surgically.

CASE 118. A 13-year-old girl presents with severe headache and fever. She has had a left earache and ear discharge for about 1 week. On examination she is ill-appearing and has a temperature of 39°C. She has mild neck stiffness. The left external auditory canal is occluded with a purulent discharge, but there is no pain on movement of the pinna. There is no swelling over the mastoid bone. The rest of the examination is normal.

- *What is the differential diagnosis?*
- *What should be done?*

This patient has several clinical features that should be cause for concern:

- (a) A purulent auditory discharge suggests the possibilities of otitis externa or otitis media associated with a tympanic membrane perforation. The former is a very painful condition and is characterized by extreme pain elicited on movement of the pinna. Its absence indicates otitis media with perforation of the tympanic membrane.
- (b) Severe headache in the presence of infection of the middle ear or paranasal sinus should suggest the possibility of intracranial extension of the infection.
- (c) Neck stiffness supports the possibility of intracranial extension of infection, including meningitis, or a cerebellar abscess, or another site of intracranial suppuration with impending herniation. Intracranial suppuration from the middle ear or paranasal sinus can occur at any level: the epidural space, subdural space, subarachnoid space (meningitis), or brain parenchyma (cerebritis or brain abscess). Another intracranial complication of middle ear or paranasal sinus infection is dural sinus thrombosis.

Management:

Although it might be tempting to perform a lumbar puncture in such a case, the possibility of focal suppuration contraindicates this. Therefore management should consist of the following, in the following order:

- (i) obtain blood cultures and culture and Gram stain of the auditory discharge;
- (ii) initiate antimicrobial therapy (the drugs are discussed below);
- (iii) brain imaging;
- (iv) if no focal lesion or no significant cerebral edema are seen, lumbar puncture should be performed.

The computer tomography scan of this patient is shown in Figure 118.1.

This shows a large brain abscess in the temporal lobe. Brain abscesses arising from the middle ear occur in the temporal lobe or cerebellum. A wide variety of infectious agents can cause such abscesses, including *Streptococcus pneumoniae*, other streptococci, *Staphylococcus aureus*, anaerobes, such as *Bacteroides* spp. and *Fusobacterium* spp., and Gram-negative bacilli. The most common are streptococci and anaerobes.

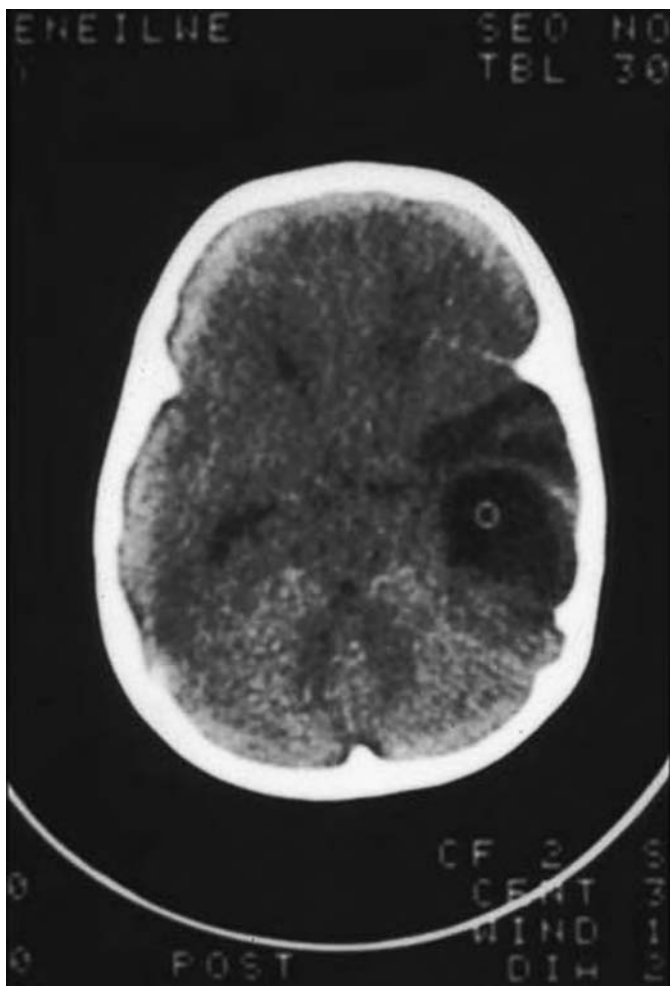


FIG 118.1. The computer tomography scan showing a large brain abscess in the temporal lobe.

Treatment should consist of antimicrobial therapy directed at the likely pathogens and surgical drainage. Suitable antimicrobial therapy would include the following:

ceftriaxone (or cefotaxime) + vancomycin + metronidazole

or

meropenem + vancomycin.

Reading:

Yogev R, Bar-Meir M: Management of brain abscess. *Pediatr Infect Dis J* 2004; 23: 157–159.

CASE 119. A 12-year-old previously well girl presents with a history of fever and cough for a few days, and difficulty breathing for 1 day. She lives in a rural area and has exposure to goats, cows, and many birds. (Her grandmother has an aviary.) She has no recent contact with sick individuals, and no knowledge of a tuberculosis contact. She has not been involved in any activities unusual for her. On examination she is very ill-appearing, with marked respiratory distress. There is mild cyanosis, but no pallor or jaundice. Her temperature is 38°C, heart rate 140/minute, and blood pressure 110/60, and her respiratory rate is 60/minute. The jugulo-venous pressure is not elevated and there is no peripheral edema. Examination of the chest shows marked intercostal retractions, and auscultation of the lungs reveals many crackles in both lower lobes. There are no signs of focal lung consolidation. The heart is not enlarged, the first and second heart sounds are normal, and there is no gallop. The rest of the examination is normal.

- *What is the differential diagnosis?*
- *What would you do?*

This patient has respiratory failure caused by a diffuse pulmonary disease, probably pneumonia. Although these pulmonary findings can also occur in left-sided heart failure, there are no other findings to indicate heart disease or a cause for heart disease. The most important action to take is to provide supplemental oxygen.

If this is indeed pneumonia, it is diffuse, a form that is sometimes referred as “atypical” pneumonia, which implies that it is not lobar. Although the patterns of pneumonia are not very specific for determining an etiological agent, this pattern does broaden the spectrum of agents that might be involved. The differential diagnosis includes the common causes of pneumonia in a 12-year-old girl, in particular *Mycoplasma pneumoniae*, *Chlamydia pneumoniae*, and *Streptococcus pneumoniae*, in addition to respiratory viruses such as influenza virus. Her exposures place her at particular risk for pneumonia caused by *Chlamydia psittaci* (from birds), *Coxiella burnetii* (from ungulates), and *Histoplasma capsulatum*. In addition she might have a hypersensitivity pneumonia caused by bird proteins (“bird fancier’s disease”). If she were an immunocompromised host, then additional agents such as cytomegalovirus, *Legionella pneumophila*, and *Pneumocystis jiroveci* infection would be important additional considerations.

The most useful diagnostic test to determine the extent of the disease and to demonstrate any complications of the disease, such as pleural effusions or cavitation, is a chest X ray. Her chest X ray is shown in Figure 119.1.

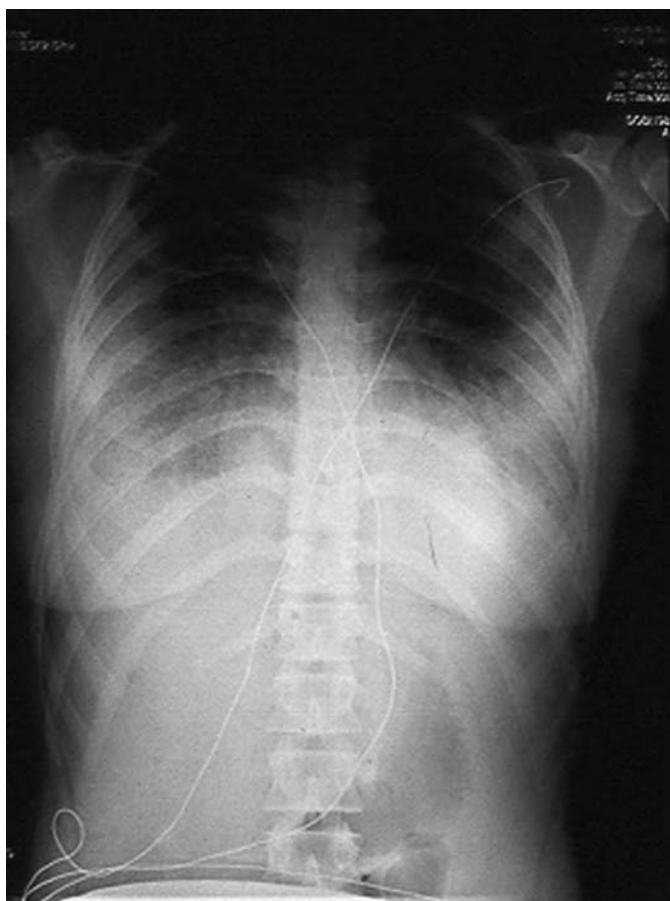


FIG 119.1. Chest X ray showing extensive, diffuse, bilateral pulmonary infiltrates.

In cases of pneumonia, the most useful test for reaching an etiologic diagnosis is examination of the sputum, by Gram stain and culture. The presence of numerous inflammatory cells and the absence of bacteria suggest an infection caused by an agent that is not stained by the Gram stain, such as *Mycoplasma pneumoniae*. Diagnosis of infections caused by *M. pneumoniae*, *C. pneumoniae*, *C. psittaci*, and *C. burnetii* depend largely on serology. Meanwhile she should be treated with a tetracycline (e.g. doxycycline) plus a third-generation cephalosporin or a penicillin; in addition a systemic corticosteroid should be administered in case this child has a hypersensitivity pneumonia.

This child was hospitalized in an intensive care unit. She required high concentrations of oxygen but did not require ventilation. She was treated with doxycycline and cefotaxime and improved over a few days. An etiology for her illness was not determined, despite extensive testing.

Reading:

Hindiyeh M, Carroll KC: Laboratory diagnosis of atypical pneumonia. *Semin Resp Infect* 2000; 15: 101–113.

CASE 120. A 12-year-old previously well boy presents with a history of fever, chills, sore throat, and neck pain for 5 days. He also has shortness of breath, nausea, and vomiting. He has had no exposures to animals and has not traveled outside his hometown in Georgia, USA. On examination he is moderately ill-appearing. His temperature is 39.1°C, heart rate 116/minute, respiratory rate 44/minute, and blood pressure 100/60 mm Hg, and his perfusion is good. He is not pale, but his sclerae are jaundiced. He has white patches on his tonsils, and examination of his lungs reveals crackles bilaterally. There is no lymphadenopathy and his cardiac examination and the rest of his examination are normal. He has a total leukocyte count of $12.5 \times 10^3/\mu\text{l}$, with 62% segmented neutrophils and 30% band forms, his total bilirubin is 4.1 mg/dl with a direct component of 3.6 mg/dl, and the aspartate aminotransaminase and alanine aminotransaminase are 58 and 35 units, respectively. His C-reactive protein concentration is 28 mg/dl.

He is admitted to hospital, and soon after admission he becomes hypotensive, necessitating his transfer to the intensive care unit.

- *What is your differential diagnosis?*
- *What would you do?*

This patient has evidence of septic shock, pharyngotonsillitis, pneumonia, and hepatitis. Which infections or infectious agents can cause this?

Viral agents:

Epstein–Barr virus
cytomegalovirus
adenovirus

Although these viruses can cause pharyngotonsillitis, hepatitis, and pneumonia, they do not cause shock, except in the case of splenic rupture in cases of Epstein–Barr virus infection.

The laboratory findings in this child are very suggestive of a severe bacterial infection (marked left shift of leukocytes and markedly elevated C-reactive protein concentration).

Bacterial agents:

Streptococcus pyogenes is the most important bacterial cause of pharyngotonsillitis. It can cause severe rapidly progressive pneumonia, but does not

cause hepatitis, unless bacteremia or toxic shock-like syndrome is present. The latter condition, which carries a very high case fatality rate, is characterized by hypotension, diffuse erythroderma, and multiorgan dysfunction. Most cases have positive blood cultures for the organism.

Staphylococcus aureus does not cause tonsillitis but can be associated with abscesses in the peripharyngeal spaces. It is a very important cause of disseminated infection, with metastatic infection in many organs including the liver, lungs, and skeleton. In addition it can cause toxic shock syndrome.

Corynebacterium diphtheriae causes pharyngotonsillitis but does not cause pneumonia or hepatitis. Shock is due to diphtheria toxin causing a toxic cardiomyopathy, which develops 2–3 weeks after the onset of illness.

Fusobacterium necrophorum (Figure 120.1), which is a normal component of the oral flora, can cause invasion of the venous drainage of the throat, resulting in a septic thrombophlebitis of the jugular vein or its tributaries and sepsis syndrome. This is called Lemierre syndrome. The initial focus of infection is in the throat, which may have been initiated by another agent, such as Epstein–Barr virus. A peritonsillar abscess may be present.

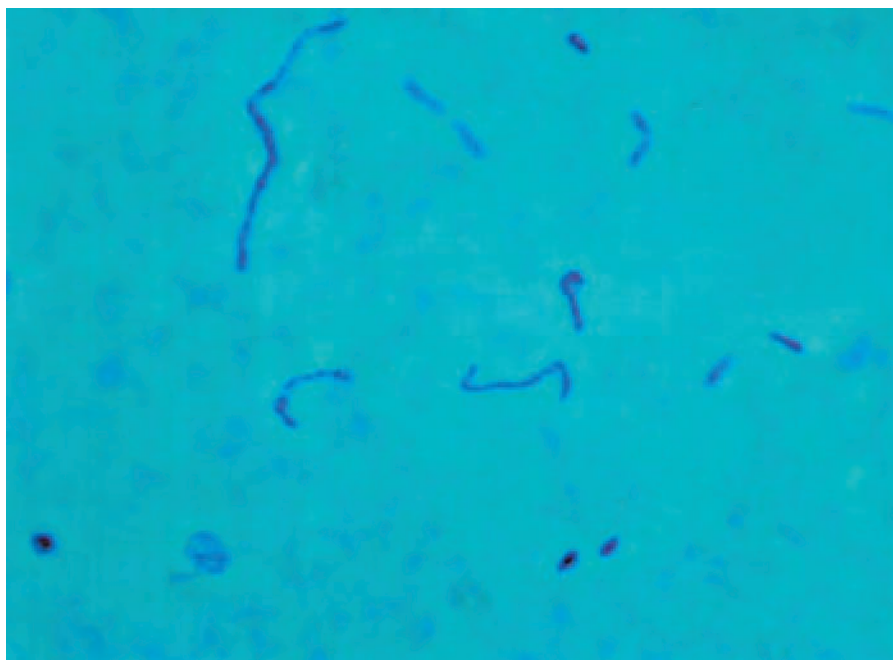


FIG 120.1. *Fusobacterium necrophorum* by phase contrast microscopy.
(Courtesy of the Centers for Disease Control and Prevention/Dr Lillian V Holderman)

Because the septic thrombophlebitis is an endovascular infection, it is frequently complicated by metastatic infection, particularly in the lung, but also in the liver, bone or joint, and skin. It may be associated with jaundice. The diagnosis is made clinically and can be confirmed by the demonstration of the organism on blood culture (Figure 120.1), and, sometimes, by demonstration of jugular vein occlusion by imaging. Therapy is primarily antimicrobial. Several antimicrobial agents are active against the organism, including clindamycin, metronidazole, a combination of a penicillin and a β -lactamase inhibitor, such as ampicillin/sulbactam, or a carbapenem. Any abscess should be drained surgically. Anticoagulation should also be considered, although this is controversial.

Management of this patient should entail the following:

- (a) ensuring adequate perfusion with aggressive intravenous fluid therapy and vasopressor therapy;
- (b) ensuring adequate oxygenation; he required supplemental oxygen.
- (c) antimicrobial therapy active against *Streptococcus pyogenes*, *Staphylococcus aureus*, and *Fusobacterium necrophorum*. He was treated initially with vancomycin, and meropenem was subsequently added to his regimen.
- (d) attempting to make a diagnosis:

blood culture: this eventually grew out *Fusobacterium necrophorum*.

chest X ray: this showed multiple foci of consolidation suggestive of embolic infection in the lungs (hematogenous pneumonia).

imaging of the neck, to look for evidence of venous occlusion: a Doppler ultrasound demonstrated a thrombus in the left internal jugular vein. As a result of this finding he was anticoagulated with low molecular weight heparin. Computer tomography imaging should be considered to look for evidence of a parapharyngeal abscess. This was not done.

He recovered and, after hospital discharge, was treated with oral clindamycin for 4 weeks.

Reading:

Ramirez S, Hild TG, Rudolph CN et al: Increased diagnosis of Lemierre's syndrome and other *Fusobacterium necrophorum* infections at a children's hospital. *Pediatrics*. 2003; 112: e380–385.

Venglarcik J: Lemierre's syndrome. *Pediatr Infect Dis J* 2003 2003; 22: 921–923.

Goldenberg NA, Knapp-Clevenger R, Hays T, Manco-Johnson MJ: Lemierre's and Lemierre's-like syndromes in children: survival and thromboembolic outcomes. *Pediatrics* 2005; 116: e543–548.

CASE 121. A 6-year-old girl underwent a major neurosurgical procedure 1 week ago to drain a large interhemispheric epidural abscess complicating sinusitis. A specimen of pus taken at surgery showed Gram-positive cocci in pairs, but the culture was negative. Because the parents had initially refused to grant permission for surgery, surgical drainage had been delayed for about 1 week. During this time the child was treated with vancomycin, ceftriaxone, and metronidazole, all intravenously, through a percutaneously inserted central venous catheter (PICC). After surgery her clinical condition improved markedly. However, despite broad-spectrum antimicrobial therapy for 2 weeks and surgical drainage, she has had a high fever. She feels well and has no other symptoms. On examination she is well appearing and very cooperative. Except for the large surgical scar along the length of her head, which is clean and without swelling, erythema, or drainage, examination of her head is normal. She has full range of ocular movements (initially she had a right third nerve palsy) and a normal neurological examination. The rest of her examination is normal.

- *What are the possible causes of fever in this child?*
- *What would you do?*

This child has several possible reasons for having fever:

- (i) intracranial infection which has not been adequately drained;
- (ii) infection introduced during the surgical procedure;
- (iii) nosocomial infection, particularly related to the intravascular catheter;
- (iv) drug fever.

The child's clinical improvement and lack of symptoms suggest that a continuing intracranial infection is unlikely. Nevertheless imaging to determine whether there is progression of the abscess should be performed. This showed the identical appearance to that seen a few days after surgery, which was a marked improvement over the preoperative appearance.

Possible sites of nosocomial should be sought clinically. Specifically the site of the intravascular catheter should be carefully examined for any signs of inflammation. In addition a blood culture drawn through the catheter should be performed. This clinical evaluation was normal, and the blood culture was negative.

The possibility of drug-induced fever should be considered. There are several different mechanisms by which drugs can cause fever:

- (i) altered thermoregulation, as can occur with anticholinergics;
- (ii) pyrogenic properties of the drug preparation such as amphotericin B deoxycholate, or contaminants within the drug preparation;
- (iii) pharmacologic effects of the drug, such as the Jarisch-Herxheimer reaction that occurs during therapy for certain infections and that is thought to be due to the release of endotoxin by dying bacteria;
- (iv) idiosyncratic reactions of certain individuals to anesthetic agents resulting in malignant hyperthermia;
- (v) presumed hypersensitivity or immune-mediated reactions: the mechanism is unknown. This is the type usually implied by the term “drug fever.” Patients may have been taking the incriminated drug for a prolonged period before the problem occurs. Although the fever may be very high, the patient is not usually ill-appearing. In adults a relative bradycardia is common. The erythrocyte sedimentation is usually markedly elevated. Although many drugs can cause this, among antimicrobial agents, β -lactams are particularly important. The only way of diagnosing this problem is to discontinue the drug and to observe whether the fever abates, which occurs within 72 hours. This patient had been receiving ceftriaxone (in addition to other agents) for about 2 weeks. Since this was considered the most likely cause of drug fever in her, this drug and metronidazole were replaced by meropenem. Her fever abated within 48 hours. No further febrile episodes occurred.

Reading:

Johnson DH, Cunha BA: Drug Fever. *Infect Dis Clin N Am* 1996; 10: 85–91.



APPENDIX

■ **Table A1: Infectious diseases reportable in the United States.**

Acquired immunodeficiency syndrome (AIDS)
Anthrax
Botulism
Brucellosis
Chancroid
Chlamydia trachomatis genital infection
Cholera
Coccidioidomycosis
Cryptosporidiosis
Cyclosporiasis
Diphtheria
Domestic arboviral disease – California, eastern equine, Powassan, St Louis, western equine
Ehrlichiosis
Enterohemorrhagic *Escherichia coli* O157:H7 and Shiga toxin-producing non-O157
Giardiasis
Gonorrhea
Haemophilus influenzae, invasive disease
Hansen's disease (leprosy)
Hantavirus pulmonary syndrome
Hemolytic uremic syndrome, postdiarrheal
Hepatitis A
Hepatitis B
Hepatitis C/non-A, non-B hepatitis
Human immunodeficiency virus infection (HIV)
Legionellosis
Listeriosis
Lyme disease
Malaria
Measles
Meningococcal disease

(continued)

(continued)

Mumps
 Pertussis
 Plague
 Poliomyelitis, paralytic
 Psittacosis
 Q fever
 Rabies, animal and human
 Rocky Mountain spotted fever
 Rubella (postnatal and congenital)
 Salmonellosis
 Shigellosis
Staphylococcus aureus infection
 Vancomycin-intermediate resistant (VISA)
 Vancomycin-resistant (VRSA)
 Streptococcal disease, invasive, group A
Streptococcus pneumoniae
 drug-resistant, invasive disease, all ages
 invasive disease children <5 years
 Streptococcal toxic shock syndrome
 Syphilis (all forms)
 Tetanus
 Toxic shock syndrome
 Trichinellosis
 Tuberculosis
 Tularemia
 Typhoid fever
 Varicella
 Yellow fever
 West Nile virus disease (all forms)

Table A2: Taxonomy of human pathogens, their usual sources, and the main clinical syndromes they cause.

Pathogens	Usual source	Clinical syndrome
VIRUSES		
DNA VIRUSES		
Herpesviridae		
Alphaherpesvirinae		
Herpes simplex 1	H, Mat	Gingivostomatitis, fever blisters, encephalitis, keratitis, neonatal infection

(continued)

Table A2: Taxonomy of human pathogens, their usual sources, and the main clinical syndromes they cause (continued)

Pathogens	Usual source	Clinical syndrome
Herpes simplex 2	H, Mat	Genital ulcers, neonatal infection
Varicella zoster virus	H	Chickenpox (varicella), zoster (shingles)
Betaherpesvirinae		
Cytomegalovirus	H, Mat	Infectious mononucleosis, hepatitis, OI – pneumonia, retinitis, enteritis
Human herpes virus 6	H	Fever, roseola, OI – hepatitis, encephalitis
Gammaherpesvirinae		
Epstein–Barr virus	H	Infectious mononucleosis, hepatitis, OI – posttransplant lymphoproliferative disease, Burkitt's lymphoma, other cancers
Human herpes virus 7	H	Fever
Human herpes virus 8	H	Kaposi sarcoma
Adenoviruses		
Multiple serotypes	H	Conjunctivitis, respiratory infection, cystitis, OI
Enteric (40, 41)		Diarrhea
Hepadnaviridae		
Hepatitis B virus	H, Mat	Hepatitis, chronic liver disease, liver cancer
Papovaviridae		
Polyoma viruses	H	OI – progressive multifocal leukoencephalopathy, nephropathy following renal transplant
Papilloma virus	H	Warts – skin, genital, laryngeal, cervical cancer
Parvoviridae		
Parvovirus B 19	H	Erythema infectiosum, aplastic crisis, hydrops fetalis, chronic anemia, OI – hepatitis, encephalitis
Poxviridae		
Chordopoxvirinae		
Orthopoxvirus		
Smallpox virus	H	Smallpox
Vaccinia virus	L	Vaccinia-local, disseminated, encephalitis, myopericarditis
Cowpox virus		
Monkeypox virus	A	Monkeypox
Molluscipox virus		
Molluscum contagiosum	H	Molluscum contagiosum
Parapoxvirus		
Orf virus	A	Orf
Yatapoxvirus		
Tanapoxvirus		

(continued)

(continued)

Pathogens	Usual source	Clinical syndrome
RNA VIRUSES		
Picornaviridae		
Enteroviruses		
Polio viruses	H	Poliomyelitis, encephalitis
Coxsackie, Echovirus	H	Fever, rash, meningitis, encephalitis
Parechovirus	H	Myocarditis, neonatal infection
Hepatovirus		
Human hepatitis A virus	H	Acute hepatitis
Rhinoviruses	H	Upper respiratory tract infection, sinusitis
Caliciviridae		
Noroviruses	H	Diarrhea
Paramyxoviridae		
Measles virus	H	Measles
Mumps virus	H	Mumps
Respiratory syncytial virus	H	Bronchiolitis, pneumonia, URI
Parainfluenza viruses (1,2,3,4)	H	Respiratory tract infection
Human metapneumovirus	H	Respiratory tract infection
Reoviridae		
Rotavirus	H	Diarrhea
Coltivirus		
Colorado tick fever virus	Tick	Colorado tick fever
Coronaviridae	A	Severe acute respiratory syndrome (SARS)
	H	Diarrhea, upper respiratory infection
Astroviridae		
Human astrovirus	H	Diarrhea
Orthomyxoviridae		
Influenza viruses A,B,C	H, A	Influenza
Retroviridae		
Lentivirus		
Human immunodeficiency virus (HIV) 1	H, Mat	HIV infection, AIDS
HIV 2	H, Mat	HIV infection, AIDS
Deltaretrovirus		
Human T-cell lymphotropic virus (HTLV)	H	T-cell leukemia
	H	Tropical spastic paraparesis
Togaviridae		
Rubivirus		
Rubella virus	H, Mat	Rubella, congenital rubella syndrome
Alphaviruses	Mos	Arthropod-borne viruses
Many arthropod-borne viruses		Equine encephalitides

(continued)

Table A2: Taxonomy of human pathogens, their usual sources, and the main clinical syndromes they cause (continued)

Pathogens	Usual source	Clinical syndrome
Flaviviridae		
Hepacavirus		
Hepatitis C	H	Hepatitis, chronic liver disease
Flavivirus		
Many arthropod-borne viruses	Mos	Yellow fever, dengue, West Nile, Japanese B encephalitis
Arenaviridae		
Lassa fever virus	A	Hemorrhagic fever
Lymphocytic choriomeningitis virus (LCM)	A, Mat	Meningitis, congenital infection
Junin, Machupo, Sabia, Guanarito	A	South American hemorrhagic fevers
Rhabdoviridae		
Lyssaviruses		
Rabies virus	A	Rabies – encephalitis
Filoviridae		
Ebola virus	A	Hemorrhagic fever
Marburg virus	A	Hemorrhagic fever
Bunyaviridae		
Bunyavirus		
California, La Crosse encephalitis	Mos	Encephalitis
Many arthropod-borne viruses		
Phlebovirus		
Sandfly fever virus	Fly	Sandfly fever
Rift Valley fever virus	Mos	Rift Valley fever
Hantaviruses		
Hantaan virus	A	Korean hemorrhagic fever
Sin nombre virus	A	Pulmonary syndrome
Nairovirus		
Congo-Crimean hemorrhagic fever virus	Tick	Hemorrhagic fever
Hepatitis D virus	H	Chronic hepatitis associated with HBV
BACTERIA		
Obligate intracellular replication, lack cell wall		
CHLAMYDIAE		
<i>C. trachomatis</i>		
Serovars B, Da, Ia, D-K	H, Mat	Genital tract infection, neonatal conjunctivitis, pneumonia
Serovars A, B, Ba, C	H, Fly	Trachoma
Serovars L1, L2, L3	H	Lymphogranuloma venereum
<i>C. psittaci</i>	A	Pneumonia
<i>C. pneumoniae</i>	H	Pneumonia

(continued)

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Pathogens	Usual source	Clinical syndrome
RICKETTSIAE		
<i>R. prowazekii</i>	Louse	Epidemic (louse-borne) typhus
<i>R. mooseri (typhi)</i>	Flea	Endemic (flea-borne) typhus
<i>R. rickettsii</i>	Tick	Rocky Mountain spotted fever
<i>R. conori</i>	Tick	Mediterranean spotted fever
<i>R. africae</i>	Tick	African tick typhus
<i>Orientia tsutsugamushi</i>	Mite	Scrub typhus
<i>R. akari</i>	Mite	Rickettsial pox
<i>R. sibirica</i>	Tick	Spotted fever
<i>R. australis</i>	Tick	Queensland tick typhus
<i>Coxiella burnetii</i>	A, tick	Q fever (hepatitis, pneumonia)
<i>Ehrlichia</i>	Tick	Ehrlichiosis
<i>Ehrlichia chaffensis</i>		Human monocytic ehrlichiosis
<i>Anaplasma phagocytophilum</i>		Human granulocytic ehrlichiosis
MYCOPLASMAS		
<i>M. pneumoniae</i>	H	Pneumonia, rash, encephalitis
<i>M. hominis</i>	H, Mat	Neonatal pneumonia
<i>M. genitalium</i>	H	Urethritis
<i>Ureaplasma urealyticum</i>	H, Mat	Neonatal pneumonia
SPIROCHETES		
<i>Treponema pallidum</i> subsp. <i>pallidum</i>	H	Syphilis
<i>Treponema pallidum</i> subsp. <i>pertenue</i>	H	Yaws
<i>Treponema pallidum</i> subsp. <i>endemicum</i>	H	Bejel
<i>Treponema carateum</i>	H	Pinta
<i>Leptospira interrogans</i>	A, E	Leptospirosis
Multiple serotypes		
<i>Borrelia</i>		
<i>B. burgdorferi</i>	Tick	Lyme disease
<i>B. recurrentis</i>	Louse	Relapsing fever (louse-borne)
<i>B. duttoni</i>	Tick	Relapsing fever
<i>B. hermsii, turicatae, parkeri</i>	Tick	Relapsing fever (tick-borne)
<i>Spirillum minus</i>	A	Rat bite fever

Gram-negative bacilli

Enterobacteriaceae – ferment glucose, oxidase-negative, reduce nitrate to nitrite, facultative anaerobes, inhabit the intestine; cause infections arising from the intestine, such as abdominal abscesses, urinary tract infections; nosocomial infections.

(continued)

Table A2: Taxonomy of human pathogens, their usual sources, and the main clinical syndromes they cause (continued)

Pathogens	Usual source	Clinical syndrome
<i>Escherichia coli</i>	A, H, End	Urinary infection, neonatal sepsis, sepsis, NI, specific virulence factors associated with specific forms of enteric infection
<i>Shigella</i> Four species: <i>S. dysenteriae</i> , <i>flexneri</i> , <i>boydii</i> , <i>sonnei</i>	H	Diarrhea, dysentery, encephalopathy
<i>Klebsiella pneumoniae</i> and other species	H, End	UTI, pneumonia, NI
<i>Enterobacter</i>	H, End	UTI, NI
<i>Citrobacter</i>	H, End	NI
<i>Serratia</i>	H, End	NI
<i>Proteus</i>	H, End	UTI, NI
<i>Morganella</i>	H, End	UTI, NI
<i>Providencia</i>	H, End	UTI, NI
<i>Hafnia</i>		
<i>Edwardsiella</i>		
<i>Pantoea</i>		
<i>Salmonella</i> >2400 serotypes	A	Diarrhea, dysentery, bacteremia
serotype typhi	H	Typhoid fever
<i>Yersinia pestis</i>	A, Flea	Plague
<i>Yersinia enterocolitica</i>	A	Diarrhea, dysentery, pseudoappendicitis
NONFERMENTERS (MANY FAMILIES)		
<i>Pseudomonas aeruginosa</i>	E	NI, pneumonia especially associated with cystic fibrosis
<i>Burkholderia cepacia</i>	E, H	Pneumonia associated with cystic fibrosis
<i>Burkholderia pseudomallei</i>	E	Melioidosis
<i>Stenotrophomonas maltophilia</i>	E	NI
<i>Acinetobacter</i> species	E	NI
<i>Elizabethkingae meningosepticum</i>	E	Neonatal sepsis, meningitis
SMALL GRAM-NEGATIVE BACILLI (COCCOBACILLI)		
<i>Bordetella pertussis</i>	H	Pertussis
<i>Brucella</i> (four species)	A	Brucellosis
<i>Capnocytophaga canimorsis</i>	A	Dog-bite wound
<i>Calymmatobacterium granulomatis</i>	H	Granuloma inguinale (Donovanosis)
<i>Campylobacter fetus</i>	A	Sepsis (OI)
<i>Campylobacter jejuni</i>	A	Diarrhea, dysentery
<i>Helicobacter pylori</i>	H	Gastritis
<i>Francisella tularensis</i>	A, arthropods	tularemia

(continued)

(continued)

Pathogens	Usual source	Clinical syndrome
<i>Haemophilus</i>		
Influenzae type b	H	Bacteremia, meningitis, septic arthritis, epiglottitis
Nontypeable	End	Otitis media, sinusitis, pneumonia, conjunctivitis
<i>Haemophilus aphrophilus</i>	H	Brazilian hemorrhagic fever
<i>Haemophilus ducreyi</i>	End	IE
<i>Legionella pneumophila</i>	H	Chancroid
Other <i>Legionella</i> species	E	Legionnaires' disease (pneumonia)
<i>Actinobacillus</i>	E	"
<i>actinomycetemcomitans</i>	End	IE
<i>Cardiobacterium hominis</i>	End	Mouth, bite-wound
<i>Eikenella corrodens</i>	End	Skeletal infections
<i>Kingella kingae</i>	A (rat)	Rat bite fever
<i>Streptobacillus moniliformis</i>		
Bartonellae		
<i>B. bacilliformis</i>	Fly	Oroya fever
<i>B. henselae</i>	A	Cat-scratch disease, bacillary angiomatosis
<i>B. quintana</i>	Louse	Trench fever, bacillary angiomatosis, IE
<i>B. elizabethae</i>	A	IE
NON-ENTEROBACTERIACEAE, FERMENTERS		
<i>Aeromonas hydrophila</i>	E	Water-associated infections
<i>Chromobacterium violaceum</i>	E	Soil-associated infections (OI)
<i>Pasteurella multocida</i>	A	Animal bite-wound infections
<i>Plesiomonas shigelloides</i>		Diarrhea
Vibrios		
<i>V. cholerae</i>	E, H	Cholera
<i>V. parahaemolyticus</i>	E	Diarrhea
<i>V. vulnificus</i>	E	OI – sepsis, oyster-associated
Gram-negative cocci		
<i>Neisseria</i>		
<i>N. meningitidis</i>	H	Septicemia, meningitis
<i>N. gonorrhoeae</i>	H, Mat	Gonorrhea, neonatal conjunctivitis
<i>Moraxella catarrhalis</i>	End	Otitis media, sinusitis
Gram-positive cocci		
CATALASE POSITIVE		
Staphylococci		
Coagulase-positive; <i>S. aureus</i>	H, End	Skin, wound, bacteremia, pneumonia, NI, IE, FB

(continued)

Table A2: Taxonomy of human pathogens, their usual sources, and the main clinical syndromes they cause (continued)

Pathogens	Usual source	Clinical syndrome
Coagulase-negative; many species Including <i>S. epidermidis</i> <i>S. haemolyticus</i> <i>Rothia mucilaginosa</i>	H, End	NI, FB iv catheters, ventricular shunts Cystitis OI – sepsis (neutropenia-associated)
CATALASE NEGATIVE		
Streptococci		
<i>S. pneumoniae</i>	H, End	Otitis media, sinusitis, conjunctivitis, pneumonia, bacteremia, meningitis
<i>S. pyogenes</i> (group A)	H	Tonsillitis, impetigo, wound, puerperal sepsis, toxic-shock like syndrome, acute glomerulonephritis, rheumatic fever
<i>S. agalactiae</i> (group B)	H, Mat	Neonatal sepsis, meningitis, OI – diabetics
<i>S. viridans</i>	End	Dental caries, endocarditis, visceral abscesses, OI – sepsis (neutropenia)
<i>S. bovis</i> (group D)	End	Endocarditis (colonic cancer-associated)
Enterococci		
<i>E. faecalis</i>	End	Endocarditis, NI, FB
<i>E. faecium</i>	"	"
<i>Pedococcus</i> spp.	E	OI – sepsis, NI vancomycin-resistant
<i>Gemella</i> sp.	End	Focal infections, IE
<i>Leukonostoc</i> spp.	E	NI, FB, OI vancomycin-resistant
Gram-positive bacilli, non-spore-forming		
<i>Actinomyces</i> several species (microaerophilic)		
<i>A. israeli</i>	End	Actinomycosis – chronic infection mouth, gut, female genital
<i>A. odontolyticus</i>		
<i>Nocardia</i>		
<i>N. asteroides</i>	E	OI – lung, brain
<i>N. nova</i>		
<i>N. farcinica</i>		
<i>N. otidiscaviarum</i>		
<i>N. brasiliensis</i>	E	Cutaneous infections
<i>N. transvalensis</i>		
<i>Corynebacteria</i>		
<i>C. diphtheriae</i>	H	Diphtheria (throat, larynx, myocarditis, neuropathy)
<i>C. urealyticum</i>	End	Chronic urinary tract infection
<i>C. jeikeium</i>	End	NI, FB

(continued)

(continued)

Pathogens	Usual source	Clinical syndrome
Many others	End	NI, FB
<i>Arcanobacterium haemolyticum</i>	H	Pharyngitis, scarlatiniform rash
<i>Rhodococcus equi</i>	E	OI – pneumonia
<i>Listeria monocytogenes</i>	E, Mat	Bacteremia, meningitis, newborn, elderly, OI
<i>Erysipelothrix rhusiopathiae</i>	A, E	Cellulitis, IE
<i>Gardnerella vaginalis</i>	End	Bacterial vaginosis
<i>Lactobacillus</i> sp.	End	
<i>Rothia denticariosa</i>	End	
<i>Streptomyces somaliensis</i>	E	Mycetoma
<i>Oerskovia</i> spp.	E	IE
<i>Tropheryma whipplei</i>	unknown	Whipple's disease
MYCOBACTERIA		
<i>M. tuberculosis</i>	H	Tuberculosis
<i>M. bovis</i>	A	Tuberculosis
<i>M. africanum</i>	H	Tuberculosis
<i>M. leprae</i>	H	Leprosy
<i>M. avium</i> complex	E	Lymphadenitis, OI
<i>M. marinum</i>	E	Water-associated granuloma
<i>M. scrofulaceum</i>	E	Lymphadenitis
Many other species	E	Focal infections, OIs
Gram-positive bacilli, spore-forming, aerobic		
BACILLUS		
<i>B. anthracis</i>	A, E	Anthrax
<i>B. cereus</i>	E	Acute food poisoning, eye
Several other species		FB, OI
Anaerobic bacteria		
GRAM-POSITIVE RODS, SPORE-FORMING (CLOSTRIDIA)		
<i>C. tetani</i>	E	Tetanus
<i>C. botulinum</i>	E	Botulism – classic, wound, infant
<i>C. difficile</i>	E, H	Antibiotic-associated colitis (NI)
<i>C. perfringens</i>	E	Food poisoning
	End	Myonecrosis (gas gangrene), hemolysis
Other histotoxic clostridia		Myonecrosis, OI

(continued)

Table A2: Taxonomy of human pathogens, their usual sources, and the main clinical syndromes they cause (continued)

Pathogens	Usual source	Clinical syndrome
ANAEROBIC GRAM-POSITIVE RODS, NON-SPOREFORMING		
<i>Bifidobacterium</i> spp.	End	Dental plaque
<i>Eubacterium</i> spp.	End	
<i>Actinomyces</i> spp.	see aerobic Gram-positive rods, above	
ANAEROBIC GRAM-NEGATIVE BACILLI		
<i>Bacteroides</i>	End	Infections arising from mucosae: mouth, gut, female genital tract
<i>Prevotella</i> species	End	As above
<i>Porphyromonas</i> species	End	As above
<i>Fusobacterium</i> species	End	Infections from mucosae, septic thrombophlebitis
ANAEROBIC GRAM-NEGATIVE COCCI		
<i>Veillonella</i>	End	As above
ANAEROBIC GRAM-POSITIVE COCCI		
<i>Peptococcus</i> species	End	As above
<i>Peptostreptococcus</i> species	End	As above
FUNGI		
YEASTS		
<i>Candida</i> species	End	Mucosal infections – mouth, vagina, NI, FB, fungemia, esophagitis, OI
<i>C. albicans</i>		
<i>C. tropicalis</i>	End	
<i>C. pseudotropicalis</i>		
<i>C. parapsilosis</i>		
<i>C. krusei</i>		
<i>C. (Torulopsis) glabrata</i>		
<i>C. lusitaniae</i>	E	OI – pneumonia, meningitis
<i>C. guilliermondii</i>		
<i>Cryptococcus neoformans</i>		
<i>Rhodotorula rubra</i>	E	OI
<i>Malassezia furfur</i>	End	Pityriasis versicolor, NI-(catheter-related)
DIMORPHIC FUNGI		
<i>Histoplasma capsulatum</i>	E	Histoplasmosis: pneumonia (bat, bird guano), OI
<i>H. capsulatum</i> var. <i>duboisii</i>	E	African histoplasmosis

(continued)

(continued)

Pathogens	Usual source	Clinical syndrome
<i>Coccidioides immitis</i>	E	Coccidioidomycosis: fever, pneumonia, OI
<i>Blastomyces dermatitidis</i>	E	Blastomycosis: pneumonia, skin
<i>Paracoccidioides brasiliensis</i>	E	South American blastomycosis
Other fungi		
<i>Sporothrix schenckii</i>	E	Sporotrichosis (cutaneous, lymphangitis)
<i>Pseudoallescheria boydii</i>	E	OI
Filamentous fungi		
<i>Aspergillus fumigatus, terrae, flavus, niger</i>	E	Aspergillosis – lung, OI, angioinvasive
<i>Zygomycetes</i>		
<i>Mucor</i>	E	Mucormycosis – OI, angioinvasive
<i>Rhizopus</i>	E	Mucormycosis
<i>Absidia</i>	E	Mucormycosis
<i>Cunninghamella</i>	E	Mucormycosis
<i>Penicillium marneffei</i>	E	OI – SE Asia
Dermatophytes		
<i>Trichophyton</i> species	H	Tinea corporis, capitis
<i>Epidermophyton</i> species	H	Tinea corporis, capitis
<i>Microsporum</i> species	H, A	Tinea corporis, capitis
There are many other fungi, living in the environment, and belonging to many families, that can cause infections as a result of injury or immunodeficiency in the host.		
PARASITES		
Intestinal Protozoa		
AMEBAE		
<i>Entamoeba histolytica</i>	H	Amebiasis: colitis, liver abscess
FLAGELLATES		
<i>Giardia lamblia (intestinalis)</i>	H, A	Diarrhea
SPOROZOA		
<i>Cryptosporidium parvum</i>	H, A	Diarrhea
<i>Cyclospora cayetanensis</i>	H	Diarrhea
<i>Isospora belli</i>	H	Diarrhea – OI

(continued)

Table A2: Taxonomy of human pathogens, their usual sources, and the main clinical syndromes they cause (continued)

Pathogens	Usual source	Clinical syndrome
MICROSPORIDIA		
<i>Enterocytozoon bieneusi</i>		OI – diarrhea, cholangitis, sinusitis
<i>Encephalitozoon intestinalis</i>		Diarrhea, OI – diarrhea, cholangitis
CILIATES		
<i>Balantidium coli</i>	H, A	Colitis
OTHER		
<i>Blastocystis hominis</i>	H	Diarrhea
Tissue and Blood Protozoa		
AMEBAE		
<i>Naegleria fowleri</i>	E	Meningoencephalitis (fresh water)
<i>Acanthameba</i>	E	Meningoencephalitis, keratitis
<i>Balamuthia mandrillaris</i>	E	Meningoencephalitis
FLAGELLATES		
<i>Leishmania donovani, infantum</i>	Fly	Visceral leishmaniasis
<i>Leishmania mexicana, tropica</i>	Fly	Cutaneous leishmaniasis
<i>Leishmania braziliensis</i>	Fly	Mucocutaneous leishmaniasis
Trypanosomes		
<i>T. cruzi</i>	reduviid bug	S. American trypanosomiasis (Chaga's disease)
<i>T. brucei rhodesiense</i>	Fly	African trypanosomiasis (sleeping sickness)
<i>T. brucei gambiense</i>		African trypanosomiasis
<i>Trichomonas vaginalis</i>	H	Vaginitis
SPOROZOA		
<i>Plasmodium falciparum</i>	Mos	Falciparum malaria (malignant tertian)
<i>Plasmodium vivax</i>	Mos	Vivax malaria (benign tertian)
<i>Plasmodium ovale</i>	Mos	Ovale malaria
<i>Plasmodium malariae</i>	Mos	Quartan malaria
<i>Babesia microti</i> and other species	Tick	Babesiosis
<i>Toxoplasma gondii</i>	A, Mat	Toxoplasmosis: mononucleosis, congenital, OI – encephalitis
MICROSPORIDIA		
<i>Brachiola (Nosema) algerae</i>		Keratoconjunctivitis, skin infection
<i>Nosema oculorum</i>		Keratitis
<i>Vittaforma corneae</i>		Keratitis, disseminated infection

(continued)

(continued)

Pathogens	Usual source	Clinical syndrome
<i>Microsporidium ceylonensis</i>		Keratitis
<i>Microsporidium africanum</i>		Keratitis
<i>Nosema connori</i>		OI – disseminated infection
<i>Encephalitozoon hellem</i>		OI – multiple sites
INTESTINAL WORMS		
ROUNDWORMS (NEMATODES)		
<i>Enterobius vermicularis</i> (Pinworm)	H	Perianal itching
<i>Ascaris lumbricoides</i>	H-E	Ascariasis, visceral larva migrans, intestinal obstruction
<i>Toxocara canis</i> and <i>cati</i>	A-E	Visceral larva migrans, pneumonitis, ocular granuloma
<i>Baylisascaris procyonis</i>	A-E	Encephalitis
Hookworms		
<i>Necator americanus</i>	H-E	Anemia, hypoalbuminemia
<i>Ancylostoma duodenale</i>	H-E	Anemia, hypoalbuminemia
<i>Strongyloides stercoralis</i>	H-E	Diarrhea, OI – visceral disease
<i>Trichuris trichiura</i>	H-E	Chronic diarrhea, rectal prolapse
<i>Anisakis</i> spp.	A	Gastritis (raw fish)
CESTODES (TAPEWORMS)		
<i>Taenia solium</i>	Pig	Tapeworm
<i>Taenia solium</i> larval stage	H	Cysticercosis
<i>Taenia saginata</i>	Beef	Tapeworm
<i>Echinococcus multilocularis</i>	A-E-H	Cysts
<i>Multiceps multiceps</i>	A	Cysts
<i>Echinococcus granulosus</i>	A	Hydatid disease (larval stage)
<i>Diphyllobothrium latum</i> (fish)	Fish	Vitamin B12 deficiency, intestinal obstruction
<i>Spirometra mansonoides</i>	Water, copepod	Eye, visceral disease
<i>Dipylidium caninum</i>		Flea
<i>Hymenolepis nana</i>	H (+/- arthropod)	Anorexia
TREMATODES (FLUKES)		
<i>Fasciolopsis buski</i>	Water plants	Intestinal disease
<i>Heterophyes heterophyes</i>	Fish	Intestinal disease
<i>Metagonimus yokagawai</i>	Fish	Intestinal disease
TISSUE AND BLOOD WORMS		
TREMATODES (FLUKES)		
<i>Paragonimus westermanii</i>	Crustacean	Lung disease
<i>Fasciola hepatica</i>	Sheep-water plants	Liver
<i>Clonorchis sinensis</i>	Fish	Biliary tract
<i>Opisthorcis viverrini</i>	Fish	Biliary tract

(continued)

Table A2: Taxonomy of human pathogens, their usual sources, and the main clinical syndromes they cause (continued)

Pathogens	Usual source	Clinical syndrome
Schistosomes		
<i>S. mansoni</i>	H-W	Liver fibrosis
<i>S. japonicum</i>	H-W	Liver fibrosis
<i>S. haematobium</i>	H-W	Urinary tract disease
<i>S. mekongi</i>	H-W	Liver fibrosis
NEMATODES		
<i>Wucheraria bancrofti</i>	Mos	Filariasis, elephantiasis
<i>Brugia malayi</i>	Mos	Filariasis
<i>Onchocerca volvulus</i>	Fly	River blindness, subcutaneous nodules
<i>Loa loa</i>	Fly	Subcutaneous nodules, conjunctival disease
<i>Dracunculus medinensis</i> (guinea worm)	W (Copepod)	Skin ulcer, cellulitis
<i>Trichinella spiralis</i>	A	Trichinosis (myositis)
<i>Ancylostoma braziliense</i> and <i>caninum</i>	A-E	Cutaneous larva migrans
<i>Gnathostoma spinigerum</i>	Fish	Subcutaneous, visceral swellings
<i>Angiostrongylus cantonensis</i>	Crustacian	Eosinophilic meningitis
<i>Angiostrongylus costaricensis</i>	Slug	Abdominal granulomas
ECTOPARASITES		
Mites		
<i>Sarcoptes scabiei</i>	H	Scabies
<i>Eutrombicula alfreddugesi</i> (chigger)	E	Dermatitis
Lice		
<i>Pediculus humanus capitis</i>	H	Head pediculosis
<i>Pediculus humanus corporis</i>	H	Body pediculosis
<i>Phthyrus pubis</i> (crab louse)	H	Pubic pediculosis
Flies		Myiasis
Calliphoridae		
Cuteribridae		
Sarcophagidae		

A = animal, including birds; E = environment; End = endogenous; FB = infections on foreign bodies, such as catheters; H = human; IE = infective endocarditis; L = laboratory; Mat = maternal; Mos = mosquito; NI = nosocomial infection; OI = opportunistic infection; URI = upper respiratory tract infection; UTI = urinary tract infection.

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List 1: Animate and inanimate sources and infectious agents associated with them.

Abbreviations: CMV = cytomegalovirus; HAV = hepatitis A virus; HBV = hepatitis B virus; HIV = human immunodeficiency virus; HSV = herpes simplex virus; VZV = varicella zoster virus

Human (close (other than sexual) contact, droplet)

Viruses: all herpes viruses, pox viruses, parvovirus, adenovirus, HBV, respiratory viruses, enteroviruses, rhinoviruses, coronaviruses, diarrhea-causing viruses, mumps, measles, papilloma viruses

Chlamydia pneumoniae, *Mycoplasma pneumoniae*

Streptococcus pyogenes, *Streptococcus pneumoniae*, *Streptococcus agalactiae*, *Staphylococcus aureus*, *Neisseria meningitidis*, *Haemophilus influenzae*, *Corynebacterium diphtheriae*, *Arcanobacterium haemolyticum*, *Mycobacterium tuberculosis*, *Mycobacterium leprae*, Shigellae, *Bordetella pertussis*, Dermatophytes, Lice, *Sarcoptes scabiei*

Human (sexual contact)

HSV, CMV, HIV, HBV, papilloma virus, *Chlamydia trachomatis*, *Treponema pallidum*, *Calymmatobacterium granulomatis*, *Haemophilus ducreyi*, *Neisseria gonorrhoeae*, *Trichomonas vaginalis*.

Human (endogenous)

staphylococci, streptococci (especially viridans group), enterococci, *Moraxella catarrhalis*, *Corynebacterium* spp., *Actinomyces* spp., many Enterobacteriaceae, for example, *E. coli*, *Haemophilus* spp., many anaerobic bacteria, *Candida* spp.

Ingestion (human feces-contaminated food, water or soil)

HAV, enteroviruses, diarrhea-causing viruses, *Salmonella*, *Shigella*, *Entamoeba histolytica*, *Giardia lamblia*, *Cryptosporidium parvum*, *Taenia solium* (cysticercosis), intestinal flukes, *Ascaris lumbricoides*, hookworms, *Strongyloides stercoralis*, *Enterobius vermicularis*

Ingestion (animal excreta-contaminated food, water, or soil)

Listeria monocytogenes, *Brucella* spp., *E. coli*, *Salmonella*, *Campylobacter*, *Yersinia enterocolitica*, *Mycobacterium bovis* (milk), *Giardia lamblia*, *Cryptosporidium parvum*, *Toxocara*, *Baylisascaris procyonis*, *Echinococcus granulosus*, *Toxoplasma gondii*

Ingestion (uncooked animal tissue)

Bacillus anthracis, *Salmonella* spp., *E. coli*, *Campylobacter*, *Yersinia enterocolitica*, *Toxoplasma gondii*, *Taenia solium* (pig), *Taenia saginata* (beef), *Hymenolepis diminuta* (rat fleas), *Diphyllobothrium latum* (fish), *Trichinella spiralis* (pig, carnivore), *Anisakis* (fish), *Paragonimus* spp., *Gnathostoma spinigerum*.

■ **Table A3: Arthropods as vectors of infectious agents.**

	Viruses	Bacteria	Protozoa	Helminths
Mosquito	Alphaviruses Flaviviruses		Plasmodia	<i>Wucheraria</i> <i>Brugia</i> <i>Dirofilaria</i>
Tick	Colorado tick Congo-Crimean	<i>Rickettsia</i> <i>Coxiella burnetii</i> <i>Ehrlichia</i> <i>Borrelia duttoni</i> <i>Borrelia burgdorferi</i> <i>Francisella</i>	Babesia	
Flea		<i>Yersinia pestis</i> <i>Rickettsia typhi</i>		
Louse		<i>Borrelia recurrentis</i> <i>Rickettsia prowazeki</i> <i>Bartonella quintana</i>		
Fly	Sandfly fever		<i>Leishmania</i> <i>Trypanosoma brucei</i> <i>Trypanosoma cruzi</i>	<i>Onchocerca</i> <i>Loa loa</i>
Triatomid bug				
Mite		<i>Rickettsia akari</i>		

Animal (contact with animal, animal tissue, animal excreta, bite, scratch)

Ebola virus, Congo-Crimean Hemorrhagic Fever, Rift Valley Fever, Lymphocytic choriomeningitis virus, Lassa fever, Hantaviruses, Rabies virus, *Herpes simiae*, *Chlamydia psittaci*, *Bacillus anthracis*, *Erysipelothrix rhusiopathiae*, *Streptobacillus moniliformis*, *Spirillum minus*, *Pasteurella multocida*, *Yersinia pestis*, *Francisella tularensis*, *Capnocytophaga canimorsus*, *Coxiella burnetii*, *Brucella* spp., *Bartonellae henselae*, *Microsporum canis*

Fresh water (non-enteral exposure)

Leptospira, *Erysipelothrix rhusiopathiae*, *Mycobacterium marinum*, *Legionella* spp., *Naegleria fowleri*, *Acanthamoeba*, *Schistosomes*, *Pseudomonas aeruginosa*, *Aeromonas hydrophila*

Sea water (enteral)

Vibrio cholerae, *Vibrio parahaemolyticus*, *Vibrio vulnificus*

Sea water (non-enteral)

Erysipelothrix rhusiopathiae, *M. marinum*, halophilic vibrios

Inanimate environment (soil, air)

Bacillus cereus, *Rhodococcus equi*, nontuberculous mycobacteria, *Nocardia* spp., Enterobacteriaceae, *Aeromonas hydrophila*, *Chromobacterium violaceum*, *Pseudomonas* spp., *Acinetobacter* spp., *Elizabethkingae meningosepticum*, *Legionella* spp., *Clostridium perfringens*, *Clostridium tetani*, dimorphic fungi, *Cryptococcus neoformans*, *Aspergillus* spp., other filamentous fungi.

Inanimate (human-contaminated)

Enteroviruses, respiratory viruses, *Mycobacterium tuberculosis*, Enterobacteriaceae, hookworms, *Strongyloides*, *Ascaris*

Mother

Rubella, CMV, HSV, HBV, HCV, HIV, VZV, enterovirus, lymphocytic choriomeningitis virus, *Treponema pallidum*, *Chlamydia trachomatis*, *Streptococcus agalactiae*, *Listeria monocytogenes*, Enterobacteriaceae, *Mycobacterium tuberculosis*, *Toxoplasma gondii*, *Plasmodium* spp.

Hospital

VZV, respiratory viruses, enteroviruses, diarrhea-causing viruses, measles, filoviruses, Enterobacteriaceae, *Pseudomonas aeruginosa*, *Stenotrophomonas maltophilia*, *Acinetobacter* spp., *Staphylococcus aureus*, coagulase-negative staphylococci, enterococci, *Clostridium difficile*, *Candida* spp., *Aspergillus* spp.

Blood

HBV, HCV, HIV, CMV, EBV, West Nile virus, *Treponema pallidum*, *Yersinia enterocolitica*, *Listeria monocytogenes*, Trypanosomes, *Plasmodium* spp., *Babesia* spp., *Toxoplasma gondii*, Filaria

Tissue

Prions, CMV, HIV, West Nile virus, rabies virus, contaminating bacteria

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Index

- Abdominal distension, 284
- Abdominal mass, hydatid disease, 287
- Abdominal pain, 12, 79, 112, 158, 169, 177, 257, 270, 273, 304, 308
 - acute HIV infection, 311
 - appendicitis, 253
 - cat-scratch disease, 143
 - hydatid disease, 287
 - leptospirosis, 139
 - pelvic inflammatory disease, 319
 - peritonitis, 315
 - pneumonia, 248
 - pyelonephritis, 244
 - typhus, 149
- Abdominal tenderness, 257
- Abscess
 - appendix, 159, 255
 - brain, 43, 66, 255
 - meningitis, 108
 - seizure, 195
 - Brodie's, 282
 - cerebellar, 327
 - dental, 271
 - epidural, 334
 - eye, 314
 - lateral pharyngeal, 264
 - liver, 13, 174
 - amebic, 174
 - lung, 249
 - odontogenic *See* Abscess
 - dental
 - pelvic, 306
 - peritonsillar, 100, 264
 - Lemierre syndrome, 332
 - perivenous, 240
 - psoas
 - tuberculosis, 226
 - retropharyngeal, 100, 264, 318
 - tuberculosis, 226
 - spinal epidural, 273
 - spinal
 - paraplegia, 262
 - subphrenic, 232
 - tubo-ovarian, 322
- Absidia*, taxonomy, 347
- Acanthameba*, taxonomy, 348
- Acetaminophen, 15, 54
- Acetylcholine, 129
- Acid-fast bacilli *See* Mycobacteria
- Acidosis, diarrhea, 80
- Acinetobacter*, taxonomy, 342
- Acquired immunodeficiency syndrome, 146, 193, 210, 242
 - reportable, 336
- Actinobacillus actinomycetemcomitans*, 153
 - taxonomy, 343
 - infective endocarditis, 297
- Actinomyces, 124, 127
 - taxonomy, 344
- Actinomyces israelii*, 125
- Actinomyces odontolyticus*, taxonomy, 344
- Actinomycosis, 125
- Acute chest syndrome, sickle cell disease, 249, 324
- Acyclovir, 42
 - Epstein-Barr virus, 39
 - herpes simplex virus, 29
 - varicella, 47

- Adenovirus, 5, 17, 20, 55, 63, 100, 159, 230, 310
 enteric
 gastroenteritis, 79
 pneumonia, 250
 taxonomy, 338
- Adnexal tenderness, pelvic inflammatory disease, 320
- Adrenal disease, enterovirus, 29
- Adrenal failure, malaria, 184
- Adrenal gland, hemorrhage, 92
- Aedes aegypti*, 77
- Aerogenous spread, 247
- Aeromonas hydrophila*, taxonomy, 343
- Africa, 41, 77, 181, 190, 198
 relapsing fever, 138
- African Tick Bite Fever *See* Rickettsia africae
- African trypanosomiasis *See* Trypanosoma brucei
- AIDS *See* Acquired immunodeficiency syndrome
- Albendazole
 ascariasis, 208
 hookworm, 203
 hydatid disease, 290
 microsporidiosis, 180
 neurocysticercosis, 42, 197
 visceral larva migrans, 202
- “Alice-in-Wonderland” syndrome, 36
- Allergy, milk-protein, 83
- Alpha-1-antitrypsin deficiency, 157
- Alphaherpesvirinae, taxonomy, 337
- Alphaviruses, taxonomy, 339
- Altered mental status, 21, 42, 104
 diarrhea, 80, 81
 infective endocarditis, 297
 intracranial suppuration, 255
 meningitis, 105, 123, 274
 neonate, 268
 pertussis, 161
 pneumonia, 248
 suppurative pericarditis, 270
 tuberculous meningitis, 209
 typhoid fever, 155
 typhus, 151
 VP shunt malfunction, 112
- Amebae, taxonomy, 347
- Amebiasis *See* Entamoeba histolytica
- Ameboma, 175
- American Heart Association, infective endocarditis, 299
- Amikacin
 endocarditis, 103
 endophthalmitis, 314
 Nocardia, 127
 nosocomial infection, 166
 vascular catheter infection, 277
- Aminoglycoside
 Pseudomonas aeruginosa, 172
 resistance, 277
- Amoxicillin/clavulanate, dental infection, 272
- Amoxicillin/clavulanic acid, 158
- Amoxicillin
 infectious mononucleosis, 155
 Kingella kingae, 154
 Lyme disease, 143
 pneumonia, 247
 salmonella, 171
 Streptococcus pyogenes, 101
 typhoid fever, 156
- Amphotericin B, 240
 drug fever, 335
 endocarditis, 103
 fungal infection, 42
 mucormycosis, 243
 Naegleria infection, 42
 nosocomial infection, 166
- Ampicillin resistance
 Haemophilus influenzae, 277
 enterococci, 277
- Ampicillin/sulbactam
 dental infection, 272
 Lemierre syndrome, 333
- Ampicillin
 infectious mononucleosis, 155
 Kingella kingae, 154
 Listeria monocytogenes, 123
 meningitis, 63
 osteomyelitis, 111
 shigella, 86
 urinary tract infection, 245
- Amyloidosis, 305
- Anaerobic bacteria, 159, 254
 brain abscess, 327
 dental infection, 272
 intracranial suppuration, 256
 lung abscess, 249
 neck abscess, 261
 pelvic inflammatory disease, 322
 retropharyngeal abscess, 264
- Anaphylaxis, ruptured hydatid cyst, 290
- Anaplasma phagocytophilum, 76, 147
 taxonomy, 341
- Anasarca, nephrotic syndrome, 315
- Ancylostoma braziliense, 203
 taxonomy, 350
- Ancylostoma duodenale, 203
- Andes Mountains, 146

- Anemia, 173, 257
 congenital rubella, 71
 congenital syphilis, 133
 hemolytic, 13, 61, 132, 258, 324
 infectious mononucleosis, 36
 malaria, 183
 microangiopathic, 80
 Mycoplasma, 252
 Oroya fever, 146
 human immunodeficiency virus infection, 65, 66
 microcytic
 hookworm, 203
 neonate, 268
 pneumonia, 323
 sickle cell disease, 59
 tuberculosis, 233
- Angiostrongylus cantonensis*, taxonomy, 350
Angiostrongylus costaricensis, taxonomy, 350
Anisakis, taxonomy, 349
Anopheles, 182
Anorexia, 12, 112, 123, 157, 263
Anterior horn cell, 31, 74, 128
Anthrax, 190 *See also Bacillus anthracis*
 reportable, 336
Antibiotic resistance, 171, 276
 Salmonella typhi, 156
 Streptococcus pneumoniae, 275
Antibody, 117
Anticholinergic drugs, drug fever, 335
Antidiphtheria globulin, 122
Antidiuretic hormone, inappropriate secretion, 109
Antigenic variation, 138
Antimicrobial
 resistance, 8
 spectrum, 9
 susceptibility, 8
 therapy, 2, 6
Antipruritic, 47
Antipyretic, 47
Antiretroviral therapy, 42, 62, 221
 highly active, 223
Antiviral therapy, 15, 29
Aortitis, syphilis, 136, 153
Aplasia cutis, 27
Apnea, pertussis, 161
Appendicitis, 159, 253, 320
 liver abscess, 174
Arachnoiditis, tuberculous meningitis, 210
Arboviruses, 42, 139, 293
Arcanobacterium haemolyticum, 100, 310
 taxonomy, 345
Arenaviridae, 41, 76
 taxonomy, 340
Argentinian hemorrhagic fever, 41
 virus, 76
Artemether-lumefantrine, 188
Arteriovenous malformation
 seizure, 195
 spinal, 263
Arthralgia, 75
 hepatitis, 14
Arthritis
 Familial Mediterranean fever, 305
 Kawasaki disease, 231
 Lyme disease, 141
 migratory, 95
 Neisseria meningitidis, 92
 rat bite fever, 291
 reactive, 109, 141
 gastroenteritis, 82
 rheumatoid, 141
 septic, 74, 80, 102, 110, 141, 170, 266, 293
 Haemophilus influenzae, 164
 Kingella kingae, 153
 salmonella, 80
 syphilis, 133
Ascaris lumbricoides, 202, 207, 251
 taxonomy, 349
Ascites, nephrotic syndrome, 315
Asia, southeast, 198
Aspergillus, 125
 ischemic necrosis, 242
 pneumonia, 250
 taxonomy, 347
Aspiration, 111
 diagnostic, 261, 267
 spinal infection, 273
Aspirin, 231
Asplenia, 88
Asthma, 25, 67, 160, 279, 317
Astrovirus, 79
 taxonomy, 339
Ataxia
 cerebellar, 51
 infectious mononucleosis, 36
Atelectasis, 249
Atrial myxoma, 298
Attack rate, 18
Autoantibody, pneumonia, 324
Autoimmune lymphoproliferative syndrome, 310
Autoimmunity, 35
Autonomic dysfunction, 73
Avascular necrosis, hip, 306

- Azithromycin
 - cat-scratch disease, 144
 - cystic fibrosis, 168
 - gastroenteritis, 86
 - onchocerciasis, 207
 - pertussis, 161
- B-lymphocyte, 35
- Babesia*, 6, 76, 138
 - blood transfusion, 173
 - taxonomy, 348
- Babinski response, tuberculous meningitis, 209
- Bacillary angiomatosis, 146
- Bacilli
 - Gram-negative, 5, 87, 102, 125, 153, 158, 164, 167, 173, 240, 250, 254, 270, 322
 - brain abscess, 327
 - ecthyma gangrenosum, 172
 - infective endocarditis, 295
 - intracranial suppuration, 256
 - nosocomial infection, 166
 - osteomyelitis, 111
 - peritonitis, 315
 - pneumonia, 250
 - Gram-positive, 124
- Bacillary angiomatosis, 66
- Bacilli, enteric, 5, 20, 268
- Bacillus anthracis*, 131
 - taxonomy, 345
- Bacillus cereus*
 - endophthalmitis, 314
 - intoxication
 - food, 78
 - taxonomy, 345
- Backache
 - poliomyelitis, 31
 - viral hemorrhagic fever, 41
- Bacteremia, 28, 48, 102, 104, 158
 - abscess, 306
 - anthrax, 131
 - ecthyma gangrenosum, 172
 - endocarditis prophylaxis, 96
 - gastroenteritis, 80
 - Haemophilus influenzae*, 164
 - human immunodeficiency virus infection, 66
 - infective endocarditis, 294
 - liver abscess, 174
 - Neisseria meningitidis*, 89, 147
 - neonate, 268
 - nosocomial infection, 165
 - polymicrobial, 159
 - salmonella, 170
 - sickle cell disease, 59
 - Streptococcus agalactiae*, 113
 - Streptococcus pneumoniae*, 114
 - Streptococcus pyogenes*, 100
 - suppurative pericarditis, 270
 - tuberculous, 219
 - typhoid fever, 155
 - urinary tract infection, 245
- Bacterial vaginosis, 321
- Bacteroides*
 - brain abscess, 327
 - taxonomy, 346
- Bacteruria, 13
- Balamuthia mandrillaris*, taxonomy, 348
- Balantidium coli*, 79, 180
 - taxonomy, 348
- Bartonella*, infective endocarditis, 297
- Bartonella bacilliformis*, 6, 146
 - taxonomy, 343
- Bartonella elizabethae*, taxonomy, 343
- Bartonella henselae*, 144, 163
 - taxonomy, 343
- Bartonella quintana*, taxonomy, 343
- Bat, 44
- Baylisascaris procyonis*, 42, 202
 - taxonomy, 349
- Beta-lactam
 - drug fever, 335
 - Pseudomonas aeruginosa*, 172
 - resistance, 277
- Beta-lactamase, 277
 - extended-spectrum, 277
 - Kingella kingae*, 154
- Betaherpesvirinae, taxonomy, 338
- Bifidobacterium*, taxonomy, 346
- Biliary atresia, 157
- Biliary obstruction, 157
- Biliary sludge, 309
- Bilirubin, 13
- Bilirubinuria, 13
- Biopsy, 243
 - bone, 281
 - lung, 64, 126, 247
- Bioterrorism, anthrax, 131
- Bird fancier's disease *See* Pneumonia
 - hypersensitivity
- Bite
 - animal, 42
 - arthropod, 300
 - insect, 46, 75, 189
 - snake, 74
- Bite cell, 260, 325
- Bladder catheterization, 245
- Bladder puncture, suprapubic, 245

- Blastocystis hominis*, taxonomy, 347
Blastomyces dermatitidis
 pneumonia, 251
 taxonomy, 347
 Blindness, 313
 onchocerciasis, 206
 Blisters, 44
 Blood glucose, hepatitis, 14
 Blood smear, 40, 114, 140, 181, 190, 259
 hemolytic anemia, 325
 relapsing fever, 139
 Blood transfusion, 14, 172
 exchange, 284
 infections, 173
 Bolivian hemorrhagic fever, 41
 virus, 76
 Bone disease, congenital rubella, 71
 Bone lucency, 281
Bordetella pertussis, 159
 taxonomy, 342
 Borneo, 139
Borrelia, 6
 relapsing fever, 76, 138
Borrelia burgdorferi, 42, 141
 taxonomy, 341
Borrelia duttoni, taxonomy, 341
Borrelia hermsii, taxonomy, 341
Borrelia recurrentis, taxonomy, 341
 Botswana, 189
 Botulism, 31, 74, 128
 reportable, 336
 Brain abscess, otitis media, 327
 Brain disease, 28, 31, 40, 42, 54, 104, 191, 193
 congenital rubella, 71
 intracranial suppuration, 255
 Lyme disease, 142
 measles, 21
 meningitis, 106
 mucormycosis, 243
 Nocardia, 127
 Rocky Mountain spotted fever, 147
 syphilis, 136, 153
 toxoplasmosis, 193
 tuberculous meningitis, 209
 Brainstem, poliomyelitis, 32
 Branchial cleft cyst, 237, 261
 Brazil, 76
 Breath sounds, 67
 Brill-Zinsser disease *See* Rickettsia prowazekii
 Bronchial obstruction, 216, 249
 Bronchiectasis, 19
 cystic fibrosis, 168
 pertussis, 161
 Bronchiolitis, 24, 249, 317
 Bronchiolitis obliterans, 19
 Bronchitis, 19, 67
 pertussis, 161
 Bronchoalveolar lavage, 64, 126, 247
 Bronchodilator, 26
 Bronchomalacia, 68
 Bronchopulmonary dysplasia, 26
 Brucella
 intramacrophage survival, 123
 taxonomy, 342
 Brucellosis, 154, 233
 reportable, 336
Brugia malayi, taxonomy, 350
 Buccal space, 272
 Bulbar *See* Brainstem
 Bulla, 131
 Bullous disease, 27
 Bunyaviridae, 41, 76
 taxonomy, 340
Burkholderia cepacia
 cystic fibrosis, 168
 pneumonia
 cystic fibrosis, 250
 taxonomy, 342
Burkholderia pseudomallei, taxonomy, 342
 Calcification
 basal ganglia, 62
 cerebral, 62
 pulmonary, 47
 Caliciviridae, taxonomy, 339
 Calliphoridae, taxonomy, 350
Calymmatobacterium granulomatis, taxonomy, 342
Campylobacter fetus, taxonomy, 342
Campylobacter jejuni, 79, 170
 taxonomy, 342
 Cancer chemotherapy, pneumonia, 250
Candida, 5, 27, 66, 70, 102, 240
 infective endocarditis, 297
 nosocomial infection, 166
 taxonomy, 346
 Candidiasis
 oral
 acute HIV infection, 312
Capnocytophaga canimorsis, taxonomy, 342
 Carbamazepine, 53
 Carbapenem
 clostridial myonecrosis, 132
 cystic fibrosis, 168
 Carcinoma
 hepatocellular, 14
 nasopharyngeal, 37

- Cardiac tamponade, 270
 - infective endocarditis, 299
- Cardiobacterium hominis*, 154
 - infective endocarditis, 297
 - taxonomy, 343
- Cardiomyopathy, human immunodeficiency virus infection, 65
- Carditis, 95
- Caries, 271
- Case fatality rate, 18, 77, 175
 - Rocky Mountain spotted fever, 148
 - relapsing fever, 138
- Cat-scratch disease, 144, 163, 238, 261
- Cataract, 71
- Catheter
 - urinary, 298
 - nosocomial infection, 165
 - vascular, 5, 58, 102, 109, 173, 240, 242, 277, 298
 - nosocomial infection, 165
- Cefixime, 246
- Cefotaxime
 - brain abscess, 328
 - epiglottitis, 319
 - meningitis, 106
 - urinary tract infection, 246
- Cefotetan, pelvic inflammatory disease, 323
- Cefoxitin, pelvic inflammatory disease, 322
- Ceftazidime, 172
 - cystic fibrosis, 168
 - endocarditis, 103
 - Pseudomonas aeruginosa*, 295
- Ceftriaxone, 114
 - brain abscess, 328
 - drug fever, 335
 - epidural abscess, 334
 - epiglottitis, 319
 - gastroenteritis, 86
 - infective endocarditis, 299
 - Lyme disease, 42, 143
 - meningitis, 106, 212, 308
 - Neisseria gonorrhoeae*, 94
 - Neisseria meningitidis*, 88, 148
 - typhoid fever, 156
 - urinary tract infection, 246
- Cell-mediated immunity, 47, 63
- Cellulitis, 48, 275
 - buccal, 164
 - orbital, 256
 - periorbital, 164
- Centers for Disease Control and Prevention, 41, 44, 182
- Central America, 206
- Cephalexin, 50
 - neck abscess, 261
 - skeletal infection, 267
- Cephalosporin
 - first-generation, 267
 - Nocardia*, 127
 - pneumonia, 325
 - Streptococcus pyogenes*, 101
 - third-generation, 158, 173
 - abdominal infection, 254
 - endocarditis, 103
 - Haemophilus influenzae*, 164
 - infective endocarditis, 295
 - intracranial suppuration, 256
 - Neisseria meningitidis*, 88
 - osteomyelitis, 111
 - peritonitis, 316
 - pneumonia, 247, 330
 - retropharyngeal abscess, 264
 - salmonella, 171
 - urinary tract infection, 246
- Cephazolin, 104
- Cercaria, 198
- Cerebral atrophy, 62
- Cerebral venous sinus thrombosis, 257
- Cerebritis, usage of term, 43
- Cerebrospinal fluid, 26, 42, 52, 105, 112, 123, 140, 163, 210, 274, 278, 325
 - congenital syphilis, 137
 - pleocytosis, 28
- Ceruloplasmin, 15
- Cestodes, taxonomy, 349
- Chancroid, reportable, 336
- Chemoprophylaxis, 165
- Chemotherapy, 39
- Chickenpox *See* Varicella
- Chlamydia*, 6
- Chlamydia pneumoniae*, 55, 63, 125, 159
 - pneumonia, 250, 329
 - taxonomy, 340
- Chlamydia psittaci*, 329
 - pneumonia, 250
 - taxonomy, 340
- Chlamydia trachomatis*
 - neonatal conjunctivitis, 94
 - neonatal pneumonia, 250
 - pelvic inflammatory disease, 321
 - reportable, 336
 - taxonomy, 340
- Chlamydiae, taxonomy, 340
- Chloramphenicol, 9
 - typhoid fever, 156
- Chloroquine, 188

- Cholangitis, 13
 liver abscess, 174
 Cholecystectomy, biliary sludge, 309
 Cholecystitis, 13, 174, 280
 Choledochal cyst, 157
 Cholelithiasis, 13
 Cholera, reportable, 336
 Chorea, Sydenham's, 95
 Chorioretinitis, 191
 congenital infection, 269
 syphilis, 133
Chromobacterium violaceum, taxonomy, 343
 Chronic granulomatous disease, 5
 Cidofovir, adenovirus, 311
 Ciprofloxacin, 131
 Neisseria meningitidis, 88
 Cirrhosis, 14
Citrobacter
 meningitis, 108
 taxonomy, 342
 Clarithromycin, *Mycobacterium avium*
 complex, 240
 Clindamycin, 50
 clostridial myonecrosis, 132
 dental infection, 272
 Lemierre syndrome, 333
 malaria, 188
 protein synthesis inhibition, 104
 retropharyngeal abscess, 264
 skeletal infection, 267
Clonorchis sinensis, taxonomy, 349
 Clostridia, histotoxic, 132
Clostridium, 314
Clostridium botulinum, 129
 taxonomy, 345
Clostridium difficile, 79
 taxonomy, 345
Clostridium perfringens, 79, 132
 taxonomy, 345
Clostridium septicum, 132
Clostridium tetani, 119
 taxonomy, 345
 Cloxacillin, *Staphylococcus aureus*, 118
 Clubbing, infective endocarditis, 297
 Clutton's joint, 152
 CMV *See* Cytomegalovirus
 Coagulopathy, 28, 40, 102
Coccidioides immitis
 pneumonia, 251
 taxonomy, 347
 Coccidioidomycosis, reportable, 336
 Coinfection, 14
 Colchicine, 305
 Colitis
 amebic, 175
 cytomegalovirus, 56
 Colorado tick fever virus, 76
 taxonomy, 339
 Coma, tuberculous meningitis, 215
 Complement, 117
 Complement deficiency, *Neisseria meningitidis*,
 89
 Computer axial tomography, 195, 210, 218,
 254, 261
 cat-scratch disease, 144
 retropharyngeal abscess, 264
 Condyloma, 133
 Confusion *See* Altered mental status
 Congenital infection, 72, 268
 Congo Crimean hemorrhagic fever, 41, 76
 virus
 taxonomy, 340
 Conjunctiva, 26
 Conjunctivitis, 27, 151
 African tick bite fever, 300
 Kawasaki disease, 230
 measles, 17
 neonatal, 92
 typhus, 151
 Constipation, 127
 Contraception, 279
 Coomb's test, 259, 325
 Cor pulmonale, cystic fibrosis, 168
 Corneal disease, 151
 Neisseria gonorrhoeae, 94
 zoster, 53
 Corneal opacity, 71
 Corneal perforation, 22
 Corneal ulcer *See* Ulcer: corneal
 Coronary artery aneurysm, 231
 Coronaviridae, taxonomy, 339
 Corticosteroid, 123, 310, 315
 autoimmune hemolytic anemia, 324
 bronchiolitis, 26
 compressive myelopathy, 263
 infectious mononucleosis, 39
 interstitial keratitis, 152
 meningitis, 106, 164
 neurocysticercosis, 197
 pneumonia, 330
 systemic lupus erythematosus, 42
 tuberculosis, 220
 tuberculous meningitis, 213
Corynebacterium diphtheriae, 34, 120, 310
 taxonomy, 344
Corynebacterium jeikeium, taxonomy, 344

- Corynebacterium urealyticum*, taxonomy, 344
Corynebacterium, infective endocarditis, 297
Coryza *See* Rhinorrhea
Cough, 117, 124, 125, 279
 adenovirus, 310
 bronchiolitis, 24
 cystic fibrosis, 167
 laryngotracheobronchitis, 316
 measles, 17
 pertussis, 159
 pneumonia, 67, 246, 323, 329
 tuberculosis, 220
Cowpox virus, taxonomy, 338
Coxiella burnetii
 infective endocarditis, 297
 pneumonia, 250, 329
 taxonomy, 341
Coxsackie virus, 29
 taxonomy, 339
Crackles, 67
 bronchiolitis, 24
Cranial nerve palsy
 Lyme disease, 142
 diphtheria, 121
 meningitis, 107
 tuberculous meningitis, 209
Creatine phosphokinase, myositis, 75
Cribriform plate, 326
Croup *See* Laryngotracheobronchitis
Cryptococcosis *See* *Cryptococcus neoformans*
Cryptococcus neoformans, 55
 chronic meningitis, 212
 taxonomy, 346
Cryptosporidiosis, reportable, 336
Cryptosporidium parvum, 66, 79, 178, 193
 taxonomy, 347
CSF *See* Cerebrospinal fluid
Culture
 bacterial, 121
 aspirate, 166, 172
 stool, 170
 blood, 40, 102, 111, 112, 140, 155, 158, 173, 181, 259, 333
 infective endocarditis, 294, 298
 intracranial suppuration, 327
 peritonitis, 316
 skeletal infection, 267
 tuberculosis, 235
 typhoid fever, 155
 cervix, 321
 ear discharge, 327
 nasopharyngeal
 pertussis, 161
 peritoneal fluid, 316
 sputum, 235, 247
 stool
 gastroenteritis, 183
 typhoid fever, 155
 throat, 96, 101
 urine, 157, 244
 viral, 59, 72
 cytomeglaovirus, 57
Cunninghamella, taxonomy, 347
Cushing's syndrome, 55
Cutaneous larva migrans, 205
Cuteribridae, taxonomy, 350
Cyclospora cayetanensis, 79, 178
 taxonomy, 347
Cyclosporiasis, reportable, 336
Cyclosporine, 37
Cyst, abdominal, 287
Cystic fibrosis, 68, 168, 250
Cysticercosis, 42
 seizure, 195
Cystitis, 198
Cytochrome oxidase, 10, 280
Cytokine, malaria, 183
Cytomegalovirus, 5, 15, 36, 55, 62, 66, 125, 157, 192, 311
 blood transfusion, 173
 congenital infection, 70, 268
 pneumonia, 329
 taxonomy, 338
 transverse myelitis, 263
Cytopathology, measles, 22

Darwin, Charles, 276
Deafness
 congenital rubella, 71
 congenital syphilis, 136
 meningitis, 109
 syphilis, 153
Dehydration
 gastroenteritis, 79
 physical signs, 77
Democratic Republic of the Congo, 181
Dengue virus, 41, 76, 139, 150, 181
 taxonomy, 340
Dental infection, 271
Dermatitis, atopic, 5, 65
Dermatome, zoster, 52
Dermatomyositis, 74
Dermatophytes, taxonomy, 347
Desquamation, congenital syphilis, 133
Developmental delay, 191
Dexamethasone, laryngotracheobronchitis, 319

- Diabetic keto-acidosis, mucormycosis, 243
 Diagnosis, 1
 microbiological, 3, 4, 6
 Diarrhea, 21, 77, 169, 172, 177
 acute infectious *See* Gastroenteritis
 amebiasis, 174
 bloody, 80
 chronic, 80
 malaria, 184
 osmotic
 lactose intolerance, 80
 typhoid fever, 155
 Dicloxacillin, neck abscess, 261
 Diphtheria, 74, 100, 120, 318, *See also*
 Corynebacterium diphtheriae
 reportable, 336
Diphyllobothrium latum, taxonomy, 349
Dipylidium caninum, taxonomy, 349
 Discharge, conjunctival, 17
 Discitis, 273
 DNA viruses, taxonomy, 337
 DNase, cystic fibrosis, 168
 Downy cells, 35
 Doxycycline, 131
 leptospirosis, 140
 Lyme disease, 143
 malaria, 188
 onchocerciasis, 207
 pelvic inflammatory disease, 322
 pneumonia, 330
 Rickettsia rickettsii, 148
 ricketsial infection, 301
 typhus, 151
Dracunculus medinensis, taxonomy, 350
 Drug eruption, 17
 Drug fever, 109, 334
 Drug interaction, 10, 88, 223, 278, 279
 rifampin, 280
 Drug metabolism, 9
 Drug toxicity
 antibiotic, 278
 hepatotoxicity, 279
 Drugs, 15
 Duke criteria, infective endocarditis,
 298
 Dysarthria, 61
 Dysentery *See* Diarrhea: bloody
 Dysrhythmia, 188

 Early antigen, 39
 Ebola virus, 40, 41, 181
 taxonomy, 340
 EBV *See* Epstein-Barr virus

Echinococcus granulosus, 288
 taxonomy, 349
Echinococcus multilocularis, taxonomy, 349
 Echocardiography, 102
 infective endocarditis, 298
 Kawasaki disease, 231
 Echovirus, 29
 taxonomy, 339
 Ecthyma gangrenosum, 172
 Edema, 98
 cerebral
 meningitis, 106
 neurocysticercosis, 197
 congenital syphilis, 135
 periorbital, 313
 pulmonary, 249
 malaria, 184
Edwardsiella, taxonomy, 342
 Efflux, 277
 Effusion, pericardial, 249
 Egypt, 198
Ehrlichia, 6, 76, 311
Ehrlichia chaffeensis, 147
 taxonomy, 341
Ehrlichia ewingii, 147
Eikenella corrodens, 154
 infective endocarditis, 297
 retropharyngeal abscess, 264
 taxonomy, 343
 Electroencephalogram, 21
Elizabethkingae meningosepticum, taxonomy, 342
 Embolus, infective endocarditis, 294
 Empyema
 epidural, 256
 sinusitis, 257
 pleural, 117, 232, 249
 amebiasis, 175
 subdural, 43, 255
 meningitis, 108
 Enanthem, varicella, 47
 Encephalitis
 African trypanosomiasis, 191
 causes, 42
 cytomegalovirus, 56
 herpes simplex virus, 28
 infectious mononucleosis, 36
 Lyme disease, 142
 measles, 21
 Mycoplasma, 252
 Neisseria meningitidis, 92
 seizure, 195
 usage of term, 43
 varicella, 51

- Encephalitozoon intestinalis*, 180
 taxonomy, 348
- Encephalomyelitis, acute demyelinating, 21
- Encephalomyelopathy, acute disseminated, 263
- Encephalopathy, 142
 acute HIV infection, 312
 cat-scratch disease, 144
 human immunodeficiency virus infection, 62, 65
 malaria, 184
 shigellosis, 80
 toxic, 80
 typhoid fever, 155
 varicella *See* Altered mental status
- Endocarditis
 infective, 58, 87, 95, 102, 154, 172
 differential diagnosis, 298
Neisseria meningitidis, 92
 rat bite fever, 292
 renal dysfunction, 278
 pacemaker, 294
- Endometriosis, 320
- Endophthalmitis, 313
- Endoscopy, tracheitis, 319
- Endothelium, malaria, 183
- Endotoxin, *Neisseria meningitidis*, 92
- Endotracheal intubation, epiglottitis, 318
- Entamoeba histolytica*, 6, 79, 174, 174, 178
 taxonomy, 347
- Enterobacteriaceae *See* Bacilli, enteric
- Enterobacter*
 meningitis, 108
 taxonomy, 342
- Enterobius vermicularis*, taxonomy, 349
- Enterococci, 102, 159, 240 *See also*
Enterococcus faecalis
- Enterococci, nosocomial infection, 166
- Enterococcus faecalis*, 5
 taxonomy, 344
 urinary tract infection, 245
- Enterococcus faecium*, taxonomy, 344
- Enterocytozoon bieneusi*, 180
 taxonomy, 348
- Enterovirus, 29, 31, 42, 74, 74, 139
 meningitis, 27
- Environmental factors, 2
- Enzyme induction, drug-interaction, 278
- Eosinophilia
 hookworm, 203
 pneumonia, 251
 schistosomiasis, 155
 visceral larva migrans, 202
- Eosinophilic granuloma, 284
- Epidemic typhus *See* *Rickettsia prowazekii*
- Epidermolysis bullosa, 27, 70
- Epidermophyton*, taxonomy, 347
- Epiglottitis, 164, 264, 318
- Epstein-Barr nuclear antigen, 39
- Epstein-Barr virus, 15, 34, 58, 100, 258, 311, 331
 blood transfusion, 173
 interstitial lymphocytic pneumonitis, 65
 taxonomy, 338
 tonsillitis, 120
 transverse myelitis, 263
- Erysipelothrix rhusiopathiae*, taxonomy, 345
- Erythema, 70
- Erythema infectiosum, 60
- Erythema marginatum, 95
- Erythema multiforme, *Mycoplasma*, 252
- Erythema, chronicum migrans, 141
- Erythroderma, 100, 104
- Erythromycin
 diphtheria, 121
 pertussis, 161
 pneumonia, 279
- Erythropoiesis, extramedullary, 70
- Eschar, 242
 African tick bite fever, 300
 anthrax, 131
- Escherichia coli*, 79, 159
 enterohemorrhagic, 80
 neonatal pneumonia, 250
 taxonomy, 342
 urinary tract infection, 157, 245
- Esophagitis, 66
- Ethambutol, 213, 221
Mycobacterium avium complex, 240
 tuberculosis, 240
- Eubacterium*, taxonomy, 346
- Eutrombicula alfreddugesi*, taxonomy, 350
- Ewing sarcoma, 284
- Exotoxin
Bacillus cereus, 314
 anthrax, 131
 scarlet fever, 100
- Exposure, 2, 3
- Eye disease, 71
 onchocerciasis, 206
 toxoplasmosis, 193
- Failure to thrive, 63, 65
- Famcyclovir, zoster, 53

- Familial Mediterranean fever, 305
Fasciola hepatica, taxonomy, 349
Fasciolopsis buski, taxonomy, 349
 Fatigue, 203
 Fecal smear, 83
 Fecal–oral *See* Route of spread: fecal–oral
 Fetus, infection, 54
 Fever, 39, 58, 63, 75, 77, 87, 94, 104, 114, 123, 124, 125, 172, 181, 189, 237, 275, 315, 331
 abdominal abscess, 158
 acute HIV infection, 311
 adenovirus, 310
 African tick bite fever, 300
 appendix abscess, 253
 bacteremia, 172
 brain abscess, 326
 cat-scratch disease, 143
 cervical lymphadenitis, 260
 hepatitis, 14
 Kawasaki disease, 230
 laryngotracheobronchitis, 316
 leptospirosis, 139
 lymphadenitis, 162
 measles, 17
 meningitis, 105, 163, 274
 neonate, 26
 nosocomial infection, 165
 of unknown origin, 174
 cat-scratch disease, 144
 osteomyelitis, 281
 pelvic abscess, 305
 PFAPA syndrome, 304
 pharyngitis, 34
 pneumonia, 55, 67, 117, 246, 279, 323, 329
 cystic fibrosis, 167
 poliomyelitis, 31
 pyelonephritis, 244
 rat bite fever, 291
 relapsing fever, 137
 retropharyngeal abscess, 263
 Rocky Mountain spotted fever, 146
 septic arthritis, 153
 sinusitis, 255
 skeletal infection, 266
 suppurative pericarditis, 269
 tuberculosis, 220, 233
 typhoid fever, 154
 typhus, 149
 varicella, 44
 ventriculitis, 112, 278
 Fifth disease *See* Erythema infectiosum
 Filoviridae, 41
 taxonomy, 340
 Fistula, tracheoesophageal, 68
 Fitz-Hugh-Curtis syndrome *See* Perihepatitis
 Flaviviridae, 41, 76
 taxonomy, 340
 Flea, 148
 Fluconazole, 42, 103, 240
 nosocomial infection, 166
 Flukes *See* Trematodes
 Fluorescent antibody test, pertussis, 161
 Fluoroquinolone, 173
 cat-scratch disease, 144
 cystic fibrosis, 168
 gastroenteritis, 86
 Nocardia, 127
 salmonella, 171
 typhoid fever, 156
 Fly, 42, 76
 ectoparasite
 taxonomy, 350
 Onchocerca volvulus, 206
 Fontanelle, meningitis, 163
 Foreign body inhalation, 160, 317
 Foreign body, intravascular, 171
 Foscarnet, 57
Francisella tularensis, 76, 163
 taxonomy, 342
 Fumagillin, microsporidiosis, 180
 Fungal infection, 220
 chronic meningitis, 210
 Fungi, 5
 opportunistic, 55
 taxonomy, 346
 Furazolidone, giardiasis, 180
 Furosemide, glomerulonephritis, 98
Fusobacterium necrophorum
 Lemierre syndrome, 332
 pneumonia, 252
Fusobacterium, 346
 brain abscess, 327
 Gait disturbance, 61
 Galactosemia, 157
 Gallbladder, 13
 hydrops, 231
 Gametocyte, plasmodium, 183
 Gammaherpesvirinae, taxonomy, 338
 Gancyclovir, 56, 269
 Ganglion, sensory, 52
 Gangrene
 Kawasaki disease, 231
 Neisseria meningitidis, 89

- Gardnerella vaginalis*, taxonomy, 345
 Gas gangrene *See* Myonecrosis
 Gastric washings, 211, 220
 tuberculosis, 235
 Gastritis, 13, 83, 280
 Gastroenteritis, 169
 adenovirus, 310
Gemella, taxonomy, 344
 Genetic factors, 2
 Genital ulcer *See* Ulcer: genital
 Genome, viral, 32
 Gentamicin, 86, 173
 cat-scratch disease, 144
 endocarditis, 102
 infective endocarditis, 295, 299
 Listeria monocytogenes, 123
 tularemia, 163
 Ghon complex, 216
 Giant cells
 multinucleate
 measles, 22
Giardia lamblia, 66, 79, 178
 taxonomy, 347
 Gibbs, 226
 Glandular fever *See* Infectious
 mononucleosis
 Glaucoma, zoster, 53
 Glomerulonephritis
 acute poststreptococcal, 98
 congenital syphilis, 133
 human immunodeficiency virus infection, 65
 infectious mononucleosis, 36
 infective endocarditis, 278, 297
Glossina, 190
 Glucose-6-phosphate dehydrogenase deficiency,
 258, 324
 malaria, 188
Gnathostoma spinigerum, taxonomy, 350
 Goodpasture's disease
 Goodpasture's syndrome, 251, 324
 Gram stain, 6, 105, 112, 114, 124, 126, 164,
 166, 172, 240, 261, 278, 313
 cerebrospinal fluid, 112
 Clostridium, 131
 Francisella tularensis, 163
 sputum, 247
 urine, 157
 Granuloma, 220
 mycobacterial infection, 240
 schistosomiasis, 198
 tuberculosis, 237
 Granulomatosis infantiseptica, 123
 Growth plate, 110
 Guatemala, 195
 Guillain-Barre syndrome, 31, 74, 121, 128
 Campylobacter jejuni, 82
 infectious mononucleosis, 36
 Guinea worm *See* *Dracunculus medinensis*

 HACEK bacteria, 153
Haemophilus aphrophilus, 153
 infective endocarditis, 297
 taxonomy, 343
Haemophilus ducreyi, taxonomy, 343
Haemophilus influenzae, 2, 20, 66, 125
 cystic fibrosis, 168
 epiglottitis, 318
 intracranial suppuration, 256
 lobar pneumonia, 247
 meningitis, 164
 pneumonia, 250
 retropharyngeal abscess, 264
 septic arthritis, 153, 266
 suppurative pericarditis, 270, 271
 taxonomy, 343
Hafnia, taxonomy, 342
 Hantaviruses, taxonomy, 340
 Headache, 39, 203, 304
 acute HIV infection, 311
 brain abscess, 326
 leptospirosis, 139
 relapsing fever, 137
 Rocky Mountain spotted fever, 147
 sinusitis, 255
 typhoid fever, 154
 typhus, 149
 ventriculitis,, 278
 VP shunt malfunction, 112
 Health department, 7, 11, 23, 32, 41, 44, 88,
 131, 165
 diphtheria, 122
 Neisseria gonorrhoeae, 94
 pertussis, 162
 syphilis, 152
 tuberculosis, 213, 225
 typhoid fever, 156
 viral hemorrhagic fever, 182
 Heart disease, 68
 congenital, 26, 58, 71, 102, 114, 293
 infective endocarditis, 297
 iron overload, 173
 Lyme disease, 142
 Heart failure, 98
 infective endocarditis, 294, 297
 rheumatic heart disease, 95
 Heart murmur, 95, 102

- Heart valve, 102
 - rheumatic fever, 95
- Helicobacter pylori*, taxonomy, 342
- Hemarthrosis, 141
- Hematemesis, 40
- Hematochezia, 83, 284
- Hematogenous spread, 110, 125, 127, 174, 213, 247, 315
 - Mycobacterium tuberculosis*, 226
 - spinal abscess, 262
- Hematuria, 13, 75, 258
 - glomerulonephritis, 98
 - infective endocarditis, 297
 - schistosomiasis, 198
- Hemiplegia
 - Rocky Mountain spotted fever, 148
 - intracranial suppuration, 255
 - zoster, 53
- Hemoconcentration, dehydration, 79
- Hemoglobinuria, 258
- Hemolytic uremic syndrome, 258
 - gastroenteritis, 80
 - pneumococcal infection, 252
- Hemorrhage
 - neonate, 268
 - splinter, 294
- Hepadnaviridae, taxonomy, 338
- Heparin, Lemierre syndrome, 333
- Hepatic failure *See* Liver failure
- Hepatic sequestration, 13
- Hepatic sludging, 13
- Hepatitis, 13, 157
- Hepatitis A virus, 181
 - taxonomy, 339
- Hepatitis B immune globulin *See* Immune globulin
- Hepatitis B virus, 321
 - blood transfusion, 173
 - taxonomy, 338
- Hepatitis C virus, 340
 - blood transfusion, 173
- Hepatitis viruses, 311
- Hepatitis
 - A, 14
 - adenovirus, 310
 - B, 14, 59
 - C, 14, 59
 - congenital rubella, 71
 - congenital syphilis, 133
 - cytomegalovirus, 56
 - D, 14
 - differential diagnosis, 15
 - E, 14
 - enterovirus, 29
 - infectious mononucleosis, 36
 - Lemierre syndrome, 331
 - neonatal, 157
 - varicella, 52
- Hepatomegaly, 157
 - acute HIV infection, 311
 - amebic abscess, 174
 - cat-scratch disease, 143
 - congenital infection, 69
 - congenital syphilis, 133
 - hepatitis, 14
 - human immunodeficiency virus infection, 65
 - neonate, 268
 - relapsing fever, 137
 - thalassemia, 173
 - toxoplasmosis, 193
 - tuberculosis, 233
 - typhoid fever, 154
 - visceral larva migrans, 202
 - yellow fever, 75
- Hereditary spherocytosis, 258
- Herpes simplex virus, 20, 26, 42, 46, 66, 70, 152, 192
 - genital, 27
 - neonatal pneumonia, 250
 - neonate, 27
 - taxonomy, 337
- Herpes viruses, 5
- Herpesviridae, taxonomy, 337
- Heterophile antibody test, 35, 259
- Heterophyes heterophyes*, taxonomy, 349
- Hibernian fever, 305
- Histoplasma capsulatum*, 55, 160
 - pneumonia, 251, 329
 - taxonomy, 346
- Histoplasma capsulatum* var. *duboisii*, taxonomy, 346
- Histoplasmosis, 233 *See also* Histoplasma capsulatum
- HIV infection *See* Human immunodeficiency virus infection
- Hodgkin's disease, 37
- Honduras, 281
- Hookworm, 203
 - taxonomy, 349
- Horner's syndrome, 118
- Host defense, 3, 5
- Howell-Jolly body, 114
- Human T-cell lymphotropic virus
 - taxonomy, 339
- Human herpes virus 6, taxonomy, 338
- Human herpes virus 7, taxonomy, 338

- Human herpes virus 8, taxonomy, 338
Human immunodeficiency virus, 36, 42, 59, 311
 blood transfusion, 173
 infection, 5, 34, 150, 220, 233, 269, 321
 acute, 312
 pneumonia, 250
 transverse myelitis, 263
 taxonomy, 339
Human metapneumovirus, taxonomy, 339
Hutchinson's teeth, 152
 congenital syphilis, 136
Hutchinson's triad, 153
Hydatid cyst *See also Echinococcus granulosus*
 seizure, 195
Hydration, 26
Hydrocephalus, 112, 215
 meningitis, 108
 tuberculous meningitis, 210
Hydrops fetalis, 61
Hygiene, 78
 cysticercosis, 197
 dental, 273
Hymenolepis nana, taxonomy, 349
Hyper IgD syndrome, 305
Hyperbaric oxygen, 132
Hyperbilirubinemia, 13
 neonatal, 284
Hyperglycemia, dehydration, 80
Hyperinflation, bronchiolitis, 24
Hyperkalemia, rhabdomyolysis, 75
Hypernatremia, dehydration, 80
Hypersensitivity pneumonia, 251
Hypertension
 glomerulonephritis, 98
 hemolytic uremic syndrome, 80
Hyperthermia, malignant, 335
Hypertonia, 31, 80, 191
Hyphae, 243
 pityriasis versicolor, 303
Hypnozoite, 183
Hypoalbuminemia, 98
Hypocalcemia, 74
 neonate, 119
 stridor, 318
Hypoglycemia, 21, 54, 104, 192
 diarrhea, 80
 malaria, 184
 seizure, 195
Hypokalemia, 74
 diarrhea, 80
Hyponatremia, 21, 104
 diarrhea, 80
 seizure, 195
Hypoproteinemia, pleural effusion, 232
Hypothyroidism, 157
Hypotonia, 31, 80
 botulism, 127
Hypoxia, 21, 54
 pertussis, 161
 pneumonia, 167
 seizure, 195
 visceral larva migrans, 202
Hypoxic-ischemic encephalopathy, 192
 seizure, 195
Ibuprofen, rheumatic fever, 95
IgM
 hepatitis A, 15
 measles, 22
Imipenem, Nocardia, 127
Immotile cilia syndrome, 68
Immune globulin, 15, 104
 botulinum, 129
 cytomegalovirus, 57
 hepatitis B, 16
 intravenous, 231
 measles, 24
 palivizumab, 26
 tetanus, 120
 varicella zoster, 45
Immune reconstitution syndrome, 223
Immunization, 120, 122 *See also Vaccination*
Immunocompromised host, 18, 24, 35, 37, 47, 55,
 132, 146, 171, 193, 220, 242, 247, 310
 measles encephalitis, 21
 parvovirus, 61
 pneumonia, 250, 329
Immunodeficiency, 68, 168
 congenital, 5
Immunofluorescence, varicella, 46
Immunoglobulin deficiency, 5
Immunologic reaction, 109
Immunomodulation, 39
Immunostaining, 6
Immunosuppression, 20, 123, 125
Immunosuppression *See also*
 Immunocompromised host
Impetigo, 98
 bullous, 27, 46
Incontinence, 262
Incubation period
 Epstein-Barr virus, 35
 hepatitis, 14
 measles, 18
 rat bite fever, 292
 varicella, 45

- Index case, 10
- Infarction, ecthyma gangrenosum, 172
- Infectious mononucleosis, 34, 100, 137, 154, 298, 310, 311
 - hemolytic anemia, 258
 - postperfusion syndrome, 58
- Inflammatory bowel disease, 83
- Influenza virus, 55, 63, 74
 - pneumonia, 250, 329
 - taxonomy, 339
- Injury, traumatic, 74
- Interferon, 15
 - rabies, 42
- Intestinal obstruction, 253
 - Ascaris lumbricoides*, 208
 - pelvic inflammatory disease, 322
- Intestinal perforation, 81, 175, 284
 - cytomegalovirus, 56
 - typhoid fever, 155
- Intestinal protozoa, taxonomy, 347
- Intoxication, 54, 74
 - botulism, 129
 - food, 77
 - pneumonia, 251
 - seizure, 195
- Intracranial hypertension, 83
- Intracranial pressure elevation, 255
- Intracranial pressure, meningitis and, 106
- Intracranial suppuration, sinusitis, 255
- Intrauterine infection *See* Congenital infection
- Intravenous drug abuse, 14
- Intussusception, 82
- Iodoquinol, amebiasis, 177
- Iron deficiency, hookworm, 203
- Iron overload, 80, 173
- Isolation, 8, 11, 181
 - airborne, 11, 23, 225
 - contact, 11, 25, 44, 156
 - droplet, 11, 25, 162
 - diphtheria, 122
 - universal, 11
- Isoniazid, 15, 213, 221, 223, 233, 240, 279
- Isospora belli*, 66, 79, 180
 - taxonomy, 347
- Ivermectin
 - ascariasis, 208
 - cutaneous larva migrans, 205
 - onchocerciasis, 206
- Janeway lesions, 297
- Japanese B encephalitis virus, taxonomy, 340
- Jarisch-Herxheimer reaction, 139
 - drug fever, 335
- Jaundice, 12, 157, 257
 - Lemierre syndrome, 331
 - pneumonia, 323
 - thalassemia, 173
 - yellow fever, 75
- Jones criteria, 95
- Kaposi sarcoma, 66
- Kawasaki disease, 17, 230
- Keratitis, 27, 152
 - interstitial, 133, 152
 - measles, 22
- Keratomalacia, 22
- Kerosene, 247
- Kidney disease, cytomegalovirus, 56
- Kingella kingae*
 - infective endocarditis, 297
 - septic arthritis, 267
 - skeletal infection, 153
 - spinal infection, 274
 - taxonomy, 343
- Klebsiella pneumoniae*, 20
 - taxonomy, 342
- Klebsiella*, urinary tract infection, 245
- Koplik spots, 18
- Kwashiorkor, 21, 240
- Labor, abdominal pain, 320
- Lactobacillus*, taxonomy, 345
- Lamivudine, 15
- Langerhans histiocytosis, 233
- Larva, *Taenia solium*, 195
- Laryngeal disorders, 318
- Laryngotracheobronchitis, 18, 318
- Lassa fever virus, 40, 76
 - taxonomy, 340
- Lateral pharyngeal space, 237, 261
 - dental abscess, 272
- Latin America, 195
- Lead poisoning, 203
- Lecithinase, 132, 314
- Legionella, 125
 - intramacrophage survival, 123
- Legionella pneumophila*, 55, 63, 329
 - taxonomy, 343
- Leiomyosarcoma, 37
 - human immunodeficiency virus infection, 66
- Leishmania braziliensis*, taxonomy, 348
- Leishmania donovani*, taxonomy, 348
- Leishmania mexicana*, taxonomy, 348
- Leishmaniasis, 131
- Lemierre syndrome, 332
 - pneumonia, 252

- Lentivirus, taxonomy, 339
- Leptospira*, 181
- Leptospira interrogans*, 140
 - taxonomy, 341
- Leptospirosis, 15, 40, 76, 138, 139, 150, 231, 293, 311
- Leucopenia
 - cytomegalovirus, 56
 - human immunodeficiency virus infection, 65
 - infectious mononucleosis, 36
 - yellow fever, 75
- Leukemia, 155, 233, 312
 - congenital, 269
- Leukocyturia, 13
- Leukonostoc*, taxonomy, 344
- Liberia, 205
- Libya, 174
- Life cycle
 - Ascaris, 207
 - Echinococcus granulosus, 290
 - hookworm, 203
 - Onchocerca volvulus, 206
 - Schistosoma, 198
 - Taenia solium, 196
 - visceral larva migrans, 202
- Limp, 30, 153, 266, 273
- Listeria monocytogenes*, 5, 59, 63, 66, 123, 268
 - blood transfusion, 173
 - neonatal pneumonia, 250
 - taxonomy, 345
- Listeriosis *See* *Listeria monocytogenes*
- Liver, 13
- Liver failure, 14, 28, 54
 - malaria, 184
 - yellow fever, 76
- Loa loa*, taxonomy, 350
- Lobar consolidation, 246
- Loperamide, 85
- Louse, 76, 148, 150
 - relapsing fever, 138
 - taxonomy, 350
- Lower motor neuron, 31, 68, 73
 - botulism, 127
- Lung biopsy *See* Biopsy: lung
- Lung cavity, 210, 225
- Lung disease, 234, 329
 - physical findings, 216
- Lung examination, 249
- Lung puncture, 247
- Lyme disease, 141
- Lymphadenitis, 100, 162
 - cat-scratch disease, 143
 - cervical, 237, 261
 - Kawasaki disease, 230
- Lymphadenopathy, 58, 189
 - acute HIV infection, 311
 - African tick bite fever, 300
 - autoimmune lymphoproliferative syndrome, 310
 - cat-scratch disease, 144
 - cervical, 34, 36, 99, 264
 - PFAPA syndrome, 304
 - hilar, 68, 210, 216
 - human immunodeficiency virus infection, 65
 - stridor, 318
 - submandibular, 271
 - syphilis, 133
- Lymphocyte, atypical, 35
- Lymphocytic choriomeningitis virus, 72, 192, 269
 - taxonomy, 340
- Lymphocytic interstitial pneumonitis, 233
- Lymphocytosis
 - infectious mononucleosis, 35
 - pertussis, 161
- Lymphoma, 37, 68, 155, 220, 233
 - B-cell, 37
 - human immunodeficiency virus infection, 66
 - Burkitt's, 37
 - Hodgkin's disease, 50
 - pleural effusion, 232
 - primary brain, 62
- Lymphopenia, human immunodeficiency virus infection, 65
- Macrolide
 - pertussis, 162
 - pneumonia, 325
 - relapsing fever, 139
 - Streptococcus pyogenes*, 101
- Magnetic resonance imaging, 263
 - skeletal infection, 267
 - spine, 273
- Malabsorption, cystic fibrosis, 168
- Malaria, 190, 269
- Malaria *See also* Plasmodium
 - hemolytic anemia, 259
- Malassezia furfur*, 303
 - taxonomy, 346
- Malformation, lung, 68
- Malnutrition, 21
 - cystic fibrosis, 168
- Malrotation, 253
- Marburg virus, 40, 41
 - taxonomy, 340

- Measles, 17, 230
 pneumonia, 250
 Measles virus, taxonomy, 339
 Mebendazole
 ascariasis, 208
 hookworm, 203
 Meckel's diverticulum, 83
 Mediastinitis, anthrax, 131
 Mediterranean spotted fever *See* Rickettsia conori
 Mefloquine, 188
 Meningismus, leptospirosis, 139
 Meningitis, 21, 26, 63, 104, 158, 164, 327
 acute HIV infection, 312
 anthrax, 131
 aseptic, 31, 231
 Lyme disease, 142
 chronic, 210
 congenital syphilis, 133
 Listeria monocytogenes, 123
 Neisseria meningitidis, 92, 147
 pneumococcal, 308
 salmonella, 80, 170
 seizure, 195, 278
 sinusitis, 255
 Streptococcus agalactiae, 113
 Streptococcus pneumoniae, 106, 274, 325
 tuberculous, 210, 223, 237
 usage of term, 42
 Meningococcal infection *See* Neisseria
 meningitidis
 Meningococemia *See* Neisseria meningitidis
 Meningoencephalitis, 26
 usage of term, 43
 Meropenem
 brain abscess, 328
 epidural abscess, 335
 intracranial suppuration, 256
 Merozoite, plasmodium, 182
 Metabolic disease, 15, 21, 28, 28, 104, 192
 diarrhea, 80
 jaundice, 157
 seizure, 195
 varicella, 54
 Metagonimus yokagawai, taxonomy, 349
 Metaphysis, 110
 osteomyelitis, 281
 Metapneumovirus, 63
 pneumonia, 250
 Methicillin resistance *See* Staphylococcus
 aureus: methicillin-resistant
 Metronidazole
 abdominal infection, 254
 amebiasis, 175
 amebic abscess, 174
 brain abscess, 328
 clostridial myonecrosis, 132
 Clostridium difficile, 86
 epidural abscess, 334
 giardiasis, 180
 intracranial suppuration, 256
 Lemierre syndrome, 333
 Mevalonate kinase, 305
 Microcephaly, 71, 191
 Microfilaria, 6, 206
 Microphthalmia, 71
 Microscopy
 dark field, 6
 electron, 6
 Microsporidia (tissue), taxonomy, 348
 Microsporidium, 180
 Microsporium, taxonomy, 347
 Minimal inhibitory concentration, 275
 Minocycline, Nocardia, 127
 Miracidium, 198
 Mites, taxonomy, 350
 Mitral valve insufficiency, 95
 Molluscum contagiosum virus, taxonomy, 338
 Monitor, fetal scalp, 27
 Monkey, 77
 Monkeypox virus, taxonomy, 338
 Moraxella catarrhalis, 2
 taxonomy, 343
 Morganella, taxonomy, 342
 Mosquito, 42, 76, 182
 MRI *See* Magnetic resonance imaging
 Muckle-Wells syndrome, 305
 Mucocutaneous disease, herpes simplex virus,
 27
 Mucormycosis, 243
 Mucor, taxonomy, 347
 Multiceps multiceps, taxonomy, 349
 Multiorgan dysfunction, 28
 viral hemorrhagic fever, 41
 Multiple sclerosis, 263
 Mumps virus, taxonomy, 339
 Muscle atrophy, 32
 Muscle disease, 74, 129
 Muscle, 31
 wasting, 32
 Mushroom, 15
 Mutation, antibiotic resistance, 276
 Myalgia, 75
 acute HIV infection, 311
 leptospirosis, 139
 poliomyelitis, 31
 relapsing, 137

- [Myalgia cont.]
 - typhus, 149
 - viral hemorrhagic fever, 41
- Myasthenia gravis, 128
- Mycobacteria, 5, 6, 220
 - intramacrophage survival, 123
 - nontuberculous, 238, 261
 - pneumonia, 250
 - taxonomy, 345
- Mycobacterium avium* complex, 66, 221
- Mycobacterium tuberculosis*, 55, 63, 125, 159, 211
 - pneumonia, 251
- Mycoplasma genitalium*, taxonomy, 341
- Mycoplasma hominis*, 250
 - taxonomy, 341
- Mycoplasma pneumoniae*, 42, 55, 63, 125, 159
 - lobar pneumonia, 247
 - pneumonia, 250, 324, 329
 - taxonomy, 341
- Myelitis
 - acute HIV infection, 312
 - transverse, 74, 263
 - infectious mononucleosis, 36
- Myelogram, 263
- Myelopathy
 - compressive, 263
 - human immunodeficiency virus infection, 66
 - vacuolar, 62
- Myocardial infarction, 231
- Myocarditis
 - diphtheria, 121
 - enterovirus, 29
 - infectious mononucleosis, 36
 - Kawasaki disease, 231
 - measles, 22
 - rheumatic fever, 95
 - typhoid fever, 155
 - varicella, 52
- Myoglobinuria, 75, 258
- Myonecrosis, 132
- Myositis, 31, 74, 129

- Naegleria fowleri*, 42
 - taxonomy, 348
- Nafcillin, 51, 104
 - cellulitis, 275
 - discitis, 274
 - endocarditis, 102
 - infective endocarditis, 295, 299
 - osteomyelitis, 111
- Nafcillin resistance, *See* *Staphylococcus aureus*:
 - methicillin-resistant
- Nausea, 12
- Necator americanus*, 203
- Neck stiffness
 - brain abscess, 326
 - cervical lymphadenitis, 260
 - lymphadenitis, 237
 - pneumonia, 248
- Necrosis, ischemic, 242
- Necrotizing enterocolitis, 284
- Necrotizing fasciitis, 48
- Neisseria gonorrhoeae*, 34, 94
 - disseminated, 293
 - pelvic inflammatory disease, 321
 - taxonomy, 343
- Neisseria meningitidis*, 2, 150
 - arthritis, 293
 - bacteremia, 147
 - suppurative pericarditis, 271
 - taxonomy, 343
- Nematodes, taxonomy, 349–350
- Neonate
 - sepsis, 113
 - tetanus, 119
- Neoplasm, seizure, 195
- Nephrotic syndrome, 315
 - congenital syphilis, 133
 - malaria, 184
- Nervous system, 31
- Neuralgia, postherpetic, 53
- Neuroblastoma, 273
- Neuromuscular junction, 31, 74, 128
- Neuromyelitis optica, 263
- Neutropenia, 5
 - human immunodeficiency virus infection, 65
 - mucormycosis, 243
 - pneumonia, 250
- Nifedipine, glomerulonephritis, 98
- Nikolsky sign, 70
- Nitazoxanide, *Cryptosporidium*, 86, 180
- Nocardia*, 5, 55, 66, 124
 - pneumonia, 250
 - bone marrow transplantation, 125
 - taxonomy, 344
- Nodule
 - onchocerciasis, 205
 - subcutaneous
 - rheumatic fever, 95
- Norovirus, 79
 - taxonomy, 339
- Nosocomial infection, 102, 109, 165, 181, 334, 353
 - infective endocarditis, 297

- Streptococcus pneumoniae, 275
 - viral hemorrhagic fever, 41
- Nucleic acid, 6
- Nutrition, 14
- Obstruction
 - airway, 36, 265, 316
- Obtunded, *See* Altered mental status
- Occupation, 3
- Odontogenic infection *See* Dental infection
- Odynophagia, acute HIV infection, 311
- Oerskovia*, taxonomy, 345
- Onchocerca volvulus*, 205
 - taxonomy, 350
- Onchocerciasis *See* *Onchocerca volvulus*
- Oncogenic, 35
- Oocyst, plasmodium, 183
- Ophthalmoplegia, 313
- Opisthorcis viverrini*, taxonomy, 349
- Opisthotonus, 214
- Opportunistic infection, 42, 62, 63, 65, 221, 233, 315
- Opsonization, 117
 - Neisseria meningitidis*, 89
- Optic atrophy, 325
- Optic neuritis, 145
 - Lyme disease, 142
- Orf virus, taxonomy, 338
- Orientia tsutsugamushi*, taxonomy, 341
- Oroya fever, 146
- Orthopantomogram, 271
- Orthopoxvirus, taxonomy, 338
- Osler's nodes, 297
- Osteogenic sarcoma, 284
- Osteoid osteoma, 284
- Osteomyelitis, 5, 48, 74, 102, 110, 266, 276
 - cat-scratch disease, 144
 - chronic, 282
 - congenital syphilis, 133
 - Kingella kingae*, 153
 - salmonella, 80, 170
 - skull, 257
 - Streptococcus pyogenes*, 100
 - vertebral, 273
- Otitis externa, 327
- Otitis media, 2, 4, 18, 100, 327
 - human immunodeficiency virus infection, 65
- Otorrhea, 326
- Ova, 6
- Ovarian cyst, rupture, 320
- Ovarian torsion, 320
- Oxacillin, 104
- osteomyelitis, 111
- Oxamniquine, 200
- Oxygenation, 26
- Pacemaker, 293
- Pain
 - back, 262, 273
 - chest, 124, 125, 270
 - pneumonia, 248
 - eye, 151, 313
 - facial, 271
 - hip, 305
 - knee, 281
 - leg, 275
 - limb, 131
 - muscle *See* Myalgia
 - neck, 225
 - Lemierre syndrome, 331
 - retropharyngeal abscess, 263
 - throat, 34, 59
 - Lemierre syndrome, 331
- Pancreatic disease, cystic fibrosis, 168
- Pancreatitis, 13
 - biliary sludge, 309
 - pleural effusion, 232
- Pantoea, taxonomy, 342
- Papilledema, intracranial suppuration, 255
- Papilloma virus, taxonomy, 338
- Papilloma, bladder, 198
- Papovaviridae, taxonomy, 338
- Papule, 45, 75
- Paracoccidioides brasiliensis*, taxonomy, 347
- Paragonimus westermanii*, taxonomy, 349
- Parainfluenza virus, 24, 55, 63, 100, 250
 - laryngotracheobronchitis, 319
 - taxonomy, 339
- Paralysis, 31
 - periodic, 74
 - tick, 74
- Parameningeal infection, 43
- Paramyxoviridae, taxonomy, 339
- Paraplegia, 262
 - spinal tuberculosis, 227
- Parasitemia, 186
- Parasites, taxonomy, 347
- Parechovirus, taxonomy, 339
- Parenteral *See* Route of spread: parenteral
- Paresis, 110
- Parinaud's syndrome, tularemia, 163
- Paromomycin, 180
- Parvovirus B 19, taxonomy, 338
- Parvovirus B19, 59, 66, 293

- Pasteurella multocida*, taxonomy, 343
PCR *See* Polymerase chain reaction
Pediculus humanus capitis, taxonomy, 350
Pediculus humanus corporis *See also* Louse
taxonomy, 350
Pediococcus, taxonomy, 344
Pelvic inflammatory disease, 320
Penicillin, 88, 117
Penicillin resistance
 Neisseria gonorrhoeae, 277
 Streptococcus pneumoniae, 275
Penicillin
 actinomycosis, 125
 clostridial myonecrosis, 132
 congenital syphilis, 137
 dental infection, 272
 diphtheria, 122
 leptospirosis, 140
 meningitis, 274
 pneumonia, 325
 rat bite fever, 292
 relapsing fever, 139
 rheumatic fever, 95
 Streptococcus pyogenes, 99, 101
 syphilis, 42, 152
 tetanus, 120
 tonsillitis, 35
Penicillium marneffei, taxonomy, 347
Peptococcus, taxonomy, 346
Peptostreptococcus, taxonomy, 346
Pericardiocentesis, 270
Pericarditis
 amebiasis, 175
 constrictive, 271
 Kawasaki disease, 231
 Neisseria meningitidis, 92
 postcardiotomy syndrome, 58
 rheumatic fever, 95
 suppurative, 270
 Haemophilus influenzae, 164
Perihepatitis, pelvic inflammatory disease, 322
Periodic fever, 305
Periosteum, 110
Periostitis, syphilis, 153
Peripheral nerve, 31, 74, 128
Peripheral neuropathy, 128
 acute HIV infection, 312
 diphtheria, 121
 human immunodeficiency virus infection, 62
Peritonitis, 112
 necrotizing enterocolitis, 284
 pelvic inflammatory disease, 322
 spontaneous bacterial, 315
Pertussis, 160
PFAPA syndrome, 305
Pharmacodynamics, 280
Pharmacokinetics, 9, 280
Pharyngotonsillitis *See* Tonsillitis
Phenytoin, 278
Phlebitis, 109
Phlegmon, retropharyngeal, 264
Photophobia, 75, 151
 African tick bite fever, 300
 leptospirosis, 139
Phthyrus pubis, taxonomy, 350
Physical therapy, 32
Picornaviridae, taxonomy, 339
Pinworm *See* *Enterobius vermicularis*
Pipestem fibrosis, 199
Pityriasis versicolor, 303
Plague, 144 *See also* *Yersinia pestis*
Plasmodium, 7, 59, 181, 193
 blood transfusion, 173
Plasmodium falciparum, 182
 taxonomy, 348
Plasmodium malariae, 183
 taxonomy, 348
Plasmodium ovale, 183
 taxonomy, 348
Plasmodium vivax, 183
 taxonomy, 348
Pleocytosis, 42
Plesiomonas shigelloides, taxonomy, 343
Pleural effusion, 117, 216, 232, 249
Pleural thickening, 220
Pneumatosis intestinalis, 286
Pneumococcus *See* *Streptococcus pneumoniae*
Pneumocystis jiroveci, 5, 55, 66, 125, 250, 329
Pneumonia, 5, 24, 117, 125, 279, 329
 adenovirus, 310
 alba, 133
 cystic fibrosis, 167
 enterovirus, 29
 giant cell, measles, 20
 human immunodeficiency virus infection,
 66
 hypersensitivity, 329
 infectious mononucleosis, 36
 Lemierre syndrome, 331
 lobar, 67, 246
 measles, 19
 mucormycosis, 243
 Neisseria meningitidis, 92
 pertussis, 161
 posttransplant, 55
 recurrent, 67

- Streptococcus agalactiae, 113
- tuberculous, 216
- typhoid fever, 155
- varicella, 47
- visceral larva migrans, 202, 208
- Pneumonitis, interstitial lymphocytic, 65
- Pneumothorax, 216, 249
- Poison ivy, 52
- Poisoning, food *See* Intoxication: food
- Polio virus, 31
 - taxonomy, 339
- Poliomyelitis, 31, 74, 128
- Polymerase chain reaction, 7, 29
 - cytomegalovirus, 57
 - pertussis, 161
- Polyoma virus, 66
 - taxonomy, 338
- Polyp, intestinal, 83
- Pontine myelinosis, 81
- Porin, 276
- Porphyria, acute intermittent, 74
- Porphyromonas*, taxonomy, 346
- Portal hypertension, schistosomiasis, 201
- Posaconazole, mucormycosis, 244
- Postpericardiotomy syndrome, 58, 298
- Postpolio syndrome, 32
- Posttransplant lymphoproliferative disease, 37
- Pott's disease *See* Tuberculosis: spine
- Poxviridae, taxonomy, 338
- PPD, 261 *See also* Tuberculin skin testing
- Praziquantel
 - neurocysticercosis, 42, 197
 - schistosomiasis, 200
- Precautions, laboratory, 163
- Pregnancy, 279
 - abdominal pain, 321
 - congenital infection, 69
 - ectopic, 320
 - malaria, 188
- Prematurity, 26
 - necrotizing enterocolitis, 285
- Prevention, 7
- Prevotella*, taxonomy, 346
- Primaquine, 188
- Prodrome, 14
 - measles, 18
 - varicella, 45
- Prophylaxis, 1, 11, 24, 88, 189
 - diphtheria, 122
 - hyposplenism, 117
 - infective endocarditis, 96, 299
 - urinary tract infection, 246
- Proptosis, 313
- Protease inhibitor, 223
- Proteus*
 - meningitis, 108
 - taxonomy, 342
- Prothrombin time, 15
- Providencia*, taxonomy, 342
- Pseudoallescheria boydii*, taxonomy, 347
- Pseudohyphae, 241
- Pseudohypparathyroidism, 62
- Pseudomembrane, 120
 - infectious mononucleosis, 36
 - Stevens–Johnson syndrome, 17
- Pseudomonas aeruginosa*, 5, 66, 295
 - cystic fibrosis, 168
 - ecthyma gangrenosum, 172
 - endocarditis, 103
 - ischemic necrosis, 242
 - pneumonia, 250
 - taxonomy, 342
- Pseudoparesis, 74
 - congenital syphilis, 133
- Pterigomandibular space, 272
- Ptosis
 - botulism, 31, 127
 - Horner's syndrome, 118
- Public health, 14, 22, 181, 235
 - tuberculosis, 223
 - typhoid fever, 156
- Pulmonary hemorrhage, 324
- Pulmonary hypertension, schistosomiasis, 200
- Pulmonary infarction, 249, 251
- Pulsus paradoxus, airway obstruction, 316, 318
- Purpura fulminans, 89
- Pus, 240
- Pustule, 27, 45, 70
- Pyelonephritis, 13, 244
- Pyrantel pamoate, hookworm, 203
- Pyrazinamide, 213, 221, 233, 240, 279
- Pyrimethamine, 188
 - malaria, 182
 - toxoplasmosis, 42, 194
- Pyrin, 305
- Quinacrine, 180
- Quinidine, 188
- Quinine, 182
- Rabies, 42
- Rabies virus, 340
- Radiography, chest, 25

- Rash, 14
 blueberry muffin, 70
 hemorrhagic, 87, 101, 172
 infective endocarditis, 294
 Neisseria meningitidis, 89
 hypopigmented, 303
 Kawasaki disease, 230
 lacy, 60
 macular, 36
 African tick bite fever, 300
 leptospirosis, 139
 rat bite fever, 291
 syphilis, 133
 viral hemorrhagic fever, 41
 maculopapular
 Neisseria meningitidis, 92
 measles, 18
 typhus, 149
 papular, 18
 petechial, 268
 infective endocarditis, 297
 Neisseria meningitidis, 89
 Rocky Mountain spotted fever, 146
 typhus, 150
 typhoid fever, 154
Rat bite fever, 293
Rectal examination, 306
Reduviid bug, 76
Reflex, deep tendon, 31
Reflux, gastroesophageal, 68
Relapsing fever, 6, 138, 269
Renal failure, 278
 diphtheria, 121
 hemolytic uremic syndrome, 80
 infective endocarditis, 293
 malaria, 184
 rhabdomyolysis, 75
 yellow fever, 76
Reoviridae, 76
 taxonomy, 339
Reptile, salmonellosis, 169
Respiratory distress, 112, 119, 120, 215, 231, 329
 laryngotracheobronchitis, 316
 pneumonia, 248, 323
 suppurative pericarditis, 269
 visceral larva migrans, 202
Respiratory failure, 73
 myositis, 75
 varicella pneumonia, 47
Respiratory syncytial virus, 24, 55, 63, 100
 pneumonia, 250
 taxonomy, 339
Respiratory tract infection, 17
 Respiratory tract, measles, 18
 Respiratory virus infection, 34, 55, 63, 66, 100, 125
 lobar pneumonia, 247
 Reticuloendothelial system, 65
 malaria, 183
 toxoplasmosis, 193
 typhoid fever, 155
 Reticulonodular pattern, miliary tuberculosis, 234
 Retinitis
 cat-scratch disease, 144
 cytomegalovirus, 56
 Retinoblastoma, 202
 Retinopathy, acute HIV infection, 312
 Retractions, bronchiolitis, 24
 Retroviridae, taxonomy, 339
 Reye's syndrome, 21, 54
 varicella, 52
 Rhabdomyolysis, 75
 Rhabdoviridae, 340
 Rhagades, 153
 Rheumatic fever, 95, 100, 141, 293, 298
 Rheumatic heart disease, 95, 297
 Rheumatoid disease, 155, 233, 266, 293
 Rhinorrhea, 24
 cerebrospinal fluid, 325
 congenital syphilis, 134
 laryngotracheobronchitis, 319
 measles, 18
 pertussis, 161
 Rhinovirus, taxonomy, 339
 Rhizopus, taxonomy, 347
 Rhodococcus equi, 125
 pneumonia, 250
 taxonomy, 345
 Rhodotorula rubra, taxonomy, 346
 Ribavirin, 15
 bronchiolitis, 26
 rabies, 42
 viral hemorrhagic fever, 40
 Rickettsia, 6, 40, 76, 139, 150, 293, 311
 eschar, 130, 190
 Rickettsia africae, 148, 301
 taxonomy, 341
 Rickettsia akari, 148
 taxonomy, 341
 Rickettsia australis, 148
 taxonomy, 341
 Rickettsia conorii, 301
 Rickettsia conori, 148
 taxonomy, 341

- Rickettsia mooseri*, 148
 taxonomy, 341
- Rickettsia prowazekii*, 76, 148, 150
 taxonomy, 341
- Rickettsia rickettsii*, 147
 taxonomy, 341
- Rickettsia sibirica*, 148
 taxonomy, 341
- Rickettsiae, taxonomy, 341
- Rickettsialpox *See* *Rickettsia akari*
- Rifabutin, 221
- Rifampin, 144, 165, 233, 279
 meningitis, 106
 Mycobacterium avium complex, 240
 Naegleria infection, 42
 Neisseria meningitidis, 88
 tuberculosis, 213, 240
 ventriculitis, 278
- Rifaximin, 86
- Rift Valley fever, 41
 virus, 40, 76
 taxonomy, 340
- Risk factors, 2, 17
- River blindness *See* *Onchocerca volvulus*
- RNA viruses, taxonomy, 339
- Rocky Mountain spotted fever, 231, 301 *See also*
Rickettsia rickettsii
- Rotavirus, 79, 178
 taxonomy, 339
- Roth spots, 102, 297
- Rothia denticariosa*, taxonomy, 345
- Rothia mucilaginosa*, taxonomy, 344
- Roundworms *See* Nematodes
- Route of infection
 ingestion, 193
 maternal, 3, 70, 133, 152, 193
 Neisseria gonorrhoeae, 94
- Route of spread
 airborne, 18, 45
 animal contact, 352
 arthropod, 352
 blood transfusion, 354
 contact, 45
 droplet, 164
 fecal–oral, 14, 32, 77, 174
 cysticercosis, 197
 human contact, 351
 inanimate environment, 353
 ingestion, 123, 352
 maternal, 27, 54, 113, 123, 353
 parenteral, 14
 pharyngeal, 32
 sexual, 351
 transplantation, 354
 water, 353
- Rubella, 157, 192, 293
 congenital, 70, 269
- Rubella virus, taxonomy, 339
- Salicylate, rheumatic fever, 95
- Salmonella, 5, 79, 170
- Salmonella enteritidis*, spinal infection, 274
- Salmonella typhi*, 140, 155, 171, 181
- Salmonella
 intramacrophage survival, 123
 septic arthritis, 153, 267
 taxonomy, 342
- Salpingitis *See* Pelvic inflammatory disease
- Sandfly fever virus, 340
- Sarcoidosis, 233
- Sarcophagidae, taxonomy, 350
- Sarcoptes scabiei*, taxonomy, 350
- Scab, varicella, 45
- Scan
 Gallium, 159
 Technetium, 111, 267, 273
- Scarlet fever, 100, 104, 230
- Schistocyte, 259
 hemolytic uremic syndrome, 80
- Schistosoma haematobium*, 198
 taxonomy, 350
- Schistosoma japonicum*, 198
 taxonomy, 350
- Schistosoma mansoni*, 198
 taxonomy, 350
 transverse myelitis, 263
- Schistosoma mekongi*, 198
 taxonomy, 350
- Schistosomiasis, 139, 155, 198
 chronic salmonella bacteremia, 171
- Schizont, plasmodium, 182
- Screening, Streptococcus agalactiae, 113
- Seizure, 278
 cat-scratch disease, 144
 diarrhea, 80
 differential diagnosis, 194
 intracranial suppuration, 255
 meningitis, 106
 neonate, 119
 theophylline toxicity, 279
- Selenium sulfide shampoo, 303
- Sepsis, 28, 54
- Sepsis syndrome, 40, 157, 315, 331
- Sequestered lobe, 68
- Serology
 amebiasis, 175

[Serology cont.]

- cat-scratch disease, 144
- congenital infections, 72
- cytomegalovirus, 59
- dengue virus, 140
- hepatitis, 15
- human immunodeficiency virus, 62
- Leptospira, 140
- Lyme disease, 143
- schistosomiasis, 201
- Streptococcus pyogenes, 98
- syphilis, 136
- toxoplasmosis, 193
- tularemia, 163
- typhoid fever, 155
- Serositis, 305
- Serratia*, taxonomy, 342
- Sexual activity, 14, 34, 36, 42, 279, 312, 320
- Sexual contacts, 16
- Sexual partner, 94
- Sexually transmitted infections, 321
- Shigella*, 79, 170, 178
 - taxonomy, 342
- Shingles *See* Zoster
- Shock, 36, 39, 104
 - diarrhea, 77
 - hypovolemic, 79
 - infective endocarditis, 293
 - Jarisch-Herxheimer reaction, 139
 - necrotizing enterocolitis, 284
 - nosocomial infection, 165
 - septic, 87, 112, 114, 165, 173, 294, 331
 - suppurative pericarditis, 270
 - viral hemorrhagic fever, 41
- Short-gut syndrome, 277
 - necrotizing enterocolitis, 286
- Shunt infection, 112
- Sickle cell disease, 2, 5, 13, 59, 114, 251, 258
 - parvovirus, 59
 - salmonella osteomyelitis, 80
- Sinusitis, 5, 255, 334
 - maxillary, 271
 - mucormycosis, 243
- Skin infection, mucormycosis, 243
- Skull fracture, basilar, meningitis, 325
- Sleeping sickness *See* *Trypanosoma brucei*
- Smallpox virus, taxonomy, 338
- Snail, 198
- Snuffles, congenital syphilis, 133
- Sore *See* Ulcer
- South Africa, 40, 154, 233, 257, 274, 287, 300

- South America, 41, 77, 198
- Soviet Union, 120
- Spasticity, 65, 68 *See also* Hypertonia
- Speech disturbance, 61
- Spherocytosis, 61
- Spinal cord, 31, 74
- Spinal muscular atrophy, 68, 128
- Spinal shock, 74
- Spirillum minus* *See also* Rat bite fever
 - taxonomy, 341
- Spirochete, 136
 - relapsing fever, 139
- Spirometra mansonoides*
 - taxonomy, 349
- Spleen, 114
- Spleen rupture, infectious mononucleosis, 36, 259
- Splenectomy, 114
- Splenic infarct, 115
- Splenomegaly, 34, 58, 257, 304
 - autoimmune lymphoproliferative syndrome, 310
 - cat-scratch disease, 143
 - congenital infection, 69
 - congenital syphilis, 133
 - human immunodeficiency virus infection, 65
 - infectious mononucleosis, 36
 - infective endocarditis, 297
 - malaria, 184
 - neonate, 268
 - relapsing fever, 137
 - thalassemia, 173
 - toxoplasmosis, 193
 - tuberculosis, 233
 - typhoid fever, 154
 - visceral larva migrans, 202
- Spondylitis, tuberculous *See* Tuberculosis: spine
- Sporothrix schenckii*, taxonomy, 347
- Stain
 - acid-fast, 6, 84, 127, 180
 - fluorochrome, 6
 - Giemsa, 139, 186, 190
 - Gram *See* Gram stain
 - hematoxylin and eosin, 6
 - immunofluorescent
 - measles, 22
 - methylene blue, 170, 303
 - Papanicolaou, 6
 - Romanowsky, 6
 - silver, 6
 - Wright's, 114
- Staphylococcus aureus*, 5, 20, 48, 55, 87, 104, 125
 - bacterial tracheitis, 319

- brain abscess, 327
- cellulitis, 275
- cystic fibrosis, 168
- endocarditis, 103
- food poisoning, 78
- impetigo, 27
- infective endocarditis, 295
- intracranial suppuration, 256
- lobar pneumonia, 247
- lung abscess, 249
- lymphadenitis, 163
- methicillin-resistant, 50, 88, 104, 111, 267, 307
 - neck abscess, 261
- neck abscess, 261
- neonatal conjunctivitis, 92
- nosocomial infection, 166
- osteomyelitis, 281
- pelvic abscess, 307
- pleural effusion, 232
- pleural empyema, 117
- pneumonia, 250
- retropharyngeal abscess, 264
- septic arthritis, 266, 293
- skin ulcer, 130, 190
- spinal abscess, 262
- spinal infection, 274
- suppurative pericarditis, 271
- taxonomy, 343
- ventriculitis, 112
- Staphylococcus epidermidis*, taxonomy, 344
- Staphylococcus haemolyticus*, taxonomy, 344
- Staphylococcus*, 102, 166, 240
 - coagulase-negative
 - nosocomial infection, 166
 - ventriculitis, 112, 278
 - methicillin-resistant
 - nosocomial infection, 166
- Starvation, 21
- Stenotrophomonas maltophilia*
 - cystic fibrosis, 168
 - pneumonia, 250
 - taxonomy, 342
- Stevens–Johnson syndrome, 17, 231
- Strabismus, 191
- Streptobacillus moniliformis* *See* Rat bite fever
- Streptobacillus moniliformis*, taxonomy, 343
- Streptococci
 - brain abscess, 327
 - dental infection, 272
 - peritonitis, 315
 - retropharyngeal abscess, 264
 - viridans group, 344
 - infective endocarditis, 295
- Streptococcus agalactiae*, 268
 - neonatal pneumonia, 250
 - neonatal sepsis, 113
 - osteomyelitis, 110
 - taxonomy, 344
- Streptococcus bovis*, taxonomy, 344
- Streptococcus* group B *See* *Streptococcus agalactiae*
- Streptococcus milleri*, intracranial suppuration, 255
- Streptococcus pneumoniae*, 2, 5, 9, 20, 55, 63, 66, 106, 125
 - brain abscess, 327
 - epiglottitis, 318
 - human immunodeficiency virus infection, 65
 - intracranial suppuration, 255
 - lobar pneumonia, 247
 - meningitis, 274
 - nephrotic syndrome, 315
 - penicillin-resistant, 106, 275
 - peritonitis, 315
 - pleural effusion, 232
 - pneumonia, 250, 325, 329
 - septic arthritis, 267
 - sickle cell disease, 114
 - suppurative pericarditis, 271
 - taxonomy, 344
- Streptococcus pyogenes*, 5, 34, 48, 95, 100, 104
 - cellulitis, 275
 - epiglottitis, 318
 - glomerulonephritis, 98
 - lymphadenitis, 163
 - neck abscess, 261
 - septic arthritis, 267, 293
 - taxonomy, 344
 - tonsillitis, 120, 310
- Streptomyces somaliensis*, taxonomy, 345
- Streptomycin
 - tuberculosis, 213
 - tularemia, 163
- Stridor
 - differential diagnosis, 318
 - diphtheria, 120
 - epiglottitis, 264
 - laryngotracheobronchitis, 316
- Stroke, 62
 - infective endocarditis, 295
- Strongyloides stercoralis*, 55
 - taxonomy, 349
- Subacute sclerosing panencephalitis, 21
- Subarachnoid space, 215

- Subdural effusion, meningitis, 108
- Sublingual space, 272
- Submandibular salivary gland, 237, 261
- Submandibular space, 272
- Submasseteric space, 272
- Sulfadiazine, toxoplasmosis, 194
- Sulfadoxine, 188
 - malaria, 182
- Sulfonamide, toxoplasmosis, 42
- Sulfur granule, 125
- Supportive care, 2, 5, 51, 73, 75, 106
 - botulism, 129
 - bronchiolitis, 26
 - diphtheria, 121
 - intracranial suppuration, 255
 - malaria, 188
 - pneumonia, 247
 - septic shock, 87, 166
 - tuberculous meningitis, 210
 - varicella pneumonia, 47
- Support, respiratory, 32
- Surgery, 7
 - infective endocarditis, 299
- Surgical debridement, mucormycosis, 243
- Surgical drainage, 48, 104
 - actinomycosis, 125
 - amebic liver abscess, 175
 - dental abscess, 271
 - intracranial suppuration, 256
 - neck abscess, 261
 - pelvic abscess, 307
 - pleural empyema, 252
 - retropharyngeal abscess, 264
 - suppurative thrombophlebitis, 167, 240
- Surgical removal, hydatid cyst, 290
- Sutton's Law, 4
- Syphilis, 150, 157, 293, 311, 321
 - chronic meningitis, 210
 - congenital, 70, 74, 133, 152, 269
 - secondary, 133
- Systemic lupus erythematosus, 42, 95, 233, 251, 277, 293, 312
 - chronic meningitis, 210
- Tachypnea, 55
 - bronchiolitis, 24
- Taenia saginata*, taxonomy, 349
- Taenia solium*, 195
 - taxonomy, 349
- Tanapoxvirus, taxonomy, 339
- Tapeworm, 290
- Tapeworms *See* Cestodes
- Temporal space, 272
- Tetanus, 119
- Tetracycline
 - Balantidium coli*, 180
 - pneumonia, 330
 - relapsing fever, 139
- Thalassemia major, 172
- Theophylline, 279
- Thiabendazole, cutaneous larva migrans, 205
- Thoracoscopy, pleural empyema, 253
- Throat culture, 35
- Throat, pain, 120
- Thrombocytopenia, 102
 - congenital syphilis, 133
 - congenital rubella, 71
 - hemolytic uremic syndrome, 80
 - human immunodeficiency virus infection, 65
 - infectious mononucleosis, 36
 - malaria, 184
 - yellow fever, 75
- Thrombophlebitis
 - Lemierre syndrome, 332
 - suppurative, 167, 240
- Tick, 40, 42, 148
 - arbovirus, 76
 - Lyme disease, 141
 - relapsing fever, 138
 - tularemia, 163
- Tinidazole
 - amebiasis, 86, 177
 - giardiasis, 180
- Tissue and blood protozoa, taxonomy, 348
- Tissue culture, 6
- T-lymphocyte, 35
- Tobramycin
 - cytic fibrosis, 168
 - Pseudomonas aeruginosa*, 295
- Togaviridae, 76
 - taxonomy, 339
- Tonsillitis, 34, 95, 98, 99, 120
 - adenovirus, 310
 - Lemierre syndrome, 331
- Total parenteral nutrition, *Malassezia furfur*, 303
- Toxic megacolon, amebic, 175
- Toxic shock syndrome, 104, 230
 - staphylococcal, 48
 - streptococcal, 48
- Toxic shock-like syndrome, 104
- Toxin, 13, 15
- Toxocara canis*, 202, 251
- Toxocara cati*, 202, 251
- Toxocara*, taxonomy, 349

- Toxoplasma gondii*, 5, 36, 42, 55, 58, 59, 137, 192, 312
 blood transfusion, 173
 congenital infection, 70
 taxonomy, 348
 Toxoplasmosis, congenital, 269
 Tracheitis, bacterial, 318
 Tracheostomy, diphtheria, 122
 Transaminase, 13, 181
 cytomegalovirus, 56
 drug toxicity, 279
 Lemierre syndrome, 331
 yellow fever, 75
 Transplantation, 5, 37, 55, 123
 adenovirus, 311
 bone marrow, 125
 liver, 14
 mucormycosis, 243
 pneumonia, 250
 Trauma, 110
 Travel, 3, 17
 Trematodes, taxonomy, 349
Treponema pallidum, 42, 59, *See also* Syphilis
 blood transfusion, 173
 taxonomy, 341
Trichinella spiralis, taxonomy, 350
Trichomonas vaginalis, 6, 321
 taxonomy, 348
Trichophyton, taxonomy, 347
Trichuris trichiura, taxonomy, 349
 Trimethoprim/sulfamethoxazole
 cat-scratch disease, 144
 Cyclospora, 180
 gastroenteritis, 86
 neck abscess, 261
 Nocardia, 127
 Pneumocystis jiroveci, 65
 salmonella, 171
 typhoid fever, 156
 urinary tract infection, 245
Tropheryma whipplei, taxonomy, 345
 Trophozoite, plasmodium, 182
Trypanosoma, 6
Trypanosoma brucei, 59, 76, 190, 300
 blood transfusion, 173
Trypanosoma brucei gambiense, taxonomy, 348
Trypanosoma brucei rhodesiense, taxonomy, 348
Trypanosoma cruzi, 59, 76
 blood transfusion, 173
 taxonomy, 348
 Trypanosomiasis, 131
 African *See* *Trypanosoma brucei*
 American *See* *Trypanosoma cruzi*
 Tsetse fly *See* Glossina
 Tubercle, 219, 237
 Tuberculin skin testing, 223, 233
 Tuberculoma, seizure, 195
 Tuberculosis, 20, 66, 154, 210, 220, 223, 279
 Tuberculosis
 meningeal *See also* Meningitis: tuberculous
 miliary, 210, 215, 233
 osteomyelitis, 281
 pleural effusion, 232
 retropharyngeal abscess, 264
 spinal, 226, 263
 Tubular necrosis, rhabdomyolysis, 75
 Tularemia, 144, *See also* Francisella tularensis
 Tumor, 263, 266
 abdominal, 287
 spinal, 263
 Tumor necrosis factor receptor, 305
 Typhoid fever, 171, 233, 259, *See also* Salmonella
 typhi
 abdominal pain, 253
 Typhus
 endemic *See* Rickettsia mooseri
 epidemic, 150
 See also Rickettsia prowazekii
 Tyrosinemia, 157
 Ukraine, 120
 Ulcer, 242
 colonic, 174
 corneal, 27
 decubitus, 73
 dendritic, 27, 152
 genital, 27
 acute HIV infection, 312
 intestinal
 typhoid fever, 155
 mouth
 PFAPA syndrome, 304
 nasal
 syphilis, 133
 peptic, 13
 skin
 anthrax, 130
 Ultrasound
 abdominal, 13, 144, 174
 lung, 252
 pelvic inflammatory disease, 323
 renal, 245
Ureaplasma urealyticum, 250
 taxonomy, 341
 Urinalysis, 13
 myositis, 75

- Urinary tract infection, 13, 157, 244, 320
Urosepsis *See* Urinary tract infection
Uveitis, 152
 Kawasaki disease, 231
 Neisseria meningitidis, 92
 zoster, 53
- Vaccination, 1, 32, 117, 120, 165
 hepatitis B, 16
 Neisseria meningitidis, 88
 pertussis, 162
 Streptococcus pneumoniae, 316
- Vaccine, 165
 hepatitis A, 15
 measles, 23
 Neisseria meningitidis, 88
 pertussis, 162
 polio, 33
 Streptococcus pneumoniae, 117, 316
 yellow fever, 77
- Vaccinia, 46
Vaccinia virus, taxonomy, 338
Valacyclovir, zoster, 53
Vancomycin, 51, 104, 114, 166, 172, 173
 brain abscess, 328
 Clostridium difficile, 86
 endocarditis, 102
 endophthalmitis, 314
 epidural abscess, 334
 infective endocarditis, 295, 299
 intracranial suppuration, 256
 meningitis, 106, 212
 osteomyelitis, 111
 skeletal infection, 267
 vascular catheter infection, 277
 ventriculitis, 278
- Varicella, 5, 27, 45, 45, 192
Varicella zoster virus, 45, 66
 taxonomy, 338
- Vascular disease, 192
Vascular injury, meningitis, 106
Vascular occlusion, 89, 172
 dehydration, 79
 Neisseria meningitidis, 89
 seizure, 195
- Vasculitis, 251
 spinal cord, 263
- Vasculopathy
 hemolytic uremic syndrome, 80
 human immunodeficiency virus infection, 62, 65
 malaria, 183
- Vector, 163
Vegetation, 103
 infective endocarditis, 296
Veillonella, taxonomy, 346
Ventilation, tetanus, 120
Ventilator, nosocomial infection, 165
Ventriculitis, 112, 278
Ventriculoperitoneal shunt, 112, 278
Verruga peruana, 146
Vesicle, 27, 45, 70
Vesicoureteral reflux, 245
Vibrio cholerae, 79
 taxonomy, 343
Vibrio parahaemolyticus, 79
 taxonomy, 343
Vibrio vulnificus, taxonomy, 343
Viral capsid antigen, 39
Viral hemorrhagic fever, 40, 181
Viral load, 57
Visceral larva migrans, 202, 251
Vitamin A, 22
Vitrectomy, 314
Voiding cystourethrogram, 245
Volvulus, midgut, 253
Vomiting, 279
 appendicitis, 253
 theophylline toxicity, 279
 ventriculitis, 278
- Waterhouse-Friderichsen syndrome, 92
Weakness, 30, 72, 323
 acute HIV infection, 311
 flaccid
 poliomyelitis, 31
 transverse myelitis, 262
 tuberculosis, 233
Wegener's granulomatosis, 251, 324
Weight loss, 203
 osteomyelitis, 281
 tuberculosis, 220, 233
West Africa, 41, 206
West Nile virus, 32, 59, 74, 150
 blood transfusion, 173
 taxonomy, 340
Wheeze, bronchiolitis, 24
Whooping cough *See* Pertussis
Widal test *See* Serology: typhoid fever
Wilson's disease, 15
Wimberger's sign, 136
Wolbachia, 206
Worm burden, 208
Wound, surgical, 58
Wucheraria bancrofti, taxonomy, 350

- X-linked lymphoproliferative disorder, 37
- Yeast, 166, 241
 - endocarditis, 103
 - pityriasis versicolor *See* *Malassezia furfur*
 - taxonomy, 346
- Yellow fever virus, 40, 41, 76, 181
 - taxonomy, 340
- Yersinia enterocolitica*, 6, 59, 79, 170, 173
 - blood transfusion, 173
 - taxonomy, 342
- Yersinia pestis*, 76, 163
 - taxonomy, 342
- Zebra, 3
- Zenker's degeneration, 155
- Zoster, 52
- Zygomycete, 242
 - taxonomy, 347